

Mitochondrial genetics

Patrick Francis Chinnery and Gavin Hudson*

Institute of Genetic Medicine, International Centre for Life, Newcastle University, Central Parkway, Newcastle upon Tyne NE1 3BZ, UK

Introduction: In the last 10 years the field of mitochondrial genetics has widened, shifting the focus from rare sporadic, metabolic disease to the effects of mitochondrial DNA (mtDNA) variation in a growing spectrum of human disease. The aim of this review is to guide the reader through some key concepts regarding mitochondria before introducing both classic and emerging mitochondrial disorders.

Sources of data: In this article, a review of the current mitochondrial genetics literature was conducted using PubMed (<http://www.ncbi.nlm.nih.gov/pubmed/>). In addition, this review makes use of a growing number of publically available databases including MITOMAP, a human mitochondrial genome database (www.mitomap.org), the Human DNA polymerase Gamma Mutation Database (<http://tools.niehs.nih.gov/polg/>) and PhyloTree.org (www.phylotree.org), a repository of global mtDNA variation.

Areas of agreement: The disruption in cellular energy, resulting from defects in mtDNA or defects in the nuclear-encoded genes responsible for mitochondrial maintenance, manifests in a growing number of human diseases.

Areas of controversy: The exact mechanisms which govern the inheritance of mtDNA are hotly debated.

Growing points: Although still in the early stages, the development of *in vitro* genetic manipulation could see an end to the inheritance of the most severe mtDNA disease.

Keywords: mitochondria/genetics/mitochondrial DNA/mitochondrial disease/mtDNA

Accepted: April 16, 2013

Mitochondria

*Correspondence address.
Institute of Genetic
Medicine, International
Centre for Life, Newcastle
University, Central
Parkway, Newcastle upon
Tyne NE1 3BZ, UK. E-mail:
gavin.hudson@ncl.ac.uk

The mitochondrion is a highly specialized organelle, present in almost all eukaryotic cells and principally charged with the production of cellular energy through oxidative phosphorylation (OXPHOS). In addition to energy production, mitochondria are also key components in calcium signalling, regulation of cellular metabolism, haem synthesis, steroid synthesis and, perhaps most importantly, programmed cell death (apoptosis).¹

Structure

MtDNA contains 37 genes, 28 on the H-strand and 9 on the L-strand. Thirteen of the genes encode one polypeptide component of the mitochondrial respiratory chain (RC), the site of cellular energy production through OXPHOS. Twenty-four genes encode a mature RNA product: 22 mitochondrial tRNA molecules, a 16 s rRNA (large ribosomal subunit) and a 12 s rRNA (small ribosomal subunit).⁵ Unlike its nDNA counterpart, mtDNA is extremely efficient with ~93% representing a coding region. Unlike nDNA, mtDNA genes lack intronic regions and some genes, notably *MTATP6* and *MTATP8*, have overlapping regions. Most genes are contiguous, separated by one or two non-coding base pairs. mtDNA contains only one significant non-coding region, the displacement loop (D-loop).⁵ The D-loop contains the site of mtDNA replication initiation (origin of heavy strand synthesis, OH) and is also the site of both H-strand transcription promoters (HSP1 and HSP2).

The mitochondrial genetic code differs slightly from nuclear DNA (nDNA). MtDNA uses only two stop codons: 'AGA' and 'AGG'⁶ (compared with 'UAA', 'UGA' and 'UAG' in nDNA), conversely 'UGA' encodes tryptophan. To compensate UAA codons have to be introduced at the post-transcriptional level. In addition 'AUA', isoleucine in nDNA, encodes for methionine in mtDNA.

Inheritance

Prevailing theory suggests that mtDNA is maternally inherited, with mtDNA nucleoids the unit of inheritance. During mammalian zygote formation, sperm mtDNA is removed by ubiquitination, likely occurring during transport through the male reproductive tract.⁷ Consequently, the mtDNA content of the zygote is determined exclusively by the previously unfertilized egg.

To date only a single case of paternal transmission in humans has been recorded.⁸ However, paternal transmission in other animals is both common and recurring. Theory suggests that the lack of paternal inheritance is due to either (i) a dilution effect; sperm contain only 100 copies of mtDNA, compared with 100 000 in the unfertilized egg, (ii) selective ubiquitination of paternal mtDNA or (iii) the 'mtDNA bottleneck' excludes the 'minor' paternal alleles.⁷ The advent of deep, next generation sequencing, allowing mtDNA can be sequenced at great depths (>20 000 fold) may enable researchers to revisit this phenomenon.

Homoplasmy and heteroplasmy

Cells contain thousands of molecules of mtDNA;⁹ and in the majority of cases their sequence is identical, known as homoplasmy. However, an

inefficient mtDNA repair, a localized oxidative environment and increased replication¹⁰ make mtDNA mutation frequent. The polyploid nature of mtDNA means that mutations often co-exist with their wild-type counterpart in various proportions (termed heteroplasmy). The proportion of mutant has important consequences in understanding mitochondrial disease (discussed later).¹¹

nDNA and mitochondrial function

According to recent data the mitochondrial proteome is estimated at ~1500 proteins.¹² Mitochondria are dependent upon the nuclear genome for the majority of the OXPHOS system and also for maintaining and replicating mtDNA as well as organelle network proliferation and destruction (Fig. 2).

