

# Modelling the effects of past and future climate on the risk of bluetongue emergence in Europe

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Vector-borne diseases are among those most sensitive to climate because the ecology of vectors and the development rate of pathogens within them are highly dependent on environmental conditions. Bluetongue (BT), a recently emerged arboviral disease of ruminants in Europe, is often cited as an illustration of climate's impact on disease emergence, although no study has yet tested this association. Here, we develop a framework to quantitatively evaluate the effects of climate on BT's emergence in Europe by integrating high-resolution climate observations and model simulations within a mechanistic model of BT transmission risk. We demonstrate that a climate-driven model explains, in both space and time, many aspects of BT's recent emergence and spread, including the 2006 BT outbreak in northwest Europe which occurred in the year of highest projected risk since at least 1960. Furthermore, the model provides mechanistic insight into BT's emergence, suggesting that the drivers of emergence across Europe differ between the South and the North. Driven by simulated future climate from an ensemble of 11 regional climate models, the model projects increase in the future risk of BT emergence across most of Europe with uncertainty in rate but not in trend. The framework described here is adaptable and applicable to other diseases, where the link between climate and disease transmission risk can be quantified, permitting the evaluation of scale and uncertainty in climate change's impact on the future of such diseases.

Keywords: climate change; vector-borne disease transmission; basic reproductive ratio; emergence; bluetongue; *Culicoides* 

## 1. INTRODUCTION

Climate change may cause vector-borne diseases to shift in distribution because the vectors' ecology and the pathogen development rate within them strongly depend on environmental conditions. In some cases, shifts to previously unexposed populations of humans and animals could have severe or even devastating consequences. Modelled projections of how vector-borne diseases will respond to climate change are needed so that measures of mitigation or adaptation can be taken.

Bluetongue (BT), a viral disease of ruminants transmitted by biting midges (*Culicoides* spp.), is considered by many to represent one of the most plausible examples of climate change driving the emergence of a

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vector-borne disease [1]. BT is widely distributed in Africa, Asia, Australia, South America and North America. In Europe, although a few sporadic outbreaks occurred in the last century, BT had never established itself in the long term. A dramatic change occurred in 1998 when an unprecedented series of outbreaks began [2], involving nine different serotypes of the virus, causing the deaths of millions of ruminants, and major economic consequences for the region. In Europe, over 110 000 outbreaks were declared to the World Animal Health Organization (OIE) between 1998 and 2010; over 80 000 of these were owing to BT virus serotype 8. The emergence of the disease in southern Europe has been attributed, in part, to the northwards spread of the afro-tropical vector, Culicoides imicola, across the Mediterranean basin [3]. This species is currently absent from northern Europe. However, BT occurred for the first time in northwest (NW) Europe in 2006, transmitted by indigenous species of *Culicoides*, most importantly members of the Obsoletus complex [4,5].

Possible causes of BT's emergence in Europe have been discussed by Purse *et al.* [3] (see the electronic supplementary material for detailed discussion on possible non-climatic drivers). They conclude that it seems improbable that biotic or non-climatic abiotic factors could have been responsible for this emergence. On the contrary, evidence is presented that the emergence of BT in southern Europe occurred in the same place and at the same time as regional warming between the 1980s and 1990s, thereby providing support for a role of climate change [3]. However, this correlation does not quantify precisely the link between climate and disease transmission parameters to explain the observed emergence. Thus, although it suggests possible mechanisms by which warming could have affected the disease, it does not identify which mechanisms are the most important. Additionally, it does not account for the emergence of BT in northern Europe in 2006. Nevertheless, this link with climate change, and the recent spread of disease in Europe, makes BT an excellent example for developing models of how climate change will influence diseases in the future, with the opportunity to validate models against the observed past outbreak occurrence.

In this paper, we present a framework for the evaluation of the effects of past and future climate on the risk of emergence of BT. The approach is based on an epidemiological model of disease transmission, the basic reproduction ratio,  $R_0$ , adapted for a two-host and two-vector disease. Four parameters of the model are known to be climate-driven. For three parameters, the equations linking the parameter to temperature have been described in the literature. To estimate the link of the fourth parameter with climate, new vector abundance models were developed. By integrating the observed high spatial resolution (25 km) climate data in the disease transmission model, the disease transmission risk can be evaluated for past periods of time. This allows us to evaluate whether the integrated climate-driven model successfully reproduces aspects of BT's past observed occurrence, including the distribution of its vectors and the emergence of the disease in NW Europe in 2006. Examining whether past epidemiological events have been driven by climate is essential before projecting models in the future [6]. In order to drive the model into the future, simulations of future climate are then integrated into the  $R_0$ model. Two steps are necessary for this. First, the model projections for past climate, obtained by integrating an ensemble of 11 regional climate models (RCMs), are compared with the  $R_0$  model driven by the observed climate data to make sure that the ensemble of climate models is able to reproduce past patterns of BT. In a second step, the  $R_0$  model is then driven by the simulations of the ensemble of 11 RCMs under a climate change scenario to evaluate the transmission risk for a future time period up to 2050. Using an ensemble of models instead of a single model, it is important to take into account the uncertainties related to the climate simulations which affect the transmission simulations.

#### 2. MATERIAL AND METHODS

## 2.1. Evaluating the basic reproduction number $R_0$ for bluetongue

The basic reproduction ratio ( $R_0$ , the number of secondary cases arising from the introduction of one infected host in a susceptible population) is a powerful tool to assess the risk of disease transmission in the event of viral introduction [7] (at the onset of the epidemic). Gubbins *et al.* [8] and Hartemink *et al.* [9] have modelled BT's  $R_0$  as a two-host disease, as cattle and sheep play different epidemiological roles [8,9]. Indeed, cattle, as opposed to sheep, are usually not clinically affected [10,11] and harbour a longer viraemia [12].  $R_0$  for BT was adapted from Gubbins *et al.* [8]

$$R_0 \propto \left(\frac{b\beta a^2}{\mu}\right) \left(\frac{\nu}{(\mu+\nu)}\right) \left(\frac{m\varphi^2}{(r_{\rm c}+d_{\rm c})} + \frac{m(1-\varphi)^2}{(r_{\rm s}+d_{\rm s})}\right),$$

where b is the probability of transmission from vector to host,  $\beta$  the probability of transmission from host to vector, a the biting rate,  $\mu$  the vector mortality rate,  $1/\nu$  the mean extrinsic incubation period, m the vector-to-host ratio,  $\varphi$  the proportion of bites on cattle, 1/r the duration of viraemia (in cattle  $r_c$  and in sheep  $r_s$ ) and 1/d the disease-induced mortality rate (in cattle  $d_c$  and in sheep  $d_s$ ).

The formula developed by Gubbins *et al.* [8] is very similar to the one derived separately by Hartemink *et al.* [9] and used to model  $R_0$  for BT across The Netherlands. Gubbins *et al.* [8] distinguished a vector-to-cattle ratio from a vector-to-sheep ratio in their formulae. Here, we propose a method that inherently estimates these two parameters together. However, we considered that this ratio had to be computed distinctively for *C. imicola* and for the Obsoletus complex because of the dissimilar distributions of these species (see the estimation of *m*).

We chose to only consider sheep and cattle as BT hosts. We did not consider other domestic hosts such as goats because of their small population sizes in Europe. We did not consider wild hosts because very little information is available to include them as a third host (no parameter estimates are available for wildlife) and because their role seems to be more important in the persistence of the disease than at the onset of the disease (see electronic supplementary material).

