



**Article title:** Plastic agriculture using worms: Augmenting polystyrene consumption and using frass for plant growth towards a zero-waste circular economy.

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1  
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4 To the Editor

5  
6 Dear Editor,

7  
8 We would like to present our manuscript titled “**Plastic agriculture using worms:  
9 Augmenting polystyrene consumption and using frass for plant growth towards a zero-  
10 waste circular economy**” for publication in your journal.

11  
12 In this research, we studied the effects of food additives on polystyrene (PS) consumption by  
13 mealworms and superworms, as well as the use of their frass for an indoor dragon fruit cactus  
14 (*Hylocereus undatus*) that is both an ornamental and food crop plant. We found that small  
15 amounts of common condiments augmented their natural PS consumption, potentially  
16 addressing PS waste often contaminated with food. We found the frass of superworms fed on  
17 PS alone did not show obvious difference from those fed on bran as determined by GC-MS,  
18 and in fact supported rooting and comparable cacti growth better than mealworm frass.

19  
20 Our research here shows promising solutions to plastic pollution and urban food production  
21 in the society today. Using purely natural solutions, worm are a feasible solution to close the  
22 loop in a circular zero waste economy that is also implementable even indoors. The study sheds  
23 light on the promise of worms that has been gaining a lot of attention for plastic waste  
24 management, and the potential of the frass for further agricultural uses. Our findings have  
25 significant impact on both ecological health and environmental quality. Preprint is on BioRxiv  
26 [doi.org/10.1101/2020.05.29.123521](https://doi.org/10.1101/2020.05.29.123521)

27  
28 We hope this article would find a home in your journal, as we believe it is useful and of  
29 interest to the scientific community and public

30  
31  
32  
33 Yours Faithfully,

34  
35 Samuel Ken-En Gan

36 On behalf of the authors  
37  
38

39 **Plastic agriculture using worms: Augmenting polystyrene consumption and using frass**  
40 **for plant growth towards a zero-waste circular economy.**

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66 **ABSTRACT**

67 Polystyrene (PS) is one of the major plastics contributing to environmental pollution with its  
68 durability and resistance to natural biodegradation. Recent research have found mealworms  
69 (*Tenebrio molitor*) and superworms (*Zophobas morio*) to be able to utilize PS as a carbon food  
70 source and degrade them without observable toxic effects. In this study, the effects of food  
71 additives on plastic consumption augmentation were studied, where small additions of sucrose  
72 and bran found to increase PS consumption. To close the plastic carbon cycle, we further  
73 evaluated the use of worm frass for dragon fruit cacti (*Hylocereus undatus*) growth and found  
74 that superworm frass supported rooting and growth better than mealworm frass and control  
75 media over a fortnight. Superworms, apart from being known fish and poultry feed, have been  
76 shown to be a suitable natural solution to the PS plastic problem that can support plant growth  
77 towards a zero-waste sustainable bioremediation cycle.

78 **Keywords:** Biodegradation, Mealworm, Superworm, Frass, Polystyrene, rooting, agricultural  
79 support, waste management

80

81

## 82 Introduction

83 Widely credited to Ray McIntire of the Dow Company (Scheirs, 2003), styrofoam or  
84 polystyrene (PS) are light polymers which have low heat conductivity (Campbell, 2012;  
85 Scheirs, 2003) and can be synthesized to different shapes and sizes, making it a highly versatile  
86 material. From insulating material for buildings to packaging food and beverages, PS is used  
87 worldwide, but it does not have an innocuous place in marine or terrestrial environments as its  
88 resistance to chemical degradation results in accumulation and pollution (Rochman et al.,  
89 2013). While one current large-scale management of PS waste is incineration, this leads to the  
90 release of toxic fumes to cause air pollution (Elizabeth Royte, 2019; Ritchie, 2018; Verma et  
91 al., 2016).

92 Superworms (*Zophobas morio*) and mealworms (*Tenebro molitor*) belong to the  
93 darkling beetle (*Tenebrionidae*) family and are naturally voracious insect pests in agriculture,  
94 consuming dry grain stock even though they are also food sources in some cultures (Sogari et  
95 al., 2019). Recently, mealworms were shown to be able to naturally consume, metabolize and  
96 mineralize the carbon in PS (Yang et al., 2015a, p. 1), an ability attributed to the commensal  
97 gut bacteria in these worms, verified by <sup>13</sup>C-carbon isotope tracing experiments among others  
98 (Yang et al., 2015b, p. 2). At the larval stage, they can be bred at high density, excreting  
99 nitrogen (Kagata and Ohgushi, 2012) and chitin rich (Finke, 2007; Soon et al., 2018) frass  
100 waste that were shown to improve growth and yields of many plants (Egusa et al., 2015;  
101 Houben et al., 2020; Poveda et al., 2019). With the potential of mealworm frass to substitute  
102 traditional NPK (Nitrogen, Phosphorus, and Potassium) based fertilizers, these worms were  
103 suggested to fuel the circular economy (Houben et al., 2020). With recent reports that  
104 superworms were able to consume PS at a rate higher than mealworms (Yang et al., 2020),  
105 there is promise in using superworms to join mealworms in the fight against plastic pollution.

106            Since food containers form the bulk of PS waste, they are often contaminated with food  
107 waste, complicating recycling methods that require clean plastic waste. In this aspect, the  
108 possible use of food contaminants to speed up plastic degradation by worms may be a natural  
109 solution that has yet to be fully exploited. Combining this with the fact that the frass can in turn  
110 be used to support plant growth, particularly food crop agriculture production, these worms are  
111 the key to turn plastic waste into fertilizers for food production with zero waste.

112            To evaluate the possibility, this study aims to investigate 1) the efficacy of food  
113 additives to augment PS degradation by mealworms and superworms; and 2) evaluate the use  
114 of the frass of super and mealworms to support dragon fruit (*Hylocereus undatus*) cacti grown  
115 (evaluated by rooting and growth), chosen as it is an easy growing indoor fruit plant with  
116 potential urban farming applications.