OXPHOS system

To date, 92 structural OXPHOS subunit genes have been identified: 13 encoded by mtDNA (Fig. 1) and 79 encoded by the nuclear genome. Briefly, complex I (NADH:ubiquinone oxidoreductase), the largest of the RC components, consists of 44 subunits: 14 enzymatic ‘core subunits’ (7 from mtDNA and 7 from nDNA)¹³ and a further 30 nDNA accessory subunits thought to maintain complex stability.¹⁴ Complex II (succinate:

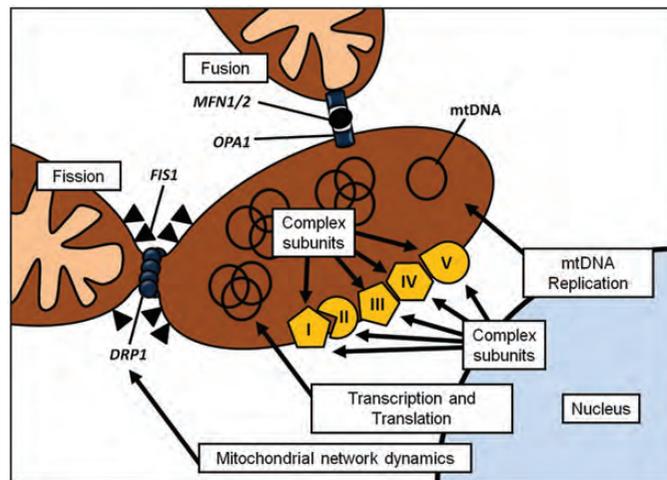


Fig. 2 Interaction between nDNA and mtDNA. Cartoon demonstrating the complex interaction between genes encoded by nDNA and the processes they control in the mitochondrion.

ubiquinone oxidoreductase) is encoded entirely by nDNA (four subunits). Complex III (ubiquinol:cytochrome *c* oxidoreductase) contains 11 subunits, 1 encoded by mtDNA (*MTCYB*) and 10 encoded by nDNA.¹⁵ Complex IV (cytochrome *c* oxidase) consists of three mtDNA-encoded subunits and a further 11 nDNA-encoded subunits. Finally, complex V (F₀F₁-ATP synthase) comprises 19 subunits, 2 encoded by mtDNA and the remaining 17 encoded by nDNA.

In addition, nDNA encodes over 35 proteins required for the RC assembly: complex I = 11 nDNA assembly factors,¹⁶ complex III = 2,¹⁵ complex IV = 18¹⁷ and complex V = 4.¹⁸

mtDNA replication

Unlike nDNA, mtDNA replication is not governed by the cell cycle (eukaryotic cell division) and is continuously recycled. MtDNA replication and integrity maintenance is handled entirely by the nDNA. In eukaryotes, mtDNA is replicated in a 'replisome' (a DNA/protein complex making up the replication machinery) by a trimeric protein complex composed of a catalytic subunit: polymerase gamma, a 140 kDa DNA polymerase encoded by *POLG* and two 55 kDa accessory subunits, encoded by *POLG2*.¹⁹ This enzyme complex performs three activities, DNA polymerase activity, 3'-5' exonuclease/proofreading activity and a 5'dRP lyase activity (required for enzymatic DNA repair).

In addition, the replisome also includes the mitochondrial single-stranded binding protein (encoded by *mtSSB*), which is involved in stabilizing single-stranded regions of mtDNA at replication forks, enhancing polymerase gamma activity. Twinkle is a 5'-3' DNA helicase, which unwinds double-stranded mtDNA, facilitating mtDNA synthesis, as well as acting as a mtDNA primase (an enzyme required to prime nucleotide synthesis).¹⁹ Several topoisomerases have been identified in humans, including the mitochondrial topoisomerases 1 (encoded by *TOP1mt*) and III α (encoded by *TOP3a*). Finally, the synergy between mitochondrial transcription factor A (encoded by *TFAM*) and mtDNA copy number suggests that TFAM may act as an mtDNA chaperone (a protein that assists the function of another protein) protecting it against oxidative damage.

mtDNA arrangement

Like its nDNA counterpart, mtDNA is also packaged in protein-DNA complexes, known as nucleoids.²⁰ MtDNA nucleoids are associated with the inner mitochondrial membrane, spaced evenly along the cristae. In addition to a single mtDNA molecule,²¹ mtDNA nucleoids contain a number of proteins.²⁰ Principally the site of mtDNA replication, it is

unsurprising that mtDNA nucleoids contain the protein machinery required for DNA replication, transcription, repair and packaging, including the mtDNA polymerase *POLG*, its accessory subunit *POLG2*, the activator of mtDNA transcription (encoded by *TFAM*) as well as mtDNA helicases and binding proteins (*twinkle* and *mtSSB*, respectively).²⁰ In addition, mtDNA nucleoids contain chaperone proteins (HSP90- β and HSP70) required for mtDNA stability.