Although transplacental transmission in hosts has been shown to occur in cattle with BT serotype 8 virus [13], and probably represents an important overwintering mechanism, it has been considered to be insignificant at the onset of the epidemic [9].

Four parameters were considered to be constant in space and time: the probabilities of transmission from vector to host (b) and from host to vector ( $\beta$ ), the duration of viraemia (1/r) and the disease-induced mortality rates (1/d) (estimates of all parameters are given in table 1). As host preferences of the two European vectors are not well described, in our model, the proportion of bites on cattle,  $\varphi$ , and on sheep,  $(1-\varphi)$ , is defined as the proportion of cattle and sheep available, respectively. The availability of cattle

| parameter                    | definition                                                 | estimation or range (value chosen)                                             | reference |
|------------------------------|------------------------------------------------------------|--------------------------------------------------------------------------------|-----------|
| b                            | probability of transmission from<br>vector to host         | $0.8 - 1.0 \ (0.9)$                                                            | [14]      |
| β                            | probability of transmission from<br>host to vector         | $0.001 {-} 0.15 \ (0.075)$                                                     | [15 - 17] |
| a                            | biting rate $(d^{-1})$                                     | $a(T) = 0.000171 \times T \times (T - 3.6966) \times (41.8699 - T)^{1/2.7056}$ | [18, 19]  |
| $\mu$                        | vector mortality rate; $(d^{-1})$                          | $\mu(T) = 0.008941 \exp(0.1547 \times T)$                                      | [20]      |
| $\frac{1}{\nu}$              | mean extrinsic incubation period (day)                     | $\nu(T) = 0.0003 \times T \times (T - 10.4057)$                                | [18]      |
| m                            | vector-to-host ratio                                       | see main text                                                                  |           |
| $\varphi$                    | proportion of bites on cattle                              | number of cattle/number of sheep and cattle                                    |           |
| $1/r (r_{\rm c}, r_{\rm s})$ | duration of viraemia (cattle,<br>sheep) (day)              | cattle: 20.6; sheep: 16.4                                                      | [21 - 23] |
| $1/d (d_{\rm c}, d_{\rm s})$ | disease-induced mortality rate (cattle, sheep); $d^{-1}$ ) | cattle: 0 sheep: $0.001 - 0.01 (0.005)$                                        | [11, 24]  |

Table 1.  $R_0$  parameters for the two-host bluetongue transmission model. T, temperature.

and sheep was based on gridded estimates of livestock density in 2005 [25].

Four parameters—*a*, the vector biting rate (the daily probability of a midge feeding on a susceptible host);  $\mu$ , the vector mortality rate;  $\nu$ , the reciprocal of viral extrinsic incubation period (the time taken for a midge to become infectious after taking an infected blood meal); and *m*, the vector-to-host ratio—are known to exhibit strong climatic dependence [8,26] and therefore vary in space and time (table 1). Equations linking the first three parameters to temperature for North American midges were obtained from published literature (table 1 and electronic supplementary material for more details on how the parameters were estimated). The relationships with temperature are shown in electronic supplementary material, figure S1.

Estimation of the vector-to-host ratio (m) is complex. Indeed, Culicoides are usually sampled in the field using light traps: however, there is no established method of converting trap catch data to population size. Here, we assume that a trap acts like a host and attracts a number of midges proportional to what a host would attract. Under this assumption, a catch does not directly estimate the vector population size but, instead, is an estimate of the vector-to-host ratio. In other words, the number of vectors caught in a trap depends on both the population size of the vectors, and the number of hosts (and traps) to which they are attracted. This assumption seems more realistic as  $R_0$  is then proportional to trap catches, whereas under the assumption that trap catches reflect a given percentage of the vector population size,  $R_0$  may be very high in areas where there are a few vectors but very low host densities (such as cities for example).

Furthermore, the vector-to-host ratio has to be estimated separately for the two vectors, C. imicola and the Obsoletus complex, as they have dissimilar ecologies and distributions [27,28]. Statistical models of trap catches for C. imicola and Obsoletus complex have been published in recent years [26,29–32], but all include variables that cannot be projected under future climate change scenarios, such as the normalized difference vegetation index. We therefore developed new vector distribution models restricting explanatory variables to ones which could be projected in the future.

Using data provided by the Spanish BT national surveillance programme on *Culicoides* trap catches set in livestock holdings from 2004 to 2006, presence-absence and abundance models were developed following the methods of Calvete et al. [26]. At each farm, the maximum catch per year was considered to be the best estimate of midge abundance. Indeed, trap catches can be extremely variable throughout the year and even between two consecutive nights, and highly dependent on local meteorological conditions [33]. Maximum catches are classically considered to be the best approximation of the midge abundance as any smaller catches could be underestimations owing to meteorological conditions being temporarily not favourable [33]. The dataset was divided into a training (n = 330) and an independent validation (n = 255) dataset. Using an information-theoretic paradigm based on Akaike's criterion, the best logistic regression models of the probability of occurrence of each vector were selected (electronic supplementary material, table S1). Five bioclimatic variables were included in the models: mean annual temperature, annual precipitation, their variation coefficients (all extracted from a 1950-2000 monthly climatic series) and a sun index, derived from a digital elevation model. Both occurrence models have a fairly good discriminatory capacity in internal (the area under the receiver-operating characteristic curve is 0.811 for C. imicola and 0.736 for the Obsolution complex) and external (0.779 for C. imicola)and 0.710 for the Obsoletus complex) validation. Midge abundance was obtained by fitting a regression equation to the predicted probability of detection (electronic supplementary material, table S2). The vector-to-host ratio was obtained by calibrating the abundance of each vector on a 0-5000 scale [8] (figure 1b,e with areas of maximal abundance (obtained for a probability of occurrence equal to 1) having a ratio of 5000. The ratios calculated for the two vectors were then summed to compute m for all vectors.

Contrary to others [9], we considered that  $R_0$  for BT could not be quantified exactly because of the lack of knowledge on specific estimates of some of the  $R_0$  parameters for the European species of vectors (such as  $a, \mu$ and  $\nu$ ). Therefore, we conservatively present  $R_0$  anomalies, the change in  $R_0$  in one time period relative to a baseline.

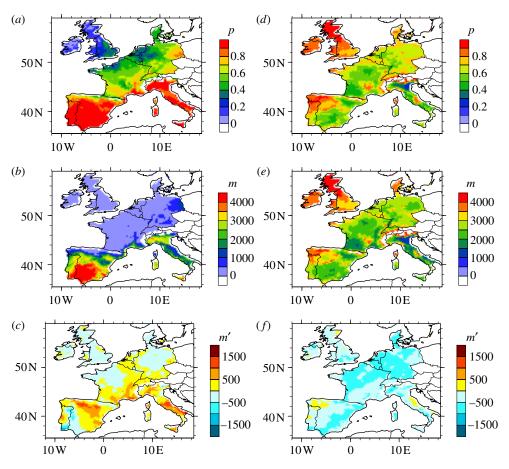


Figure 1. Mean probability of presence, vector-to-host ratio, and anomalies for *C. imicola* and the Obsoletus complex. (a) Mean probability of presence for *C. imicola* (August–September–October (ASO) 1961–1999). (b) Vector-to-host ratio for *C. imicola*. (c) Vector-to-host ratio anomaly for *C. imicola* for 2000–2008 (relative to the 1961–1999 mean). (d) Mean probability of presence for the Obsoletus complex (ASO 1961–1999). (e) Vector-to-host ratio for the Obsoletus complex. (f) Vector-to-host ratio anomaly for the Obsoletus complex for 2000–2008 (relative to the 1961–1999 mean).