117

## 118 **Materials and methods**

### 119 **Insect rearing and frass collection**

120 Superworms (*Zophobas morio*) and mealworms (*Tenebrio molitor*) fed on bran were purchased  
121 as fish and bird feed sold in pet stores in the Clementi, Singapore. They were weighed and  
122 transferred to polypropylene (PP) containers (impervious to the worms) with the respective test  
123 food condiments (see Figure 1A & B for experimental setup). The collection of worm frass  
124 was performed by sifting the contents of the containers with a mesh sieve to remove uneaten  
125 PS/food and worm parts (if any). The worms were kept in cardboard boxes with a constant  
126 humidity of ~50% and a temperature of ~25°C (previously reported to be ideal for PS  
127 consumption by worms, Yang et al., 2018a) monitored by assembled Arduino devices (not  
128 shown).

129

130 **PS consumption rate experiments**

131 The natural rate of PS consumption (mg of PS / g of worm per day) by superworms and  
132 mealworms in our setup were determined by rearing them separately in replicates. To control  
133 for different worm sizes, the experimental setups were based on total worm weights of between  
134 6.22 -10.76 g and 300-390 mg of PS balls with diameters between 0.4 to 0.5 cm (Art friend,  
135 Singapore) in replicate set ups (Figure 1A & B). For food additives testing, PS balls were  
136 premixed with 25 mg of either cinnamon (Masterfoods, Australia), bran (Bob's Redmill,  
137 America), table sucrose (Lippo group) or no additive (control) in polypropylene containers. To  
138 improve adherence of food additives to PS balls, 0.9 ml of water was added to the mix.  
139 Unconsumed PS balls were collected after 4 days and weighed on an analytical balance. The  
140 final total live worm weights (Table 2) were used for calculation. Experiments were repeated  
141 in sextuplicates.

142

143 **Worm frass and Dragon fruit (*Hylocereus undatus*) experiment setups**

144 Frass obtained from superworms and mealworms previously reared solely on PS balls were  
145 evaluated as media for *Hylocereus undatus* cacti. Stock cacti were grown from seeds in air-  
146 conditioned office environments for more than four years prior and expanded from the same  
147 original pot. The grafting method of budding offshoots was used to expand cacti successfully  
148 multiple times on used Chinese tea leaves (termed tea leaves) throughout the four years. For  
149 this evaluation experiment, the same grafting method was used to transplant 48 selected cacti  
150 branches of as similar size as possible onto the test media and grown in cleaned plastic  
151 wineglasses in individual setups (Figure 1C) with the funnelled bottom to support upright

152 supplanting of the cacti. Media tested included: used leaves, bran (Bob's Redmill, America),  
153 superworm frass, and mealworm frass. For each set-up, the media covered the bottom of the  
154 grafted cacti, supporting it to stay upright to form the soil line.

155 The grafted cacti were lined up against a window ledge and watered every 3 times a week to  
156 wet the media. As much as possible, equal conditions were applied for 11 replicates. The height  
157 of the straightened grafted cacti (from the tip to the bottom of the stem, excluding roots) were  
158 measured before grafting and after a period of two weeks. Only the rooting for living cacti  
159 below the soil line are considered (to rule out confounding existing aerial roots which occur  
160 above the soil line, and that rooting below the soil line demonstrate direct effects of the media).  
161 Observed rooting of the grafted cacti were recorded qualitatively with photographs. Dead cacti  
162 (supplementary table S1) were also recorded.

163

#### 164 **GC-MS analysis of superworm frass**

165 For characterisation, GC-MS analysis was performed as adapted and modified from a previous  
166 report (Yang et al., 2015b). PS balls or frass (20 mg) from superworms reared on either  
167 polystyrene or bran were dissolved in a consistent manner by gram of frass to fixed volume of  
168 gas chromatography grade dichloromethane solvent for standardized comparisons of peak  
169 heights later, and incubated in 2 ml microfuge tubes on a shaker rack for 10 minutes and  
170 subsequently centrifuged (14.8k RPM, 5 minutes using table top centrifuge) to remove  
171 undissolved solids. The solvent soluble samples were filtered using a 0.45 µm teflon syringe  
172 filter and analysed on a GC-MS system (HP 6890 gas chromatography HP-5MS column and  
173 HP 5973 mass spectrometry). The GC oven temperature was set to 50°C for 1 minute and  
174 250°C for 5 minutes by ramp up rate at 10°C/minute.

175

## 176 **Results**

### 177 **Augmentation effects of food additives/condiments on polystyrene consumption.**

178 Mealworms and superworms were reared on PS with/without common food additives:  
179 cinnamon, sucrose, and bran. Bran was the initial food source the worms were supplied in, and  
180 thus was used as a control. It was also previously reported to increase the rate of PS  
181 consumption when supplemented at half the weight of PS (Yang et al., 2018a). From our  
182 results, addition of all three food additives significantly increased the rate of PS consumption  
183 in mealworms ( $p < 0.1$ ) (Figure 2). Whereas only the addition of sucrose increased the rate of  
184 PS consumption in superworms. Small amounts of sucrose or bran (25 mg) were found to more  
185 than double the PS consumption rate from an average of 1.035 and 1.40 mg / g of worm per  
186 day to 1.79 (not statistically significant) and 2.14 ( $p < 0.05$ ) when bran was used as an additive,  
187 and 1.9 ( $p < 0.1$ ) and 3.55 mg / g ( $p < 0.05$ ) of worm per day when sucrose was added to PS  
188 for superworms and mealworms respectively. In mealworms coted with small amounts of  
189 sucrose, the mealworms were observed to significantly consume more PS than those coted with  
190 bran (Figure 2). Comparing the efficacy of sucrose on mealworms and superworms,  
191 mealworms significantly ate more PS. With the exception of superworms fed on PS with  
192 sucrose additives that recorded a slight increase in weight 1.79% ( $p < 0.1$ ), no significant worm  
193 weight change were observed in either the mealworms or superworms over the period of four  
194 days on the PS diets (See Table S1 in the supplementary data).

195

### 196 **Effect of Superworm and Mealworm frass on Plant growth and rooting.**

197 We next sought to determine if frass from worms solely fed on PS can be used as an  
198 alternative growth media for plants to evaluate the zero-waste circular economy solution. From

199 our results, the superworm frass supported a significantly higher proportion of rooting for the  
200 grafted dragon fruit cacti offshoots compared to those grown on spent tea leaves or bran  
201 (Figures 3 & 4). In superworm frass media, nine cacti rooted (90%) compared to the tea leaves  
202 with five cacti rooting (45.5 % rooted,  $p < 0.05$ ); or to those grown on bran, four cacti rooted  
203 (36.4 % rooted,  $p < 0.05$ , see Table 1). With respect to cacti height growth, plants grown on  
204 the superworm frass media gained an average height of 0.5 cm that was not significantly  
205 different from those grown on tea leaves (average gain of 0.14 cm). Mealworm frass media  
206 alone significantly impaired the growth of plants which lost an average height of 0.52 cm ( $p <$   
207  $0.05$ ), due to water loss, possibly also reflecting the water holding abilities of the frass. It was  
208 also observed that 5 out of a total of 11 cacti across triplicates died when grown on mealworm  
209 frass alone.