Transcription and translation

Transcription of mtDNA is 'prokaryotic like' and was thought of a two-component system involving a protein complex containing the mitochondrial RNA polymerase (*POLRMT*) and two transcription factors (TFB1M and 2M).^{22,23} However, recent research indicates that TFB1M does not modulate mtDNA transcription in the presence of TFB2M, rather it acts as a dimethyltransferase which stabilizes the small subunit of the mitochondrial ribosome. RNA transcription is regulated by mitochondrial transcription factor A (*TFAM*).²⁴

Briefly, each strand is transcribed as a polycistronic precursor mRNA molecule (i.e. the mRNA contains all of the genes in one molecule). Light-strand transcription is initiated from the light-strand promoter; however, heavy-strand transcription initiates from two heavy strand promoters: HSP1 and HSP2 (Fig. 1).²⁵ Transcript elongation is performed by *POLRMT*, enhanced by both 'transcription elongation factor mitochondrial' (*TEFM*) and termination of mature transcripts is carried out by mitochondrial termination factor 1 (*MTERF1*).²⁵

Translation of the 13 mtDNA protein coding genes occurs in the mitochondria. The mitoribosomes are partly coded by mtDNA (*MTRNR1* and *MTRNR2*, Fig. 1), but require a further 81 nDNA proteins. Translation is initiated by two mitochondrial initiation factors: mtIF1 and mtIF3.^{26,27} mtIF3 begins initiation by dissociating the 'mitoribosome' (the mitochondrial ribosomes) allowing assembly of the initiation complex.²⁸ mRNA is then bound to the small subunit, aligning the start codon to the peptidyl site of the mitoribosome. Peptide elongation is controlled by a number of nuclear-encoded genes, including mitochondrial elongation factor Tu (mtEFTu),^{29,30} which binds the tRNA to the mitoribosome and mitochondrial elongation factor G1 (mtEFG1), required to move the newly added amino acid along one position and allowing amino acid inclusion.³¹ Translation termination is carried out solely by mitochondrial release factor 1a (mtRF1a),³² which recognizes the stop codons (UAA and UAG)³³ and triggers hydrolysis of the bond between the terminal tRNA and the nascent peptide.

Controlling mitochondrial network dynamics

Mitochondria are often depicted as distinct organelles; however, they actually form a complex reticulum that is undergoing continual fusion and fission (Fig. 2).³⁴ It is likely that fusion has evolved as a mechanism to promote intermitochondrial cooperation—allowing the sharing and dissemination of mtDNA and mitochondrial proteins. Fission promotes mitochondrial compartmentalization,³⁴ a mechanism that is needed to distribute mitochondria during cell division. Mitochondrial network dynamics, much like mtDNA replication, is controlled completely by nDNA, although likely involves mtDNA–nDNA communication.³⁴

Mitochondrial fusion

The principle player in mitochondrial fusion is mitofusin (Mfn) and mammalian mitochondria contain two similar mitofusin proteins: Mfn1 and Mfn2 (Fig. 2),³⁴ sharing 80% sequence homology. Studies indicate that both Mfn1 and Mfn2 uniformly localize to the mitochondrial outer membrane and overexpression leads to peri-nuclear clustering on mitochondria.³⁴ Mitochondrial fusion is also dependent upon OPA1 expression (Fig. 2),³⁴ where inhibition of gene expression causes an increase in mitochondrial fragmentation, conversely the overexpression of *OPA1* breaks the network into spheres.

Mitochondrial fission

DNM1L, dynamin 1 like, controls mitochondrial fission in mammalian cells (Fig. 2).³⁴ *DNM1L* codes for a principally cytosolic protein; however, it also localizes to fission sites on the mitochondria. Similar to Mfn1, the overexpression of ‘mutant’ *DNM1L* results in a breakdown of mitochondrial networks. Due to its dynamin similarity, two different functions have been proposed for *DNM1L*. It has been hypothesized that *DNM1L* may mechanically mediate membrane fission through GTP hydrolysis; alternatively, it may act as a signalling molecule, conscripting and activating separate fission enzymes such as Dnm1: the yeast homologue of Drp1.

Areas of agreement

Mitochondrial disease

An area where all mitochondrial researchers would agree is the capacity for mitochondrial dysfunction to manifest as disease. Mitochondrial disease is principally a chronic loss of cellular energy, where a failure to meet cellular energy demand results in a clinical phenotype. The clinical

spectrum of mitochondrial disease is diverse (Fig. 3); however, tissues where there is a high metabolic demand, such as the central nervous system (CNS) or heart, are typically affected.

The broad clinical spectrum of mitochondrial dysfunction, coupled with the heterogeneity of mtDNA variation, makes the prevalence of mitochondrial DNA (mtDNA) difficult to calculate. Estimates, based on clinical observations, indicate that as many as 1 in 5000 people in the North East of England have manifested mitochondrial disease,³⁵ with similar figures reported in other parts of the world.^{36–38}

Identifying and diagnosing mitochondrial genetic disease: general principles

Mitochondrial dysfunction should be considered in the differential diagnosis of any progressive, multisystem, disorder. However, clinical diagnosis can be difficult if patients do not present with ‘classical mitochondrial’ disease (see later).