#### 2.2. Climate data

Temporal variation in  $R_0$  was derived from three sources of climate data: a high-resolution, observed climate dataset for 1961–2008; and two ensembles of climate model simulations provided by the ENSEM-BLES research theme 3 (available at http:// ensemblesrt3.dmi.dk/). Observed temperature and rainfall are estimated from the E-OBS gridded dataset (25 km resolution) which is derived through the interpolation of station measurements [34].

Regional scenarios for climate change impacts assessment require finer spatial scales than those provided by general circulation models (GCMs), which have a coarse resolution (about 300 km). The ENSEMBLES European project (http://ensembles-eu.metoffice.com/) provides improved RCMs, at spatial scales of 25 km, for both recent past (1961–2000) and future climate scenarios (1950–2050). Models covering the European domain with a regular  $0.25^{\circ}$  step consistent with the observation grid were retained. Two ensembles of simulations are considered, the control experiments (SimCTL) and the scenario experiment (SimA1B).

In the SimCTL experiment (1961–2000), all RCMs are forced at their boundaries by the ERA40 reanalysis (the 'best guess' of the observations that uses both modelling and different sources of observations through

data assimilation) [35]. Observed external forcing (greenhouse gases, solar, volcanic, aerosols) is applied to all RCMs.

In the SimA1B experiment (1961–2050), the RCMs are forced at their boundaries by a GCM with a coarser resolution (about 300 km) forced by the A1B emission scenario (median scenario in terms of CO<sub>2</sub> emissions) [36]. Different GCMs are used to drive the RCMs. This explains, in part, the large spread in the different scenarios with respect to the control run (in which all RCMs have the same boundary conditions, namely they are all driven by the ERA40 reanalysis). As each RCM realization is based on a different model (with different physical parametrization), and as the GCM which provides the RCM boundary conditions vary from one RCM to another, we can assume that the various RCM projections are independent of one another.

The 11 selected RCMs (and operational centre which developed them) are: C4IRCA3 (Met Éireann, Ireland), CNRM-RM4.5 (CNRM, Météo-France), DMI-HIRAM5 (DMI, Denmark), ETHZ-CLM (ETHZ, Switzerland), ICTP-RegCM3 (ICTP, Italy), KNMI-RACMO2 (KNMI, Netherlands), METO-HC (Met Office, UK), MPI-M-REMO (MPI, Germany), OURANOSMRCC (OURANOS, Canada), SMHIRCA (SMHI, Sweden) and UCLM-PROMES (UCLM, Spain).

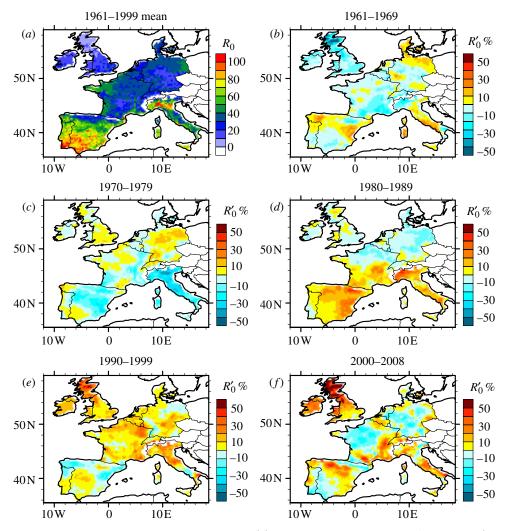


Figure 2. Long-term mean and modelled  $R_0$  decadal variability. (a) Long-term mean  $R_0$  for the ASO season (the average is computed for the 1961–1999 period). The magnitude has been scaled to vary between an arbitrary range between 0 and 100%. (b-f)  $R_0$  relative anomalies (%) with respect to the reference mean (1961–1999) for different decades.  $R_0$  is estimated from the observed climate dataset.

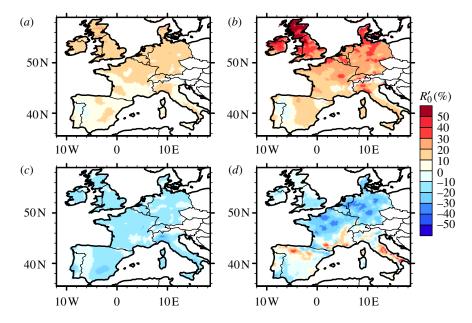


Figure 3. Sensitivity of  $R_0$  parameters to climate change.  $R_0$  relative anomalies (%) for the period 2000–2008 with respect to the 1961–1999 climatology (ASO) based on the climate observations. All parameters are assumed to be constant except one in each panel: in (a) the biting rate; in (b) 1/mean extrinsic incubation period; in (c) the vector mortality rate; and in (d) the vector-to-host ratio (m).

RCM-simulated precipitation and temperature were bias-corrected on a monthly mean basis with respect to the E-OBS observed dataset over the 1961–1999 reference period (see the electronic supplementary material).

# 2.3. Integration of climate data in the transmission model

We acknowledge that the entomological data used to build the models covered only a restricted geographical area (Iberia). Thus, model projections were limited to Western Europe and results of midge distribution were carefully examined (see §3) before computing  $R_0$ anomaly maps. For some outputs, a distinction was made between southwest (SW) Europe (below the northern border of Spain, i.e. the area where *C. imicola* is abundant) and NW Europe (above Switzerland, where *C. imicola* is absent).

In Europe, BT is highly seasonal and occurs mostly in late summer to autumn. For each climate dataset, the temperature-dependent parameters  $(a, \mu, \nu)$  included in the  $R_0$  model are computed for August, September and October (ASO) and then averaged to build a seasonal mean for each year. The vector-to-host ratio (m) is computed on an annual basis, and then integrated with the seasonal mean of the other parameters to compute  $R_0$ for ASO. For each of the two ensembles (SimCTL and SimA1B), the ensemble mean is then estimated by averaging the  $R_0$  values for each individual RCM simulation. These estimates were averaged to give long-term and decadal means. To evaluate the specific effect of each parameter on  $R_0$  anomalies (figure 3), all parameters but one were held constant. To investigate the relative influences of change in temperature versus rainfall from the SimA1B ensemble on future change in  $R_0$  anomaly, each was held constant (temperatures at 20°C and 25°C, rainfall at 250 and 500 mm) in turn, while the other was allowed to vary (figure 6). Changes in temperature and rainfall over the 1960–2050 period are shown in the electronic supplementary material, figure S3. For a given ensemble, the spread of simulated  $R_0$  (figure 7) is defined as 1 s.d. of all RCM projections with respect to the ensemble mean. The multi-model sign consistency is computed assigning +1(-1) to each RCM projection if an increase (decrease) in  $R_0$  is simulated. This is averaged and multiplied by 100 to display percentages.