210

211 A loss of 0.43 cm was also observed in plants grown on bran but was not significantly different  
212 from the other test media. There were no significant differences between the number of rooting  
213 cacti of mealworm frass to both tea leaves and bran. Superworm frass significantly supported  
214 rooting better compared to the other media (Figure 3, Figure S1).

215

### 216 **GC-MS analysis of superworm frass**

217

218 To investigate the presence of PS and possible by-products e.g., styrene, the superworm frass  
219 were collected and analysed using Gas chromatography–mass spectrometry (GC-MS).  
220 Analysis of the PS balls alone showed peaks corresponding to styrene and molecules containing  
221 benzyl groups, but no notable corresponding peaks were observed in the filtered frass from the  
222 superworms reared on PS balls (Figure 5). The frass samples had notable peaks corresponding  
223 to 9-oleamide ( $C_{18}H_{35}NO$ ) fatty acid primary amides (FAPA) along with smaller peaks  
224 corresponding to mainly other FAPAs, short chain alkanes, alcohols, and cycloalkanes (Figure

225 5, Table 3). In general, there were no notable significant differences between the GC-MS  
226 analysis of the frass of the superworms fed on PS and bran.

227

## 228 **Discussion**

229 We set out to investigate the effects of food additives on the rate of PS consumption by  
230 mealworms and superworms, and the feasibility of their frass alone to support plant growth,  
231 assessed by the growth height gain and rooting of grafted dragon fruit cacti offshoots.

### 232 *Food additives*

233 Of the food additives, small additions of table sucrose (25 mg) were found to be the  
234 most effective, conferring mealworms greater consumption of the PS balls when compared to  
235 mealworms and superworms coted on bran and sucrose respectively. Mealworms and  
236 superworms fed with sucrose additives experienced the highest increase in PS degradation,  
237 ~2.5 ( $p < 0.05$ ) and ~ 1.8-fold ( $p < 0.1$ ) respectively. Bran, previously reported to double the  
238 rate of PS consumption when supplemented at half the PS substrate weight (Yang et al., 2018a),  
239 was also found in our study to increase the rate of PS consumption by ~1.7 (not statistically  
240 significant) and ~1.5 ( $p < 0.05$ ) folds in superworms and mealworms, respectively. This was  
241 higher than cinnamon- supplemented PS, and in the absence of food additives. It should be  
242 noted that cinnamon elicited a significantly higher rate than no additive control for superworms,  
243 but not for mealworms. While this might be due to multiple factors, ranging from different  
244 taste receptors to metabolic/microbial processing of cinnamon, cinnamon did not have negative  
245 impact on mealworm consumption of PS. Given that most PS waste are food packages, the  
246 findings here bode well since both mealworms and superworms can consume organic waste  
247 and be reared in high densities. There was no significant weight loss over the 4 days of  
248 experimentation (see Table 2). Although a previous study showed possible hindering of  
249 mealworms life cycle on a plastic diet (Matyja et al., 2020), we did not observe notable  
250 abnormalities during our worm breeding other than delayed stages (which is beneficial for  
251 plastic degradation) given that the larvae ate more than adult beetles. In fact, our superworm

252 beetles laid eggs to give rise to a second generation of superworms growing on a pure plastic  
253 diet as their parent generation did (data not shown).

#### 254 *Comparison of superworms to mealworms*

255 Comparing the rate of PS consumed by weight of worms per day, there were no  
256 significant difference between mealworms and superworms (for control conditions in Figure  
257 2), which was contrary to a recent report that showed superworms to be superior to mealworms  
258 in PS consumption (Yang et al., 2020). This was not unexpected as the study calculated and  
259 used the rate of PS consumption per individual worm as the basis of comparison. As the  
260 difference in mass of a single mealworm compared to a superworm could be as high as 20  
261 folds, calculating by weight rather than number of worms may have normalized the  
262 underestimation of mealworm productivity. Given that in future real-life application, actual  
263 counting of worms is not feasible to deal with the tonnes of PS waste generated daily, we  
264 adopted weight as the measurement for future scalability purposes.

265

#### 266 *Zero-waste circular economy*

267 Both mealworms and superworms are known fish (Henry et al., 2015) and poultry  
268 (Finke, 2007) feed, and now with the added advantage of being valuable plastic degraders  
269 (Yang et al., 2018a, 2018b, 2020, 2015a, 2015b). As a potential cost-effective feed source for  
270 urban poultry and fish farming, the worms could already contribute to addressing both plastic  
271 and food production problems. While further research is necessary to ensure that plasticizers  
272 or other plastic degradation products do not bioaccumulate and get introduced into the food  
273 chain to humans (see reviews on plasticizer accumulation in the food chain, EFSA Panel on  
274 Contaminants in the Food Chain (CONTAM), 2016; Toussaint et al., 2019), there is great

275 promise in the use of the worms themselves as a solution to plastic waste. In fact, one added  
276 advantage of mealworms and superworms over other beetles larvae, is that unlike black soldier  
277 flies that are commonly used for food waste (Palma et al., 2019), the mature darkling beetles  
278 have fused wings/elytra and do not fly, making their biocontainment significantly easier for  
279 plastic degradation setups. Thus, any large-scale setups can be performed with minimal  
280 concern for their escape, leaving their frass to be addressed in a zero-waste circular economy.