A detailed family history is important; a clear maternal inheritance (without male transmission) indicates a primary mtDNA defect, whilst an autosomal inheritance pattern is indicative of nDNA interaction. In many

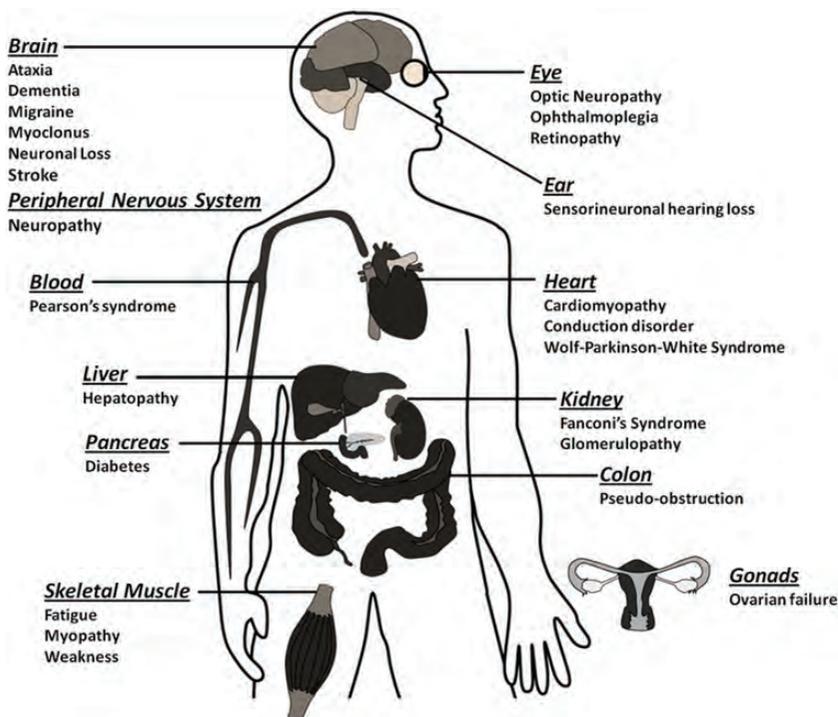


Fig. 3 Clinical spectrum of mitochondrial disease. Schematic diagram showing the organ and corresponding disease affected by mitochondrial dysfunction.

cases blood and/or CSF lactate concentration,³⁹ neuroimaging,^{40,41} cardiac evaluation and muscle biopsy for histological or histochemical evidence can indicate mitochondrial disease. However, establishing a molecular genetic diagnosis is preferred.

Molecular genetic testing can be carried out on DNA extracted from blood (useful for the identification of some mtDNA and nDNA mutations),^{42,43} but DNA extracted from the affected tissue is preferred (as pathogenic mtDNA mutations are often not detectable in blood).⁴⁴ Southern blot analysis can be used to identify mtDNA rearrangements and 'common' mutations can be targeted by Sanger sequencing of either mtDNA or nDNA.

The genetics of mitochondrial disease

The complex interaction between the two cellular genomes means mitochondrial disease can arise through either (i) a primary mtDNA defect or (ii) a defect in a nuclear-encoded mitochondrial protein.

mtDNA and disease

Understanding mtDNA variation

mtDNA integrity is constantly attacked by mitochondrial reactive oxygen species (ROS) generated during cellular OXPHOS.⁴⁵ ROS are potent genotoxic agents, which cause mutagenic and cytotoxic effects. The proximity of mtDNA to the site of mitochondrial ROS production (principally complexes I and III of the RC) is the major cause of oxidative lesions and mtDNA instability and is directly responsible for the higher nucleotide instability when compared with nDNA.

Despite being packaged in mitochondrial nucleoids and possessing DNA repair pathways evolved to cope with oxidative damage, including base excision repair mechanisms,⁴⁶ mtDNA mutation rates are significantly higher than nDNA. Mutation creates two distinct classes of mtDNA variant: (i) single-base-pair variants and (ii) mtDNA rearrangements (deletions and insertions). Single-base-pair variants are typically inheritable and are either common in the populace (as proposed neutral variants) or enriched in individuals with disease (as mtDNA mutations). Understanding the complex nature of mtDNA variation is critical to understanding its affect on disease and there are a few key points that must be understood before assessing an mtDNA variant.

Consequences of mtDNA heteroplasmy

MtDNA heteroplasmy (described earlier) has a complex relationship with disease. The clinical expression of a heteroplasmic pathogenic

mtDNA mutation is directly correlatable with the relative proportion of wild-type and mutant genomes.⁴⁷ For common point mutations, a typical threshold of 80–90% mutant is required to manifest as disease at the cellular level,^{48,49} and tissue levels correlate loosely with the severity of the clinical phenotype. However, there is emerging evidence that mutation levels can change over time, increasing in post-mitotic tissues, such as brain and muscle and decreasing in mitotic tissues including blood. This can present a challenge when interpreting some clinical molecular genetic tests.^{44,50,51}

Common mtDNA variation

Evolutionarily, common inherited mtDNA mutations have created stable population subgroups separated by common sequence variation known as haplogroups. Many of the major sub-divisions occurred over 10 000 years ago, developing as humans migrated into new geographic areas. Over 95% of Europeans belong to 1 of 10 major haplogroups, H, J, T, U, K (a subgroup of U), M, I, V, W and X, with each haplogroup defined by specific sequence variants within the population.⁵² These common, inherited, mtDNA variants are usually not heteroplasmic, and due to their selection neutrality have become fixed in the population. However, different haplogroups have been associated with a variety of human diseases, including primary mitochondrial disorders such as Leber's hereditary optic neuropathy (LHON, an age-related loss of vision), where background mitochondrial haplogroup has a direct, functional, effect on the RC protein complex assembly;⁵³ but has expanded to include age-related neurodegenerative disorders such as Parkinson's disease (PD)⁵⁴ Alzheimer's disease^{55,56} and age-related macular degeneration.⁵⁷