### 3. RESULTS

#### 3.1. Modelled vector-to-host ratios

Modelled vector-to-host ratios for *C. imicola* for the 1961-1999 period (figure 1*b*), driven by the observed climate data, reproduce the past situation in Spain and southern Portugal, the only parts of SW Europe in which this species was known to occur before 1998. The anomaly for the period 2000-2008 (i.e. the change in modelled ratio during this time period compared with that of 1961-1999) reproduces remarkably well the recent spread of *C. imicola* to areas of northern Spain and southeast France (figure 1*c*). The precise time of introduction is not known, but *C. imicola* was

detected for the first time in 2000 in Corsica [37], in southeast France in 2003 [38] and in northern Spain in 2007 [39]. The species is known to be spreading in these areas. However, the model does not reproduce its first detection in Catalonia in 2002 [40]. In Italy, the situation is not as clear: *C. imicola* was first detected in 2000 [41] but entomological surveillance was unable to detect a range of expansion between 2002 and 2007 [42]. The model over-predicts the presence of *C. imicola* in north Italy, where it has not yet been detected.

Regional data on the distribution of the indigenous Obsoletus complex do not yet exist. Nevertheless, modelled vector-to-host ratios for the Obsoletus complex for the 1961–1999 period confirm its known, very widespread distribution (figure 1e). Negative anomalies of the Obsoletus complex for 2000–2008 occur across almost the entire region suggesting that, recently, climate has caused the density of this complex to decrease (figure 1f).

### 3.2. Past $R_0$ anomalies

The simulated mean  $R_0$  (figure 2a), again based on observed climate data, depicts an increasing North-South gradient and correctly identifies southern Spain and Portugal as key areas at risk of BT for the 1961–1999 period. Although no BT occurred in north Italy during that period (see OIE Handistatus II Annual animal disease status Europe/2002/Bluetongue Animal health status at http://www.oie.int/hs2/sit\_mald\_cont. asp?c\_mald=10&c\_cont=4&annee=2002), the model projects high risk of disease transmission in the event of viral introduction. In the 1960s and 1970s (figure 2b,c), most areas had negative  $R_0$  anomalies and hence low risk of disease transmission (in the event of viral introduction) relative to the 1961-1999 mean. In the 1980s (figure 2d), areas of Spain, southern France and NW Italy displayed positive  $R_0$  anomalies, suggesting an increase in the risk of disease transmission in the event of viral introduction. In the 1990s and early 2000s, strong positive  $R_0$  anomalies in NW Europe including the UK are highlighted (figure  $2e_{f}$ ). The climate conditions over NW Europe could thus have been favourable to BT transmission for 15 years before the virus was introduced in 2006.

In NW Europe, these positive  $R_0$  anomalies are linked to changes in the biting rate (a) (figure 3a), and particularly the extrinsic incubation period ( $\nu$ ) (figure 3b). By contrast, changes in the vector mortality rate and *Culicoides* density do not explain this increase (figure 3c,d). In SW Europe, the influence of changing biting rate and extrinsic incubation period on the  $R_0$ anomaly is reduced compared with the NW, but there is a substantial contribution from changes in the vector-to-host ratio in parts of Spain, France and Italy, related to spread in the distribution of *C. imicola* (figure 3d).

The first ever occurrence of BT in NW Europe [43] occurred in the year for which the model has the largest positive anomaly since 1961 (figure 4a, histogram). The origin of the viral introduction in 2006 is still not known; however, our model shows that climate rendered that year in northern Europe at higher risk of a BT

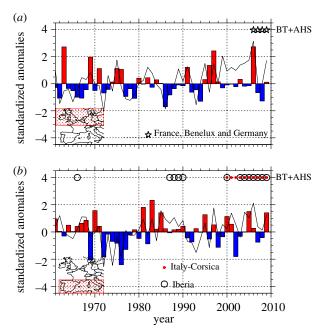


Figure 4.  $R_0$  recent evolution. (a)  $R_0$  standardized anomalies (histogram) over northwest and (b) southwest Europe. The standardized anomalies are computed retrieving the longterm mean and then weighting by the standard deviation (ASO 1961–1999). The area to compute the indices is displayed in the lower left corner. The observed BT and AHS outbreaks are represented for different areas by the associated markers.  $R_0$  is estimated from the observed climate dataset. The line represents anomalies of  $R'_0$  owing to the variations of the three virus transmission parameters  $(a, \mu \text{ and } \nu)$  with the vector-to-host ratio, m, held constant.

outbreak (in the event of viral introduction) than any of the previous 45 years. The fact that anomalies are negative for 2007 and 2008 in the NW, while the epizootic continued to spread, is not necessarily a discrepancy as  $R_0$  estimates only the initial spread of a disease, and in 2007 (and 2008), there were already significant numbers of infected holdings from which the disease disseminated. Other years of relatively high risk were 1963 and the mid-1990s. In SW Europe (figure 4b), there is a higher proportion of years with positive anomalies than in NW Europe, although they are generally of smaller magnitude. No BT was detected in SW Europe between 1961 and 1999. Nevertheless, there were outbreaks of a closely related viral disease of equids, African horse sickness (AHS), also transmitted by C. imicola, in Iberia in 1966 and in 1987–1991, both periods of positive anomaly for BT risk. The only decade without any recorded activity of either BT or AHS was the 1970s, a decade of consistently high negative  $R_0$  anomaly. Finally, considering BT in SW Europe in the last decade, the year with the strongest negative anomaly, 2002, stands out as the only year without new serotype introduction, and an unusually low reported BT incidence (see OIE Handistatus II Annual animal disease status Europe/2002/Bluetongue Animal health status at http://www.oie.int/hs2/sit\_mald\_cont. asp?c\_mald=10&c\_cont=4&annee=2002).

In figure 4a, b, the line represents the anomalies of  $R'_0$ owing to the variations of the three virus transmission parameters (i.e.  $a, \mu$  and  $\nu$ ) with the vector-to-host ratio, m, held constant. It shows that in NW Europe, over the last 10 years, 2006 stands out as the year when both m and the other transmission parameters were favourable for transmission. Conversely, between 2000 and 2005, all the transmission parameters but mwere favourable. In SW Europe, the anomalies of  $R'_0$ are more concordant than those of  $R_0$  with the epizootics of AHS which occurred between 1987 and 1990 on the Iberian peninsula, and with the epizootics of BT in 2004 and 2006 in southern Europe. This suggests that the climate in those years may have favoured disease transmission because of effects on the ability of the vectors to transmit the causative virus, rather than effects on the vector population size.

### 3.3. Future $R_0$ anomalies

The outputs from SimCTL (the ensemble of simulations fitted to past and present climatic conditions) integrated within the BT model reproduce very well the variability in  $R_0$  for the past period (1960–2000), in both NW (figure 5a) and SW (figure 5b) Europe. This confirms that simulated climate data can successfully drive the integrated BT model. Integrating SimA1B (the ensemble of simulations run under the climate change special report on emission scenario A1B), the mean  $R_0$  across the 11 RCMs is simulated to increase gradually between the present and 2050, but more rapidly in NW than in SW Europe (4.3%) versus 1.7% per decade). Model outputs also exhibit greater spread across the different RCM projections in the NW. Given susceptible ruminant host populations, our models suggest that by 2050,  $R_0$  will have increased by 30 per cent in NW and 10 per cent in SW Europe, with respect to 1961–1999 mean modelled risk levels in each of the two regions. Nevertheless, even in 2050, the absolute risk of BT transmission remains twice as high in SW than in NW Europe.

## 3.4. Influences of temperature and rainfall on $R_0$ anomalies

For NW Europe (figure 6a-d),  $R_0$  anomalies are mainly driven by changes in temperature, particularly via its influence on extrinsic incubation period, and, to a lesser extent, the biting rate. By contrast, with temperature set constant, there is very little trend in future  $R_0$  anomaly (less than 0.5% change per decade).