### 281 *Frass analysis*

282 We did not focus our frass analysis on the mealworm frass as they did not support cacti  
283 growth, and the literature on plastic degrading mealworms was already quite extensive  
284 (Houben et al., 2020; Yang et al., 2018b). Through GC-MS analysis, we did not detect styrene  
285 in the frass of the PS fed superworms despite it being detected in the PS ball control analysis,  
286 nor were there any major notable additional degradation products in the filtered frass of  
287 superworms fed on PS compared to those fed on bran. Further evaluation using more sensitive  
288 methods to rule out possible bioaccumulation, including testing a wider range of PS products  
289 such as coloured or other PS products with additives should be performed before  
290 implementation in real-life settings. As was found in previous reports (Yang et al., 2018a,  
291 2018b, 2020, 2015a, 2015b), microplastics can be present in the frass and these can indeed  
292 pose a problem as they accumulate. However, it is possible that with repeated re-digestion of  
293 the frass by the worms, this problem can be mitigated. A possible industrial set up is to have a  
294 multi-layered process where frass is then fed to another chamber of worms and the process  
295 repeated until complete removal of microplastics. It is possible that both mealworms and  
296 superworms can be utilized in such a setup as our other experiments (not shown) showed that  
297 apart from occasional cross-eating, they can be bred together.

298 For ease of operation, the dragon fruit cacti (*Hylocereus undatus*), an easy to grow  
299 indoor plant that is both an ornamental and food crop was chosen for the evaluation of frass for  
300 urban farming. The superworm frass alone was found to be better at supporting rooting (90%  
301 rooting compared to 45.4% in used tea leaves) and was at least as effective as spent tea leaves  
302 (the media used in the last four years to expand the cacti) in supporting growth as determined  
303 by cacti height gain over a fortnight. Mealworm frass on the other hand, resulted in a lot of  
304 failed grafts (likely due to its observed poorer ability to retain moisture), while bran media  
305 resulted in poor height and rooting support, with shrinkage due to water loss in some replicates.  
306 It is possible that short chain growth promoting alkene semiochemicals which were detected  
307 from GC-MS analysis (e.g. Heptacosane, Nonadecane and Octadecane, Jishma et al., 2017), as  
308 well as chitin in the superworm frass may have augmented rooting (chitin was previously  
309 reported to support rooting, Winkler et al., 2017), or that there was some other auxin like  
310 compound present that would require further analysis. It should also be noted that the  
311 superworm frass was less pungent than the ammonia smelling mealworm frass, providing more  
312 reasons beyond rooting and comparable cacti growth for the use of superworm frass.

313 The lack of support of mealworm frass on dragon fruit cacti growth is unexpected given  
314 a previous report (Houben et al., 2020; Poveda et al., 2019). This difference may be due to the  
315 usage of 100% frass for our evaluation or due to the different nutritional requirements of the  
316 dragon fruit cacti, or though unlikely, the difference in frass from mealworms that are fed  
317 purely on PS. It may be possible to further evaluate mealworm frass for other food crops.

318

### 319 *Further research*

320 Given that there was no known benefit of mealworm frass in the dragon fruit cacti in  
321 our setups, and that consumption of PS by both mealworms and superworms showed no

322 difference, the use of superworms over mealworms is proposed here to be the better candidate  
323 in closing the loop from plastic to fish/poultry feed to frass-supported agriculture. Much  
324 remains to be studied on the possible accumulation of plasticizers or other plastic-derived  
325 chemicals as well as the frass on a variety of other food crops, but current results are promising  
326 with previous studies showing about 40 to 50% degradation of polystyrene monomers in  
327 mealworms (Yang et al., 2015a) and superworms (Yang et al., 2020), respectively in the span  
328 of a fortnight as determined by respirometry experiments. With further incubations of the waste  
329 frass supplemented with food additives and even tailored microbial assimilation of the PS  
330 polymers, total degradation could be made more complete if necessary, addressing even  
331 microplastics. Since many plants are able to clear up toxins from the environment (Cristina  
332 Negri and Hinchman, 1996), it is possible that any potential toxic substances arising from other  
333 plastics, could be combined with phytoremediation (Cristina Negri and Hinchman, 1996; Negri  
334 and Hinchman, 1996).

335 There are exciting research based on enzymes isolated from bacteria present in plastic eating  
336 worms (Austin et al., 2018; Danso et al., 2019; Palm et al., 2019; Yoshida et al., 2016), but the  
337 implementation of these processes towards complete degradation into harmless substances at  
338 industrial scale will require further research and engineering in the face of an urgently  
339 increasingly pressing problem of plastic waste made worse by lockdown measures during the  
340 COVID19 pandemic. In the meantime, the natural solution of worms can be investigated  
341 further for more immediate implementation, especially their simultaneous roles for urban  
342 farming in both fish/poultry feed and their frass for food crops. Worms are naturally more  
343 resistant to environmental factors compared to pure enzymes and can overcome obstacles for  
344 enzymes in plastic crystallinity or accessibility of the polymer chains, such that while protein  
345 engineering of such enzymes (Ma et al., 2018) are promising, there is still much to optimize  
346 before large scale implementation compared to worms.

347           The setups of both PS consumption by worms and frass-supported cacti growth were  
348 all performed indoors, demonstrating the possibility of worms to be an environmentally  
349 friendly urban solution to plastic waste and food sustainability that can be implemented widely,  
350 even within homes.

## 351 **Conclusion**

352           In conclusion, with evidence that food additives augment rather than antagonize PS  
353 degradation, and that the frass can be used to support food crop growth while the worms are  
354 themselves sources of poultry and fish feed, the answer in the worms is a very fitting scalable  
355 solution to both the plastic pollution and food (aquaculture and agriculture) production  
356 problems.

357

## 358 **Declarations and Conflict of Interests**

359 The authors declare no conflict of interest with this work.

360

## 361 **Author Contributions**

362 DWSK, BYXA, JYY, SKEG performed the worm culturing and plant growth experiments. ZX  
363 performed the GCMS experiments. DWSK, JYY, SKEG, analysed the results and wrote the  
364 manuscript. SKEG designed and supervised all aspect of the study. All authors read and  
365 approved the manuscript.