Rare mtDNA variation

Rare, inherited, point mutations are a major cause of disease in humans, with an estimated incidence of 1 in 5000.⁵⁸ They primarily occur in protein coding and tRNA genes and ultimately result in a reduction of cellular energy, through either a reduction in mitochondrial RC enzyme activity or an impairment of mitochondrial protein synthesis.⁵⁹ Unlike common inherited variants, rare point mutations are often heteroplasmic.

In contrast to point mutations, primary mitochondrial rearrangements of mtDNA are not inheritable; they are primarily, sporadic, large-scale deletions, typically heteroplasmic and usually result in disease. To date around 120 different mtDNA deletions have been identified in patients with mitochondrial disease.⁶⁰ Similarly to mtDNA point mutations, the ratio of deleted versus 'wild-type' molecules is critical to disease aetiology, with mtDNA deletions manifesting disease at a lower heteroplasmic threshold (~50–60%).⁶¹ The exact mechanism of deletion formation is under debate and current research indicates two likely

models of deletion formation: (i) replication error and (ii) mtDNA repair inefficiency.^{62,63}

'Classical' mtDNA diseases

LHON is a common cause of inherited blindness that typically presents with bilateral, painless, sub-acute visual failure in young adult males. LHON was the first maternally inherited disease to be associated with an mtDNA point mutation.⁶⁴ Today, clinical diagnosis is usually confirmed by molecular genetic analysis for one of three 'common' mtDNA mutations, which all affect genes coding for complex I subunits of the RC: m.3460G>A, m.11778G>A and m14484T>C.⁶⁵ Mitochondrial dysfunction causes a specific loss of retinal ganglion cells,⁶⁶ whilst preserving the remaining retinal layers. The optic nerve also shows characteristic degeneration and an accumulation of mitochondria suggesting an impairment of axoplasmic transport. LHON mutations are typically homoplasmic; however, not all patients harbouring a pathogenic LHON mtDNA mutation develop visual failure. Studies of LHON have identified common mtDNA variants that may modulate LHON expression;^{67,68} additionally environmental factors, such as cigarette smoke⁶⁹ and oestrogen levels may play a role.⁷⁰ However, the majority of research has focused on the identification of a nuclear-encoded susceptibility allele.^{67,71-74}

Non-syndromic and aminoglycoside-induced sensorineuronal hearing loss is associated with m.1555A>G, a homoplasmic point mutation in the 12sRNA gene.⁷⁵ The variant alters a highly conserved region of 12sRNA, mutating the molecule to more closely resemble its bacterial homologue. *In vitro* experiments on m.1555A>G mutant cell lines demonstrated that exposure to aminoglycoside would impair growth; however, not all symptomatic individuals have been exposed to aminoglycoside.⁷⁵

Surprisingly, given that they make up only 5% of mtDNA, the vast majority of pathogenic mtDNA point mutations occur in the tRNA genes (Fig. 1).^{76,77} In addition, pathogenic tRNA mutations are typically heteroplasmic.

Mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes (MELAS) is typically a childhood, multisystem disorder. Patients commonly manifest with generalized tonic-clonic seizures, recurrent headaches, anorexia with recurrent vomiting and postlingual hearing loss,⁷⁸⁻⁸⁰ but can manifest with impaired: motor ability, vision and mental acuity due to the cumulative effect of multiple stroke-like episodes. MELAS is commonly (80% of cases) caused by a A>G transition at m.3243 in *MTTL1*,⁸¹ but is also associated with variants in *MTND5*.⁸² Biochemically, MELAS manifests as defects of complex I and IV activity; however, care must be taken when interpreting the findings as biochemical results can often appear normal.

Myoclonus epilepsy with ragged red fibres (MERRF) is a neuromuscular disorder primarily caused by m.8344A>G in *MTTK*.⁸³ Clinically, patients with m.8344A>G present with myoclonus, epilepsy, muscle weakness, cerebellar ataxia and dementia, although neurological symptoms can develop with age.⁸³ Clinical severity is correlated with patient heteroplasmy with high levels of mutant mtDNA often causing, severe complex I or IV deficiency and occasionally a combined complex I and IV deficiency. Much like MELAS, the genotype–phenotype correlation of m.8344A>G can be extended beyond MERRF. M.8344A>G has been identified in diverse mitochondrial phenotypes such as Leigh’s syndrome.

m.7472insC, affecting *MTTS* (Fig. 1), was first identified in a large Italian family presenting with hearing loss, ataxia and myoclonus. This mutation was later found in several unrelated families, all showing a wide clinical spectrum, including isolated hearing loss, ataxia and MERRF. This mutation has been found at increasing frequencies in families presenting with maternally inherited hearing loss.