For SW Europe, the results are less robust (figure 6e-h). When temperature is set constant and only rainfall varies (figure 6f,h), the increase in  $R_0$  anomaly is 2 or 1.3 per cent per decade (20°C and 25°C, respectively), mainly via the effect of rainfall on vector-to-host ratio (m). However, the relation is complex, leading to opposite trends in  $R_0$ anomaly when rainfall is set constant (figure 6e, q). When rainfall is set at low values (p = 250 mm), the increase in temperature causes decrease in  $R_0$  anomaly while, at higher rainfall levels, increasing temperature leads to a slight increase in  $R_0$  anomaly. In other words, if SW Europe is dry, the vector-to-host ratio is simulated to decrease as temperatures increase, leading to the decrease in BT transmission risk. Conversely, if SW Europe is wetter, the increase in temperature then leads to a moderate increase in  $R_0$ .

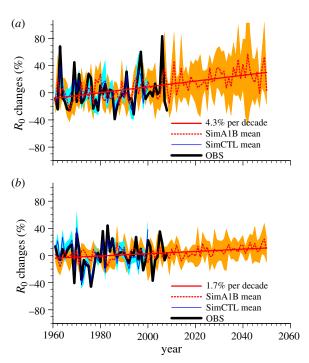


Figure 5. Simulated  $R_0$  future evolution.  $R_0$  relative anomalies (%) with respect to 1961–1999 time period in ASO for (*a*) northwest and (*b*) southwest Europe (see figure 4 for the domain definition).  $R_0$  estimated from the climate observations (OBS) is displayed in black, the  $R_0$  ensemble mean based on the SimCTL (SimA1B) RCM ensemble is displayed in blue (red). The blue (orange) envelope highlights the spread (defined as 1 s.d. of each RCM realization to the ensemble mean) within the SimCTL (SimA1B) ensemble.

# 3.5. Evaluation of the uncertainty of future $R_0$ anomalies

Focusing on the effects of future regional changes of climate (figure 7),  $R_0$  is simulated to increase by the SimA1B RCM ensemble over most of Western Europe. For the 2011–2030 period, climatic changes are projected to induce a significant increase in  $R_0$  in Ireland, Wales, southeast France and NW Iberia (figure 7a) consistent between the different climate models in terms of direction (figure 7c), and with a moderate spread in the projected magnitude (figure 7b). For the 2031-2050 period, Ireland, a larger part of Britain and NW Iberia show similar patterns (figure 7d-f). Over that period, changes in climate also induce a significant increase in BT risk in southeast France, but with a large spread in magnitude (figure 7e). By contrast, a limited area of southern Spain exhibits a small decrease in  $R_0$  which is consistent between models and in both time periods.

### 4. DISCUSSION

The dramatic emergence of BT in Europe—a disease with a known strong dependence on climate [44]—has enabled testing of our novel and generic framework to assess the effects of climate on the transmission of a disease. Applied to BT, the framework gives the best evidence to date that BT's emergence across Europe is related, at least partly, to climate change. It enabled the assessment of the changing risk of BT transmission (in the event of an introduction of the pathogen) in space and time, and also in which climate-sensitive biological mechanisms were involved. It further allowed us to investigate the change in risk of disease transmission in the future, given climate change, and to assess the relative influences of temperature and rainfall on this change in disease risk. This type of framework can be applied to other diseases for which the epidemiology is well described and where the links between the  $R_0$ parameters and climatic variables have been quantified.

The main limitations of the application of this framework to BT are owing to paucity of species-specific entomological knowledge on the parameters that determine vectorial capacity of European vectors. The complete list and relative importance (including competence) of the *Culicoides* species involved as well as their distribution and fine estimates of their biological parameters (such as the biting rate, extrinsic incubation period (EIP), mortality rate) and of the vector-to-host ratio would enable more robust modelling.

For example, the fact that the densities of *C. imicola* are overestimated in northern Italy could show that we omitted a factor influencing the establishment of this species. Conte *et al.* [27,28] have shown that soil type and vegetation cover impacted on its distribution in Italy. This information could be used to refine the distribution models. This can be done quite easily for variables which do not vary in time such as the soil type (supposing the data were available for the whole study area), but remains more difficult for other environmental variables that may be important, such as forest cover, which has varied over the last 50 years in Europe.

Modelling of the vector-to-host ratio component of the  $R_0$  disease model is problematic. First, the *Culicoides* population caught in light traps may not be fully representative of those that transmit BT virus. In one study, CDC (Center for Disease Control) traps baited with UV light tended to overestimate the numbers of C. imicola vacuumed off a sheep while underestimating those of C. obsoletus (although only 11 C. imicola were caught during the 8 days of the study [45]). Biteau-Coroller [46] captured once 23 per cent more and once 26 per cent less C. imicola in an Onderstepoort Veterinary Institute (OVI) trap than in a drop trap. Carpenter et al. [47] suggest that OVI light traps underestimate the role of some potential vector species such as *Culicoides chiopterus*. On the whole, while it is likely that light trap catches do not fully represent the population that feed on hosts, there is still no consensual method to correct for the bias. A second problem is that it remains unclear whether the number of midges caught in a trap should be treated as an estimate of the vector population size (as did Gubbins et al. [8] and Hartemink et al. [9]) or the vector-to-host ratio (as assumed here). We consider there to be problems with the former approach. First, there is no information on how to relate the number of midges caught in a trap to the number in the region being modelled: [9], for example, multiplied by an arbitrarily chosen constant (100) to convert a trap catch to the midge population per square-kilometre. Secondly, and as described earlier, it leads to problematically extreme estimates of  $R_0$  at extreme host densities (high or low). We consider that our novel approach, which uses trap catch as an estimate of the vector-to-host ratio, has the advantage of bypassing the problems

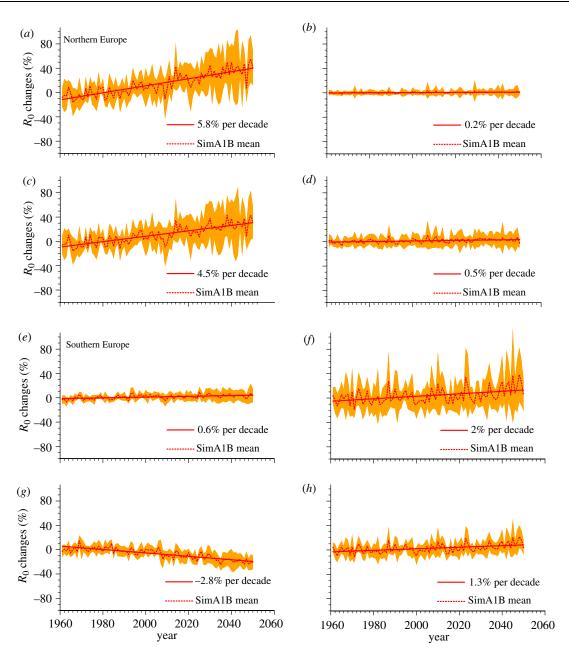


Figure 6. Sensitivity of  $R_0$  to changes in rainfall and precipitation.  $R_0$  relative anomalies (%) with respect to 1961–1999 time period in ASO for (a-d) northwest and (e-h) southwest Europe (see figure 4 for the domain definition). The anomalies are computed fixing (a,e) either precipitation constant to 500 mm or (c,g) to 250 mm or (b,f) temperature constant to 25°C or (d,h) to 20°C.

outlined above and ensures that  $R_0$  is proportional to trap catch. Nevertheless, formal testing of the two hypotheses has not yet been undertaken.