366

367

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373

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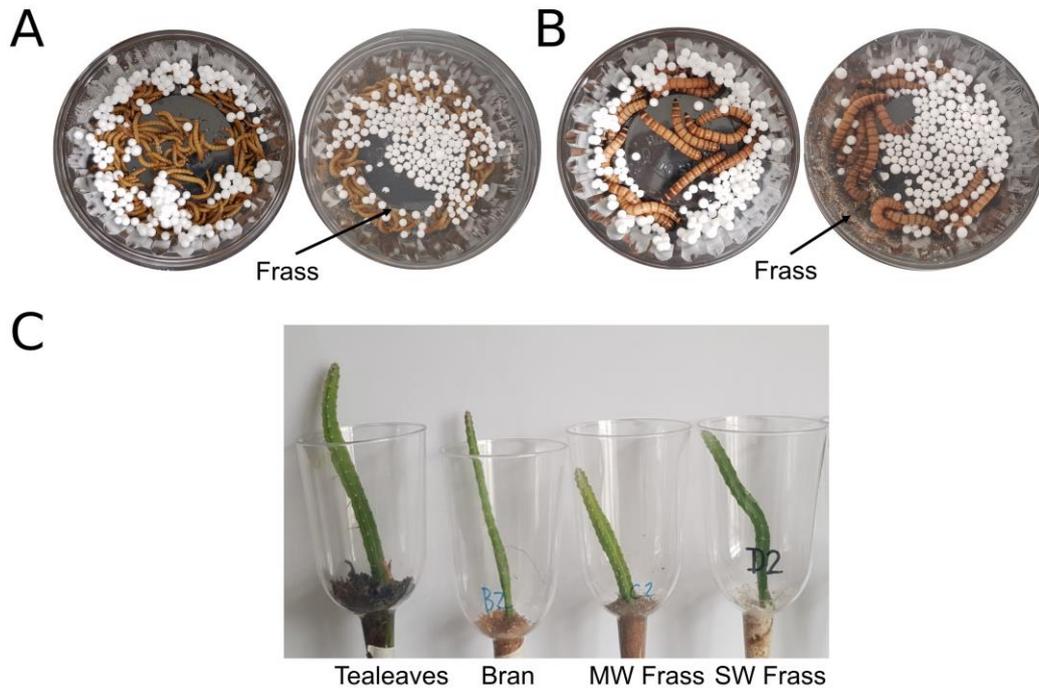
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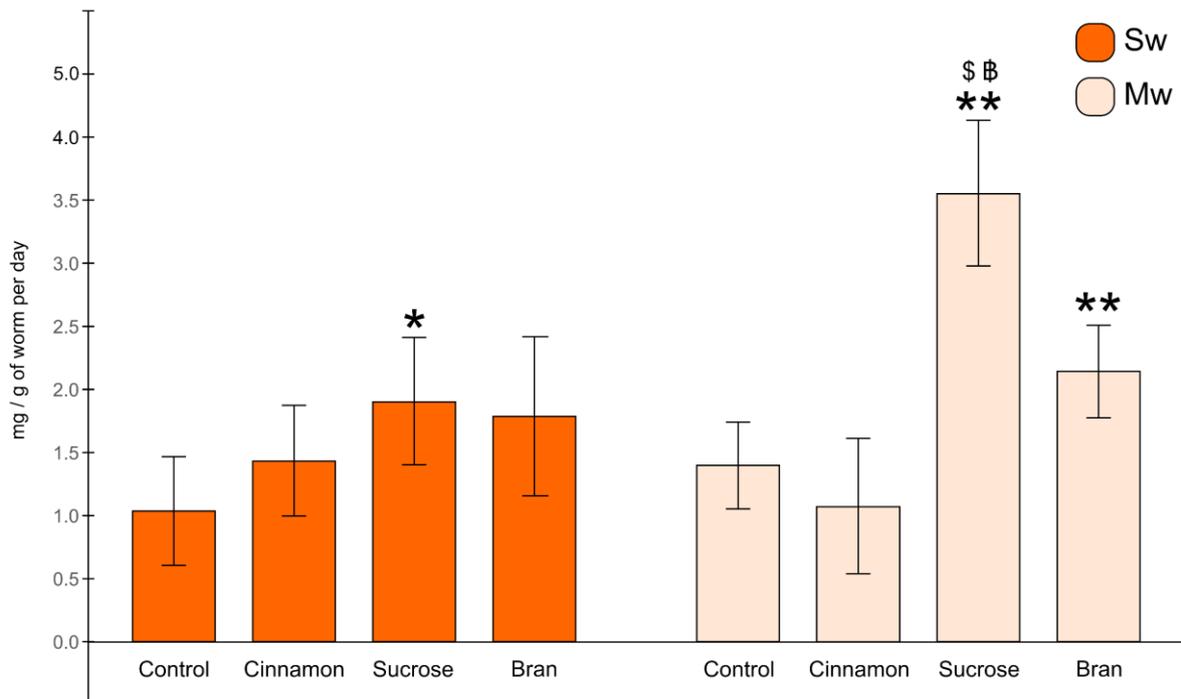
**Table 1.** Effect of different medias on number and proportion of rooting cacti. n=44, df=1

Media	Control	Proportion of Cacti rooted	Observed Total Number of Cactus Rooted	Observed Total Number of Cactus Not Rooted	Total	Pearson's $\chi^2$ P-value	Pearson's $\chi^2$ totals
Tea leaves	Bran	45.5%	5	6	11	.66	0.2
	Bran	36.4%	4	7	11	.66	0.2
Mealworm frass ^	Tea leaves	15%	2	4	6	.49	0.5
	Bran					.63	0.2
Superworm frass ^	Tea leaves	90%	9	1	10	.03**	4.7
	Bran					.01**	6.4

*Note.* \*\* denotes  $p < .05$ . ^ Five out of eleven cactus plants with mealworm frass died. One out of eleven cactus plants with superworm frass died. Rooting was counted only if they appeared below the soil line as the cacti often had pre-existing aerial roots.