Pathogenic rearrangements of mtDNA are typically large-scale deletions and to date over 120 different pathogenic mtDNA deletions have been identified.⁶⁰ As described previously, mtDNA deletions are typically sporadic and not inheritable. Clinical severity is directly correlatable with the level and tissue distribution of the rearrangement and mitochondrial dysfunction is simply a result of the removal of key mitochondrial genes. Homoplasmic tRNA gene loss is particularly detrimental as mitochondria cannot synthesize a functional OXPHOS system. mtDNA deletions are associated with three main clinical phenotypes: Kearns–Sayre syndrome (KSS),⁸⁴ sporadic progressive external ophthalmoplegia (PEO)⁸⁵ and Pearson’s syndrome.⁸⁶

KSS is an early onset, sporadic, disorder characterized by PEO and pigmentary retinopathy; however, cases can also present with cerebellar syndrome, heart block, diabetes and shortness of stature. Mitochondrial dysfunction manifests as ragged red fibres (RRFs), an accumulation of dysfunctional mitochondria in the sub-sarcolemmal region of a muscle fibre (detectable when a muscle section is stained with Gomori trichrome stain).⁸⁵

Large-scale deletions and duplications of mtDNA are a known cause of Pearson’s bone-marrow–pancreas syndrome, a rare infant disorder characterized by infantile sideroblastic anaemia and occasionally including severe exocrine pancreatic insufficiency.⁸⁶

mtDNA variation and mitochondrial disease

Nuclear–mitochondrial disease can be classified into four distinct groups: (i) disorders resulting from a reduction in mtDNA stability; (ii) disorders

resulting from mutations in nuclear-encoded components or assembly factors of the OXPHOS system; (iii) disorders resulting from mutations affecting mitochondrial translation and (iv) disorders due to defects in genes controlling mitochondrial network dynamics.

Disorders resulting from a reduction in mtDNA stability

A growing number of disorders have become associated with mtDNA instability, primarily a result of impaired mtDNA replication. Mutations in *POLG*, the gene encoding the only mtDNA polymerase, are by far the commonest cause of mtDNA stability disorders. Mutations in the *POLG* gene can cause either point mutations (through impaired mtDNA proofreading) or deletions (through impaired polymerase activity) in mtDNA.¹⁹ The first pathogenic mutations in *POLG* were identified in families with autosomal dominant PEO (adPEO); however, the spectrum of disease associated with *POLG* mutations has been expanded to include autosomal recessive PEO, adult onset ataxia, Alpers' syndrome, parkinsonism and premature ovarian failure.⁸⁷

adPEO, characterized by multiple mtDNA deletions, is caused by mutations in *PEO1*, which encodes 'twinkle' the putative mitochondrial helicase.⁸⁸ It is thought that twinkle mutations result in an accumulation of replication intermediates, causing replication stalling and eventually depletion. adPEO is also associated with mutations in *ANT1*,⁸⁹ the gene coding adenine nucleotide translocase. Mutations in *ANT1* impair ADP-ATP exchange through the mitochondrial membrane, causing a nucleotide imbalance (affecting replication) and a severe reduction in cellular energy.

In addition to structurally altering mtDNA, several disorders have been identified that are caused by a reduction in mtDNA copy number.¹⁹ Alpers syndrome, characterized by diffuse and progressive cerebral atrophy,⁹⁰ has been associated with mutations in *POLG*,^{91,92} which cause impairment of the replicative machinery.⁹³

Recessive mutations in thymidine phosphorylase cause mitochondrial neurogastrointestinal encephalopathy, characterized by mtDNA depletion, multiple deletions and point mutations. mtDNA depletion has also been identified in early onset hypotonia with myopathy and hepatic involvement, caused by mutations in either thymidine kinase (*TK2*) or deoxyguanosine kinase (*DGUOK*).⁹⁴ Mutations in both of these genes cause a reduction in the mtDNA nucleotide pooling, reducing replication efficiency.

Disorders resulting from mutations in nuclear-encoded components or assembly factors of the OXPHOS system

Isolated complex I deficiency is by far the commonest biochemical defect found in mitochondrial disorders; however, it is also the most complex

aetiology and clinical spectrum.⁹⁵ Complex I deficiency is associated with a broad range of clinical phenotypes ranging from lethal neonatal disease to adult onset neurodegenerative disorders.^{96,97} A high level of genetic heterogeneity, coupled with weak genotype–phenotype correlations, makes it difficult to predict the genetic basis on pure clinical grounds.⁹⁵ This is important because of the different inheritance patterns and different natural histories of the different genetic causes. However, some patterns are starting to emerge.