The observation that in southern Europe annual anomalies of  $R'_0$  (i.e.  $R_0$  modelled with varying viral transmission parameters but constant vector-to-host ratio) tended to be more concordant with epizootics than anomalies of  $R_0$  itself (with vector-to-host ratio also varying) is interesting. While there are insufficient data to draw robust conclusions, one possibility relates to the absence of lags in our model. Our model presumes near-instantaneous effects of climate variation on  $R_0$  (at least within the same three month period). In reality, the viral transmission parameters (a,  $\mu$ and  $\nu$ ) may be expected to respond quite rapidly to changes in climate, but there are likely to be significant lags in the response of the vector-to-host ratio, while the population size builds up and/or spreads over the years. If correct, this effect would probably affect the utility of annual anomalies in  $R_0$  (but not  $R'_0$ ), but should affect decadal anomalies in  $R_0$  less.

Despite these weaknesses, our novel framework successfully describes many aspects of BT's emergence, as demonstrated by the good concordance between model outputs and the observed distribution of the vectors and the disease. This shows that these potential biases have only had a moderate impact on the analysis. Nevertheless, given these approximations, we recommend not to compute absolute values of  $R_0$  but, until further data are available, to focus analysis on trends and anomalies.

Another limit of the application is that we explored only the effects of changing climate over time on BT's  $R_0$ , holding constant other factors which may also vary in time such as host densities. The fact that variation in climate alone successfully reproduces many aspects of this past emergence does not mean that

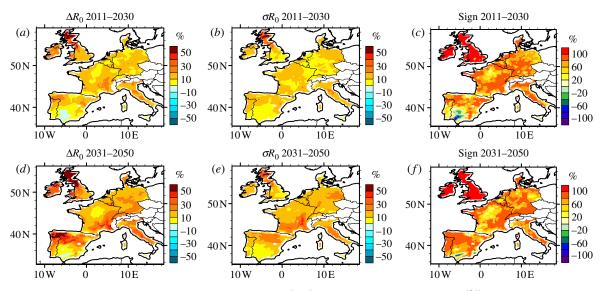


Figure 7. Simulated  $R_0$  changes, spread and sign consistency. (a,d) Simulated mean  $R_0$  changes (%) estimated from the SimA1B ensemble (with respect to the 1961–1999 climatology). (b,e) Multi-model spread defined as 1 s.d. of all RCM projections with respect to the ensemble mean. (c,f) Sign consistency (%), if all RCM projections agree on an increase, then the value is 100% and conversely for a decrease, -100%.

climate is the only driver, but it provides strong evidence that climate has played an important role. Indeed, ignoring a major driver would most likely result in substantial spatial and temporal discrepancies between predictions and the observed situation.

Further, this framework provides mechanistic insight into the drivers of the emergence, highlighting for the first time the role of climate-drivers of virus transmission, particularly the extrinsic incubation period, in NW Europe and a relatively larger role of climate-drivers of vector densities in SW Europe. It also confirms the role of temperature as a major driver of change in NW Europe (through the changes in EIP it produces), and a more complex situation in SW Europe where temperature influences differently the vector-to-host ratio and therefore  $R_0$  depending on whether the area is drier or wetter. When rainfall is low, the increase in temperature will lead to a decrease in risk, whereas when it is high, the increase of temperature will lead to an increase in risk. Finally, it has also permitted quantification of the effects of future changes of climate on BT risk of transmission in the event of viral introduction, projecting the  $R_0$  for BT to increase still further across much of Western Europe over the next 40 years, indicating an increasing threat to susceptible livestock from this disease.

Future simulations of the effects of climate on the disease must be interpreted with caution as there are still uncertainties related to the actual state-of-the-art climate model biases and associated with the selected emission scenario (A1B). Such approaches can be updated on a continuous-flow basis when climate data emanating from new models and/or forced by new emission scenarios are made available. Further research should also focus on testing the effects of a wider range of climatic variables on transmission parameters, including other variable statistics which could be more pertinent than mean values such as cumulative (e.g. degree days) or fluctuation [48] functions and evaluating the uncertainty of these relations. In parallel, climatic models and simulations need to be developed for these variables.

We have assessed only the effects of climate (mainly temperature and precipitation) on future  $R_0$  via direct effects on vectors and viral transmission parameters. This approach, namely holding constant non-climatic drivers, corresponds to the first of three contexts in which the effects of climate change on health can be evaluated [49]. The other contexts require the inclusion of non-climatic disease drivers in models. For BT, substantial knowledge gaps would need to be filled in order to take into account both the indirect knock-on effects of climate change [49] (on social, economic, political and land-use [50] changes), and non-climatic disease drivers. Although other possible non-climatic drivers of past BT occurrence have been discounted (see electronic supplementary material) [3]; for future periods, non-climatic drivers such as changes in livestock densities (or composition), and the development of novel control tools which reduce the exposure of naive hosts to vectors should not be ruled out as they could have major impacts on the future occurrence of BT. Indeed, we cannot be sure that other disease drivers will not arise and counteract or supplant these effects in the future.

The real future of BT in Europe will result from the combination of both climatic and non-climatic future changes. Further development of our approach would be the inclusion of additional, non-climatic drivers of BT spread; this implies that spatio-temporal estimates of the drivers, and their future trends, are available, and that their link to disease transmission is quantified. The ultimate aim would be to disentangle the interactions between drivers in order to then apply this approach for all drivers combined. Much more knowledge is needed about the different BT episystems (vectors, hosts, pathogens, biological controlling mechanisms and all the environmental factors), and on their future trends, before one can hope to reach this point.

Finally, simulating the effects of future climate on the risk of transmission of a disease in the event of viral introduction is very different from predicting where and when a disease will occur. The consequence of this is that the validation of this type of model is complex. Indeed, a vear with no epizootic does not mean that the conditions were not favourable for an epizootic; it might just be that no pathogen was introduced or that animals had been vaccinated. Thus, we can only verify the sensitivity of the model by comparing the first year of emergence of a new epizootic with the model's projections. Predicting where and when a disease will occur is out of the scope of the framework proposed here and, perhaps, remains a problem too complex to be addressed [51] as there may be changes over time in hosts, vectors, pathogens and the environment (including climate) and, most importantly, as this also depends on the probability of introduction of the pathogen.