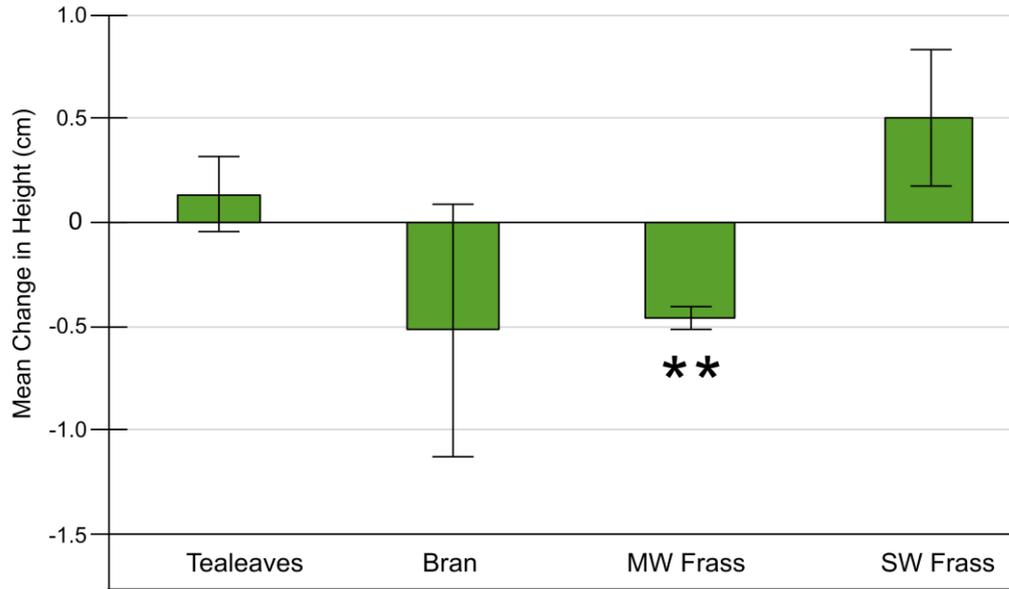


496 **Figure 1:** Representative images of the setups for testing PS consumption rates by (A)  
497 mealworms; (B) Superworms. For both A and B, the left are the initial setups, and the right  
498 showed the setup after four days where frass was produced from the PS consumption. (C) Setup  
499 of the dragon fruit cacti grafted onto the test media of tea leaves, bran, MW, and SW frass in  
500 reused plastic wineglasses. The funnel end allowed the grafted cacti to be covered by less frass  
501 and also stay upright.



504

505 **Figure 2** Average rate of PS consumption (mg / g of worm per day) by superworms (Sw) and  
 506 mealworms (Mw) with and without food additives (cinnamon, sucrose and bran). Additives  
 507 were mixed with PS balls and sprayed with DI water to allow the additives to adhere to the  
 508 styrofoam balls. The residual PS were weighed after four days. Results are reported as standard  
 509 error of means from 6 replicates, statistical analysis were performed with two tailed Student's  
 510 T-test. \* =  $p < 0.1$ , \*\* =  $p < 0.05$  versus corresponding controls of the same worm species; £  $p$   
 511  $< 0.05$  versus Bran of the same worm species (co-feeding bran had been previously been  
 512 reported to boost PS consumption); \$  $p < 0.05$  versus corresponding setup of a different worm  
 513 species.



514  
515 **Figure 3:** Mean cacti height differences grown on the respective media over a fortnight with  
516 standard error from 11 replicates. \*\* = significant changes in cacti height compared to tea  
517 leaves control ( $p < 0.05$ , two-tailed student's T-test). MW = mealworm frass, SW = superworm  
518 frass.



519

520 **Figure 4:** Dragon fruit cacti grown on frass, bran and tea leaves after a fortnight. (A)

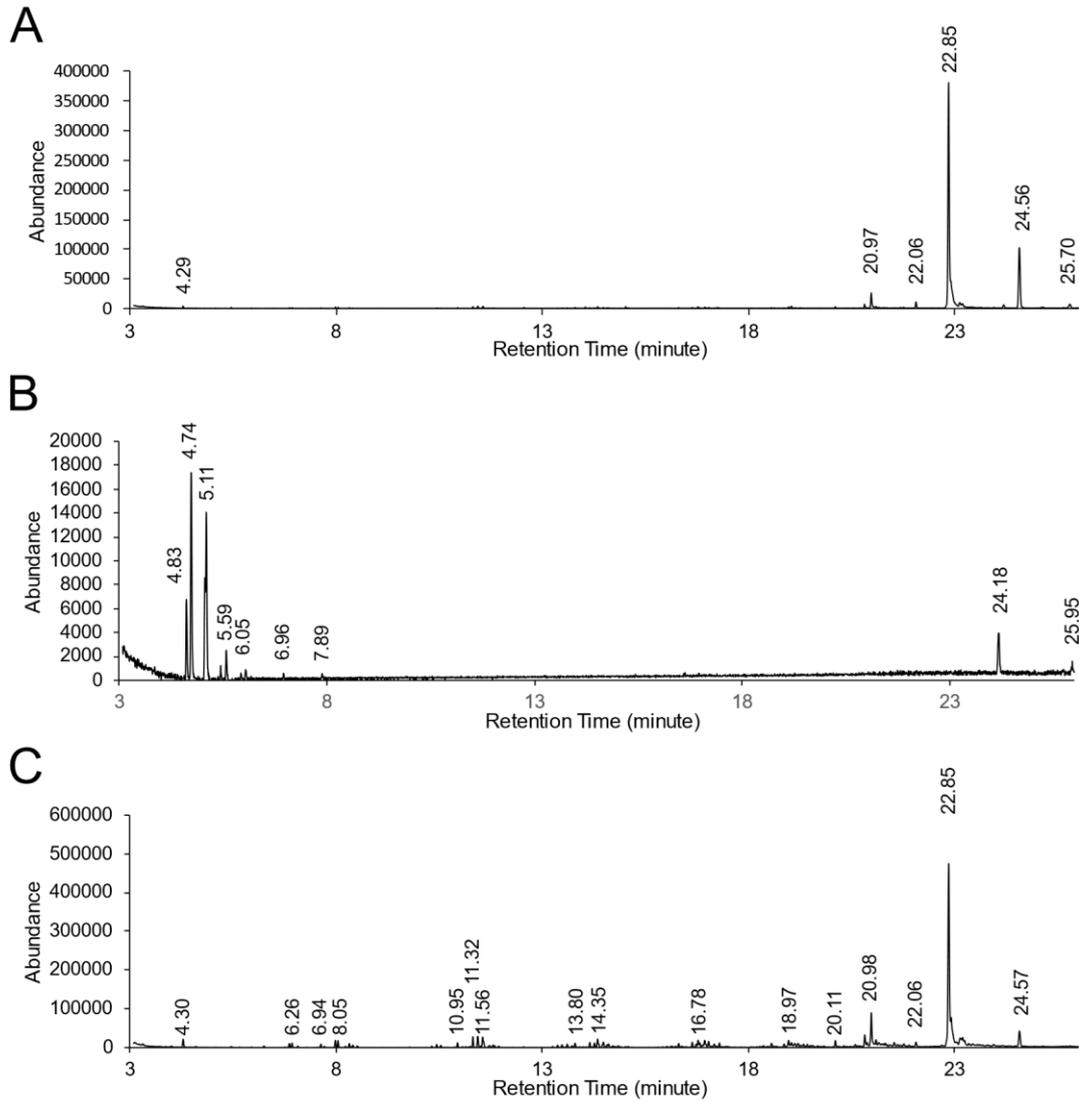
521 Representative pictures of cacti grown on tea leaves (a, b), Bran (c, d), Mealworm frass (e, f)

522 and Superworm frass (g, h). (B) Dead cacti from mealworm frass setups. Aerial roots occurring

523 above the soil line were not counted, as many of them pre-existed prior to the start of the

524

experiment.