There are at least 46 nuclear-encoded subunits of complex I (compared with 7 mtDNA encoded subunits) and so it is unsurprising that nDNA mutations have been identified in 14 of the structural subunits. Pathogenic mutations in *NDUFS1*,⁹⁸ *NDUFS3*,^{95,99} *NDUFS4*,¹⁰⁰ *NDUFS7*,¹⁰¹ *NDUFS8*,¹⁰² *NDUFV1*,^{98,103} *NDUFA10*,¹⁰⁴ *NDUFB3*⁹⁵ and *NDUFA2*¹⁰⁵ typically manifest as Leigh or Leigh-like syndromes.^{60,106} Conversely, mutations in *NDUFS2*,¹⁰⁷ *NDUFS6*,¹⁰⁸ *NDUFV2*,¹⁰⁹ *NDUFA1*, *NDUFA11*¹¹⁰ and *ACAD9*¹¹¹ are typically associated with hypertrophic cardiomyopathy and encephalopathy. In addition, mutations in complex I assembly proteins can manifest as disease: Leigh syndrome (*NDUFAF2* and *NDUFAF5*),^{112,113} encephalopathy (*NDUFAF4*)¹¹⁴ and cardioencephalomyopathy (*NDUFAF1*).¹¹⁵

Complex II is completely encoded by nDNA and is composed of four polypeptide subunits: *SHD-A*, *-B*, *-C* and *-D*. Mutations in *SHD-A* are rare, but are associated with Leigh's syndrome. Surprisingly, mutations in *SHD-B*, *-C* and *-D* appear to be a common cause of inherited paragangliomas and pheochromocytomas.¹¹⁶

Complex III deficiency typically causes a severe multisystem early onset disorder, which is recessively inherited and rare.^{117,118} Identified mutations in *BCS1L*, a complex III assembly protein, presenting with neonatal proximal tubulopathy, hepatic involvement and encephalopathy. Subsequently, a deletion in human ubiquinone–cytochrome *c* reductase binding protein of complex III (*UQCRB*) was identified in a consanguineous family presenting with hypoglycaemia and lactic acidosis;¹¹⁹ and a missense mutation was identified in *UQCRC*, a ubiquinone-binding protein, in a large consanguineous Israeli-Bedouin kindred.¹²⁰ More recently, a mutation in *TTC19* (a complex III structural subunit gene) was identified in individuals with a progressive neurodegenerative disorder in late infancy,¹²¹ expanding the phenotype of complex mutations beyond early infant disorders.

Mutations in complex IV result in severe, typically fatal, infantile disease and to date mutations in four complex IV structural subunits have been identified. A homozygous mutation in *COX6BI*, identified in brothers from a consanguineous Saudi Arabian family, presented with gait instabilities visual disturbances, progressive neurological deterioration and leukodystrophic brain changes.¹²² Mutations in *COX10*, a homologue of yeast haem A:farnesyltransferase, are associated with Leigh syndrome^{123,124} and

a multisystem disorder.¹²³ Atypically, mutations in *COX7B*¹²⁵ are associated with facial dysmorphisms and congenital abnormalities,¹²⁶ and a single mutation in the structural subunit gene, *COX4I2*, was identified in adult Arab Muslim patients with exocrine pancreatic insufficiency, dyserythropoietic anaemia and calvarial hyperostosis.¹²⁷

In contrast, a number of mutations have been identified in complex IV assembly factors. Complex IV assembly gene disorders include *SURF1* (Surfeit locus protein 1), associated with Leigh Syndrome;^{128,129} *C12ORF62* (chromosome 12 open reading frame 62), associated with fatal, neonatal, mitochondrial IV deficiency;¹³⁰ *COA5* (cytochrome *c* oxidase assembly factor 5), associated with neonatal hypertrophic cardiomyopathy¹³¹ and *FASTKD2*, associated with cytochrome *c* oxidase-defective encephalomyopathy.¹³²

Mutations in nDNA-encoded complex V subunit genes also appear very rare. A mutation in *ATP5E* (ATP synthase, H⁺ transporting, mitochondrial F1 complex, epsilon subunit) was identified in an Austrian woman with complex V deficiency,¹³³ and a single gene defect has been identified in the complex V assembly factor gene *ATPAF2*, resulting in impaired complex V activity.¹³⁴

Disorders resulting from mutations affecting mitochondrial translation

Several nDNA mutations have been identified which influence the efficiency of mitochondrial translation. Mitochondrial ribosomal protein S16 (*MRPS16*) and mitochondrial ribosomal protein S22 (*MRPS22*) are components of the mitoribosome. Mutations in these genes are known to cause severe, infantile, lactic acidosis, developmental defects in the brain, and facial dysmorphisms (*MRPS16*) and fatal neonatal hypertrophic cardiomyopathy and kidney tubulopathy (*MRPS22*).¹³⁵

Mutations in *PUS1*, pseudouridine synthase 1, have been shown to cause myopathy, lactic acidosis and sideroblastic anaemia.¹³⁶ The mutation, in the catalytic core of the protein, is thought to disrupt the conversion of uridine to pseudouridine, required for tRNA synthesis.

Disorders due to defects in genes controlling mitochondrial network dynamics

Mutations in *OPA1* are primarily a cause of optic atrophy,⁶⁶ but additional phenotypes, such as deafness and neuromuscular disease, have also been seen. Interestingly, mutations in *OPA1* also appear to cause the formation of mtDNA deletions, indicating that Opa1 is also important to mtDNA maintenance.

Much like *OPA1*, defects in *MFN2* cause a disturbance of mtDNA maintenance through impairment of mitochondrial network dynamics.⁶⁶ Mutations in *MFN2* are typically associated with Charcot-Marie-Tooth

disease (CMT2A) and hereditary motor and sensory neuropathy (CMT with HMSN type VI).⁶⁶

DNM1L (dynamamin 1-like), another GTPase, is required for fission of mitochondria.¹³⁷ To date, only a single *DNM1L* has been identified in an infant presenting with both defective mitochondrial and peroxisomal fission.¹³⁸ The patient presented in the first days of life with severe microcephaly, abnormal brain development, optic atrophy with hyperplasia and lactic acidemia.¹³⁸

Areas of controversy?