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## REFERENCES

- Rogers, D. J. & Randolph, S. E. 2006 Climate change and vector-borne diseases. *Adv. Parasitol.* **62**, 345–381. (doi:10.1016/S0065-308X(05)62010-6)
- 2 Mellor, P. S., Carpenter, S., Harrup, L., Baylis, M. & Mertens, P. P. 2008 Bluetongue in Europe and the Mediterranean Basin: history of occurrence prior to 2006. *Prev. Vet. Med.* 87, 4–20. (doi:10.1016/j.prevetmed.2008.06.002)
- 3 Purse, B. V., Mellor, P. S., Rogers, D. J., Samuel, A. R., Mertens, P. P. & Baylis, M. 2005 Climate change and the recent emergence of bluetongue in Europe. *Nat. Rev. Microbiol.* 3, 171–181. (doi:10.1038/nrmicro1090)
- 4 Mehlhorn, H., Walldorf, V., Klimpel, S., Jahn, B., Jaeger, F., Eschweiler, J., Hoffmann, B. & Beer, M. 2007 First occurrence of *Culicoides obsoletus*-transmitted Bluetongue virus epidemic in Central Europe. *Parasitol. Res.* **101**, 219–228. (doi:10.1007/s00436-007-0519-6)
- 5 Meiswinkel, R., van Rijn, P., Leijs, P. & Goffredo, M. 2007 Potential new *Culicoides* vector of bluetongue virus in northern Europe. *Vet. Rec.* 161, 564–565.
- 6 Randolph, S. E. 2010 To what extent has climate change contributed to the recent epidemiology of tick-borne diseases? *Vet. Parasitol.* **167**, 92–94. (doi:10.1016/j. vetpar.2009.09.011)
- 7 Heffernan, J. M., Smith, R. J. & Wahl, L. M. 2005 Perspectives on the basic reproductive ratio. J. R. Soc. Interface 2, 281–293. (doi:10.1098/rsif.2005.0042)
- 8 Gubbins, S., Carpenter, S., Baylis, M., Wood, J. L. & Mellor, P. S. 2008 Assessing the risk of bluetongue to UK livestock: uncertainty and sensitivity analyses of a temperature-dependent model for the basic reproduction number. J. R. Soc. Interface 5, 363–371. (doi:10.1098/ rsif.2007.1110)

- 9 Hartemink, N. A., Purse, B. V., Meiswinkel, R., Brown, H. E., de Koeijer, A., Elbers, A. R. W., Boender, G.-J., Rogers, D. J. & Heesterbeek, J. A. P. 2009 Mapping the basic reproduction number (R0) for vector-borne diseases: a case study on bluetongue virus. *Epidemics* 1, 153–161. (doi:10.1016/j.epidem.2009.05.004)
- 10 Elbers, A. R., Backx, A., Meroc, E., Gerbier, G., Staubach, C., Hendrickx, G., van der Spek, A. & Mintiens, K. 2008 Field observations during the bluetongue serotype 8 epidemic in 2006. I. Detection of first outbreaks and clinical signs in sheep and cattle in Belgium, France and the Netherlands. *Prev. Vet. Med.* 87, 21–30. (doi:10.1016/j. prevetmed.2008.06.004)
- 11 Elbers, A. R., Backx, A., Mintiens, K., Gerbier, G., Staubach, C., Hendrickx, G. & van der Spek, A. 2008 Field observations during the Bluetongue serotype 8 epidemic in 2006. II. Morbidity and mortality rate, case fatality and clinical recovery in sheep and cattle in the Netherlands. *Prev. Vet. Med.* 87, 31–40. (doi:10.1016/j. prevetmed.2008.06.003)
- 12 Bonneau, K. R., DeMaula, C. D., Mullens, B. A. & MacLachlan, N. J. 2002 Duration of viraemia infectious to *Culicoides sonorensis* in bluetongue virus-infected cattle and sheep. *Vet. Microbiol.* 88, 115–125. (doi:10. 1016/S0378-1135(02)00106-2)
- 13 Menzies, F. D. et al. 2008 Evidence for transplacental and contact transmission of bluetongue virus in cattle. Vet. Rec. 163, 203–209. (doi:10.1136/vr.163.7.203)
- 14 Baylis, M., O'Connell, L. & Mellor, P. S. 2008 Rates of bluetongue virus transmission between *Culicoides sonor*ensis and sheep. *Med. Vet. Entomol.* 22, 228–237. (doi:10.1111/j.1365-2915.2008.00732.x)
- 15 Carpenter, S., Lunt, H. L., Arav, D., Venter, G. J. & Mellor, P. S. 2006 Oral susceptibility to bluetongue virus of *Culicoides* (Diptera: Ceratopogonidae) from the United Kingdom. J. Med. Entomol. 43, 73–78. (doi:10. 1603/0022-2585(2006)043[0073:OSTBVO]2.0.CO;2)
- 16 Gerry, A. C., Mullens, B. A., Maclachlan, N. J., Mecham, J. O. & Dada, C. E. 2001 Seasonal transmission of bluetongue virus by *Culicoides sonorensis* (Diptera: Ceratopogonidae) at a southern California dairy and evaluation of vectorial capacity as a predictor of bluetongue virus transmission. J. Med. Entomol. 38, 197–209. (doi:10.1603/0022-2585-38.2.197)
- 17 Nunamaker, R. A., Mecham, J. O., Holbrook, F. R. & Lockwood, J. A. 1997 Applications of dot-blot, ELISA, and immunoelectron microscopy to field detection of bluetongue virus in *Culicoides variipennis sonorensis:* an ecological perspective. J. Med. Entomol. 34, 24–28.
- 18 Mullens, B. A., Gerry, A. C., Lysyk, T. J. & Schmidtmann, E. T. 2004 Environmental effects on vector competence and virogenesis of bluetongue virus in *Culicoides*: interpreting laboratory data in a field context. *Vet. Ital.* **40**, 160–166.
- 19 Mullens, B. A. & Holbrook, F. R. 1991 Temperature effects on the gonotrophic cycle of *Culicoides variipennis* (Diptera: Ceratopogonidae). J. Am. Mosq. Control Assoc. 7, 588–591.
- 20 Gerry, A. C. & Mullens, B. A. 2000 Seasonal abundance and survivorship of *Culicoides sonorensis* (Diptera: Ceratopogonidae) at a southern California dairy, with reference to potential bluetongue virus transmission and persistence. *J. Med. Entomol.* **37**, 675–688. (doi:10. 1603/0022-2585-37.5.675)
- 21 Goldsmit, L., Barzilai, E. & Tadmor, A. 1975 The comparative sensitivity of sheep and chicken embryos to bluetongue virus and observations on viraemia in experimentally infected sheep. Aust. Vet. J. 51, 190–196. (doi:10.1111/j.1751-0813.1975.tb00053.x)