525

526 **Figure 5:** Representative GC-MS graphs from (A) frass of PS fed superworms, (B) PS balls

527 control, and (C) frass from superworms fed on bran. A table of proposed chemicals

528 corresponding to the identities of the different peaks are provided in Table 3

529

530

531

**Table 2.** Change in worm weight after four days with different additives. Each set-up was performed in sextuplicate. Statistical analyses were performed using two-tailed Student's T-test \* =  $p < 0.1$ , with the average change in worm weight calculated in (g) and (%).

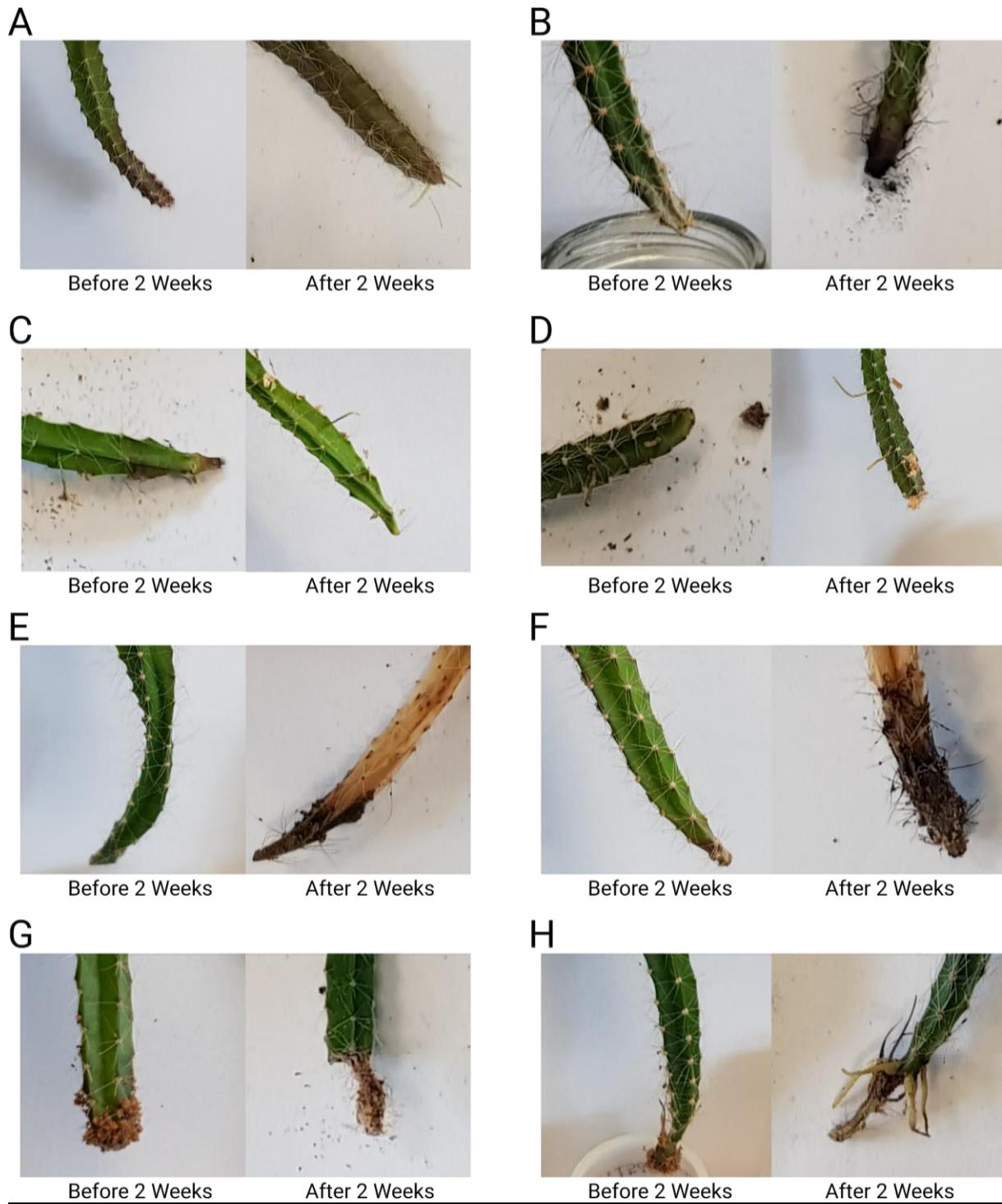
Set-Up	Initial Worm Weight (g)	Final Worm Weight (g)	Change in worm weight (g)	Average change in worm weight (g)	SEM	Average Change in worm weight (%)	t-test P value (two-Tailed)	
<b>Superworms</b>	<b>Control</b>	6.95	7.19	0.24	<-0.01	0.08	-0.04	0.97
		7.42	7.46	0.04				
		7.46	7.55	0.09				
		10.40	10.38	-0.02				
		10.70	10.67	-0.03				
		10.33	9.99	-0.34				
	<b>Cinnamon</b>	6.98	6.93	-0.05	-0.11	0.09	-1.24	0.27
		7.08	7.11	0.03				
		6.77	6.97	0.20				
		10.66	10.48	-0.18				
		10.40	10.14	-0.26				
		10.33	9.94	-0.39				
	<b>Sucrose</b>	7.01	7.30	0.29	0.16	0.07	1.79	0.06*
		7.40	7.73	0.33				
		6.62	6.90	0.28				
		10.55	10.53	-0.02				
		10.31	10.36	0.05				
		10.71	10.72	0.01				
	<b>Bran</b>	6.57	6.67	0.10	-0.06	0.08	-0.69	0.50
		7.26	7.13	-0.13				
		7.05	7.30	0.25				
10.10		9.99	-0.11					
10.48		10.15	-0.33					
10.39		10.25	-0.14					
<b>Mealworms</b>	<b>Control</b>	6.79	6.36	-0.43	-0.24	0.22	-2.85	0.32
		6.29	5.37	-0.92				
		6.44	5.65	-0.79				
		10.49	10.58	0.09				