- 22 Melville, L. F., Weir, R., Harmsen, M., Walsh, S., Hunt, N. T. & Daniels, P. W. 1996 Characteristics of naturally occurring bluetongue viral infections of cattle. In *Bluetongue disease in Southeast Asia and the Pacific* (eds T. D. St George & P. Kegao), pp. 245–250. Canberra, Australia: ACIAR.
- 23 Veronesi, E., Hamblin, C. & Mellor, P. S. 2005 Live attenuated bluetongue vaccine viruses in Dorset Poll sheep, before and after passage in vector midges (Diptera: Ceratopogonidae). Vaccine 23, 5509–5516. (doi:10.1016/j. vaccine.2005.07.039)
- 24 Savini, G., Goffredo, M., Monaco, F., Di Gennaro, A., Cafiero, M. A., Baldi, L., de Santis, P., Meiswinkel, R. & Caporale, V. 2005 Bluetongue virus isolations from midges belonging to the Obsoletus complex (*Culicoides*, Diptera: Ceratopogonidae) in Italy. *Vet. Rec.* **157**, 133–139.
- 25 Robinson, T. P., Franceschini, G. & Wint, W. 2007 The Food and Agriculture Organization's Gridded Livestock of the World. *Vet. Ital.* **43**, 745–751.
- 26 Calvete, C., Estrada, R., Miranda, M. A., Borras, D., Calvo, J. H. & Lucientes, J. 2008 Modelling the distributions and spatial coincidence of bluetongue vectors *Culicoides imicola* and the *Culicoides obsoletus* group throughout the Iberian peninsula. *Med. Vet. Entomol.* 22, 124–134. (doi:10.1111/j.1365-2915.2008.00728.x)
- 27 Conte, A., Goffredo, M., Ippoliti, C. & Meiswinkel, R. 2007 Influence of biotic and abiotic factors on the distribution and abundance of *Culicoides imicola* and the Obsoletus Complex in Italy. *Vet. Parasitol.* **150**, 333–344. (doi:10.1016/j.vetpar.2007.09.021)
- 28 Conte, A., Ippoliti, C., Savini, L., Goffredo, M. & Meiswinkel, R. 2007 Novel environmental factors influencing the distribution and abundance of *Culicoides imicola* and the Obsoletus Complex in Italy. *Vet. Ital.* 43, 571–580.
- 29 Baylis, M., Bouayoune, H., Touti, J. & El Hasnaoui, H. 1998 Use of climatic data and satellite imagery to model the abundance of *Culicoides imicola*, the vector of African horse sickness virus, in Morocco. *Med. Vet. Entomol.* **12**, 255–266. (doi:10.1046/j.1365-2915.1998.00109.x)
- 30 Baylis, M., Mellor, P. S., Wittmann, E. J. & Rogers, D. J. 2001 Prediction of areas around the Mediterranean at risk of bluetongue by modelling the distributions of its vector using satellite imaging. *Vet. Rec.* **149**, 639–643. (+erratum *Vet. Rec.* 2002, 150(13) 404). (doi:10.1136/ vr.149.21.639)
- 31 Conte, A., Giovannini, A., Savini, L., Goffredo, M., Calistri, P. & Meiswinkel, R. 2003 The effect of climate on the presence of *Culicoides imicola* in Italy. J. Vet. Med. B 50, 139–147. (doi:10.1046/j.1439-0450.2003.00632.x)
- 32 Tatem, A. J., Baylis, M., Mellor, P. S., Purse, B. V., Capela, R., Pena, I. & Rogers, D. J. 2003 Prediction of bluetongue vector distribution in Europe and north Africa using satellite imagery. *Vet. Microbiol.* 97, 13–29. (doi:10.1016/j.vetmic.2003.08.009)
- 33 Baylis, M., el Hasnaoui, H., Bouayoune, H., Touti, J. & Mellor, P. S. 1997 The spatial and seasonal distribution of African horse sickness and its potential *Culicoides* vectors in Morocco. *Med. Vet. Entomol.* **11**, 203–212. (doi:10. 1111/j.1365-2915.1997.tb00397.x)
- 34 Haylock, M. R., Hofstra, N., Klein Tank, A. M. G., Klok, E. J., Jones, P. D. & New, M. 2008 A European daily highresolution gridded dataset of surface temperature and precipitation or 1950–2006. J. Geophys. Res. 113, D20119. (doi:10.1029/2008JD010201)
- 35 Uppala, S. M. et al. 2005 The ERA-40 re-analysis. Q. J. R. Meteorol. Soc. 131, 2961–3012. (doi:10.1256/qj.04.176)
- 36 Nakicenovic, N. & Swart, R. (eds) 2000 Special report on emissions scenarios: a special report of Working Group

III of the Intergovernmental Panel on Climate Change, 599 pp. Cambridge, UK: Cambridge University Press.

- 37 Delécolle, C. & de La Rocque, S. 2002 Contribution à l'étude des *Culicoides* de Corse. Liste des espèces recensées en 2000/ 2001 et redescription du principal vecteur de la fièvre catarrhale ovine: *C. imicola* Kieffer, 1913 (Diptera, Ceratopogonidae). *Bull. Soc. Entomol. France* 107, 371–379.
- 38 Baldet, T., Mathieu, B., Delécolle, J.-C., Gerbier, G. & Roger, F. 2005 Emergence de la fièvre catarrhale ovine dans le Bassin méditerranéen et surveillance entomologique en France. *Rev. Elev. Méd. vét. Pays trop.* 58, 125–132.
- 39 Goldarazena, A., Romon, P., Aduriz, G., Balenghien, T., Baldet, T. & Delecolle, J. C. 2008 First record of *Culicoides imicola*, the main vector of bluetongue virus in Europe, in the Basque Country (northern Spain). *Vet. Rec.* 162, 820–821. (doi:10.1136/vr.162.25.820)
- 40 Sarto I Monteys, V. & Saiz-Ardanaz, M. 2003 Culicoides midges in Catalonia (Spain), with special reference to likely bluetongue virus vectors. Med. Vet. Entomol. 17, 288–293. (doi:10.1046/j.1365-2915.2003.00441.x)
- 41 De Liberato, C., Purse, B. V., Goffredo, M., Scholl, F. & Scaramozzino, P. 2003 Geographical and seasonal distribution of the bluetongue virus vector, *Culicoides imicola*, in central Italy. *Med. Vet. Entomol.* **17**, 388– 394. (doi:10.1111/j.1365-2915.2003.00456.x)
- 42 Conte, A., Gilbert, M. & Goffredo, M. 2009 Eight years of entomological surveillance in Italy show no evidence of *Culicoides imicola* geographical range expansion. *J. Appl. Ecol.* 46, 1332–1339. (doi:10.1111/j.1365-2664. 2009.01723.x)
- 43 Thiry, E. et al. 2006 Bluetongue in northern Europe. Vet. Rec. 159, 327. (doi:10.1136/vr.159.10.327)
- 44 Mellor, P. S., Boorman, J. & Baylis, M. 2000 Culicoides biting midges: their role as arbovirus vectors. Annu. Rev. Entomol. 45, 307–340. (doi:10.1146/annurev.ento.45.1.307)
- 45 Gerry, A. C., Sarto i Monteys, V., Moreno Vidal, J. O., Francino, O. & Mullens, B. A. 2009 Biting rates of *Culicoides* midges (Diptera: Ceratopogonidae) on sheep in northeastern Spain in relation to midge capture using UV light and carbon dioxide-baited traps. *J. Med. Entomol.* 46, 615–624. (doi:10. 1603/033.046.0329)
- 46 Biteau-Coroller, F. 2006 Surveillance et évaluation du risque de transmission des maladies vectorielles émergentes: apport de la capacité vectorielle. Exemple de la fièvre catarrhale du mouton. Doctorat en Epidémiologie, Université de Montpellier II, Montpellier, France, 238 pp.
- 47 Carpenter, S., Szmaragd, C., Barber, J., Labuschagne, K., Gubbins, S. & Mellor, P. 2008 An assessment of *Culicoides* surveillance techniques in northern Europe: have we underestimated a potential bluetongue virus vector? *J. Appl. Ecol.* **45**, 1237–1245. (doi:10.1111/j.1365-2664. 2008.01511.x)
- 48 Paaijmans, K. P., Read, A. F. & Thomas, M. B. 2009 Understanding the link between malaria risk and climate. *Proc. Natl Acad. Sci. USA* **106**, 13844–13849. (doi:10. 1073/pnas.0903423106)
- 49 McMichael, A. J., Woodruff, R. E. & Hales, S. 2006 Climate change and human health: present and future risks. *Lancet* **367**, 859–869. (doi:10.1016/S0140-6736 (06)68079-3)
- 50 Tubiello, F. N., Soussana, J. F. & Howden, S. M. 2007 Crop and pasture response to climate change. *Proc. Natl Acad. Sci. USA* **104**, 19686–19690. (doi:10.1073/pnas. 0701728104)
- 51 Tabachnick, W. J. 2010 Challenges in predicting climate and environmental effects on vector-borne disease episystems in a changing world. J. Exp. Biol. 213, 946–954. (doi:10.1242/jeb.037564)