	10.46	10.70	0.24				
	10.76	11.11	0.35				
<b>Cinnamon</b>	6.22	5.74	-0.48				
	6.23	5.89	-0.34				
	6.45	5.89	-0.56				
	10.54	10.63	0.09	-0.20	0.16	-2.36	0.27
	10.44	10.90	0.46				
	10.45	10.09	-0.36				
<b>Sucrose</b>	6.36	5.57	-0.79				
	6.45	4.77	-1.68				
	6.34	5.87	-0.47				
	10.56	10.34	-0.22	-0.46	0.29	-5.43	0.18
	10.59	10.71	0.12				
	10.52	10.80	0.28				
<b>Bran</b>	6.40	5.84	-0.56				
	6.11	5.76	-0.35				
	6.59	6.20	-0.39				
	10.69	10.84	0.15	-0.17	0.12	-1.98	0.23
	10.75	10.89	0.14				
	10.50	10.50	0.00				

**Table 3 List** of individual chemicals from a mass search of the peaks detected in GC-MS.

	PK	RT (mins)	Library search results
Frass from Superworms reared on PS	1	4.29	2,4-Dimethyl-1-heptene
	2	11.44	1-Octadecanol
	3	19.04	Pentanamide, 4-methyl-
	4	20.81	Cyclopentylacetone 2-Propanone
	5	20.97	Hexadecanamide
	6	22.06	Hexacosane
	7	22.85	9-Octadecenamide, (Z)-
	8	23.12	9-Octadecenamide, (Z)-
	9	23.18	1-Heptadecanamine
	10	24.19	Benzonitrile, m-phenethyl-
	11	24.56	Pentacosane
	12	25.79	Tricosane, 2-methyl-
PS Balls	1	4.63	Benzene, ethyl
	3	5.07	1,3,5,7-Cyclooctatetraene
	4	5.1	3-Hexene, (E)-
	7	6.05	Benzene, propyl-
	9	25.95	Vanadium, ( $\eta^7$ -cycloheptatrienylum)( $\eta^5$ -2,4-cyclopentadien-1-yl)-
Frass from Superworms reared on Bran	1	4.3	2,4-Dimethyl-1-heptene
	2	7.99	Heptane, 4-methylene-
	3	8.05	1-Hexadecanol
	4	11.32	1,1-dimethyl-2-propylcyclohexane
	5	11.44	4-Decene, 3-methyl-
	6	11.56	Cyclohexane, 1,2,3-trimethyl-
	7	14.35	Heneicosane
	8	14.48	Ethanone, 1-cyclopentyl-
	9	16.64	2-Undecene, 4,5-dimethyl
	10	16.78	Pentacosane
	11	16.94	Trifluoroacetyl-3,7-dimethyloctano
	12	17.04	Cyclohexane, 1,2,4-trimethyl-
	13	18.98	Heneicosane
	14	20.11	cyano-8-pentadecene
	15	20.82	9-Octadecenamide
	16	20.98	Hexadecanamide
	17	21.09	Nonahexacontanoic acid
	18	22.86	9-Octadecenamide,
	19	22.91	9-Octadecenamide,
	20	23.13	9-Octadecenamide,
	21	23.19	Tetracosane

	22	24.57	Pentacosane
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538 **Figure S1:** Representative pictures of cacti before and after 2 weeks of growth on tea leaves  
539 (A, B), Bran (C, D), Mealworm frass (E, F) and Superworm frass (G, H) based media. “Before  
540 2 weeks” refer to the start of the experiment, and “After 2 weeks” refers to the end of the  
541 experiment.

**Table S1.** Effect of different media on mean change in height of the cacti. Each set-up was performed with eleven replicates. Statistical analyses were performed using the two-tailed Student's T-test \*\* =  $p < 0.05$ .

Plant Media	Initial Height (cm)	Final Height (cm)	Change in Height (cm)	Mean Change in Height (cm)	SEM	Control	t-test <i>P</i> value (2-tailed)
<b>Tea leaves</b>	9.4	9.5	0.1	0.14	0.18	Bran	.32
	13.0	13.5	0.5				
	10.4	10.6	0.2				
	7.9	6.7	0.8				
	12.6	11.6	-1.0				
	15.8	15.5	-0.3				
	10.0	10.0	0.0				
	10.9	10.6	-0.3				
	8.2	8.0	-0.2				
	9.0	10.1	1.1				
	20.1	20.7	0.6				
<b>Bran</b>	7.6	8.0	0.4	-0.52	0.60	Tea leaves	.32
	10.9	11.5	0.6				
	6.9	6.7	-0.2				
	10.3	10.8	0.5				
	6.1	5.8	-0.3				
	10.1	10.1	0.0				
	8.5	8.5	0.0				
	12.3	12.2	-0.1				
	7.0	6.9	-0.1				
	8.1	8.1	0.0				
	17.2	10.7	-6.5				
<b>Mealworm Frass</b>	8.1	7.6	-0.5	-0.43	0.06	Tea leaves	.01**
	7.5	7.3	-0.2				

	6.1	5.4	-0.7			
	7.7	7.1	-0.6			
	9.9	9.6	-0.3			
	11.9	11.6	-0.3			
	13.1		Dead			
	12.1		Dead			
	7.1		Dead			
	9.5		Dead			
	6.2		Dead			
	10.4	12.5	2.1			
	11.2	13.0	1.8			
	12.3	13.0	0.7			
	7.6	7.7	0.1			
	9.1	10.5	1.4			
<b>Superworm Frass</b>	11	9.4	-1.6	0.50	0.33	
	18.1	18.7	0.6			
	10.8	10.8	0.0			
	5.9		Dead			
	8.2	7.9	-0.3			
	7.3	7.5	0.2			

Note. \* denotes  $P < .05$ . 5 out of 11 cactus plant replicates with mealworm frass died. 1 out of 11 cactus plant replicates with superworm frass died