The mitochondrial bottleneck

Mutations in mtDNA are often heteroplasmic, with severity correlating with increasing percentage of mutant. Observations indicate that the amount of a variant inherited from a heteroplasmic mother varies between offspring.^{139,140} This is important when investigating disease aetiology, as an asymptomatic mother, with a sub-clinical heteroplasmy level, can give birth to children with significantly higher levels of an mtDNA mutation.

The 'mitochondrial bottleneck theory' attempts to explain this phenomenon.¹⁴⁰ Briefly, the reduction of mtDNA during early development 'redistributes' mtDNA to daughter cells (effectively sharing mtDNA content amongst daughter cells). Oocyte maturation is associated with the rapid replication of mtDNA. This reduction-amplification leads to a purportedly random shift in mtDNA mutational load between cells. Researchers agree that the bottleneck is due to a rapid reduction in mtDNA levels during embryonic development; however, the exact mechanism of segregation is hotly debated. There are currently three leading theories of the mtDNA bottleneck mechanism:¹⁴⁰ (i) variation in heteroplasmy is due to an unequal segregation of mtDNA during cell division, (ii) variation in heteroplasmy is due to an unequal segregation of mtDNA nucleoids during cell division and (iii) variation in heteroplasmy is due to the selective replication of a specific sub-population of mtDNA.

Growing points

Assigning variant causality

Optimal mitochondrial function requires the synergistic cooperation of both mtDNA and nDNA; hence, the investigation of dysfunction requires the interrogation of both genomes. Correctly determining the pathogenicity of potential mutants (in either genome) is critical to understanding

mitochondrial disease. This underpins the genetic counselling and subsequent prenatal diagnosis of mitochondrial disorders.

Despite the complexity of both mtDNA point mutations and deletions, as well as the potential for heteroplasmy, assigning pathogenicity to mtDNA variants is analogous to nDNA mutations and is comprehensively described by DiMauro and Schon.¹⁴¹ Briefly, the mutation must be present in cases significantly more than asymptomatic controls; if heteroplasmic, the proportion of mutated mtDNA must be higher in patients compared with controls (and subsequently higher in clinically affected tissues compared with unaffected tissues). More importantly, the mutated mtDNA must segregate with defined clinical outcome (described previously). Other criteria, such as evolutionary conservation must be interpreted with care, as very rare neutral variants (so-called 'private polymorphisms') or homoplasmic changes (such as in LHON) may be wrongly miss-classified using this approach.¹⁴¹ Assigning pathogenicity to tRNA mutations is slightly more challenging; tRNA variants are common; however, a small number of tRNA mutations are responsible for a disproportionate majority of mitochondrial disease.⁷⁷ McFarland *et al.*⁷⁷ provide a comprehensive scoring system which can be used to accurately determine tRNA mutation pathogenicity.

Whole-exome sequencing (WES)¹⁴² has emerged as the preferred method for identifying Mendelian disease genes, and is proving valuable in the diagnostic evaluation of phenotypically and genetically heterogeneous disorders such as mitochondrial disease.^{95,143} Initially, candidate mutations can be identified by prioritizing known mitochondrial genes, such as the 1500 proposed in 'MitoCarta',¹⁴⁴ or Mitop2.¹⁴⁵ Secondly, WES can drive the discovery of novel mitochondrial disease genes or provide a link to previous disease genes that demonstrate an overlapping clinical phenotype.^{146–151} However, as with all new technologies, care must be taken when interpreting WES data in novel disease genes. Variants identified in poorly characterized genes will require extensive biochemical and functional laboratory analysis to assign causality. Additionally, WES is not wholly comprehensive, not capturing non-coding or regulatory regions and often failing to sequence large portions of the exome.^{142,152} However, as technology improves and bioinformatic analysis becomes streamlined, WES is likely to become a major facet in identifying nuclear genes that affect mitochondrial function.

Managing mitochondrial disease

There are limited treatment options for patients with mitochondrial diseases. The main emphasis is on disease prevention and the management of complications. Effective genetic counselling, especially given a

family history of mitochondrial disease, is crucial. However, the clinical variability, coupled with the unpredictable inheritance of a heteroplasmic 'mutant dose' (through the bottleneck), makes a definite diagnosis difficult.^{153,154}

Empiric recurrence risks are available for common homoplasmic mutations (i.e. for LHON), but genetic counselling for heteroplasmic mutations is difficult because of the genetic bottleneck (described earlier). Increased knowledge of the natural history of specific mitochondrial disorders has informed clinical practice. Particular attention to cardiac, ophthalmological and endocrine complications (especially diabetes), can lead to prompt supportive management.¹⁵⁵ However, there are no specific disease-modifying treatments at present, although some drugs show promise.¹⁵⁶

An area that has had some *in vitro* and pre-clinical success is the development of 'gene therapies'.¹⁵⁷ There are currently three strategies for applying gene therapy to mitochondrial disease: (i) the rescue of an RC defect by expression of a 'replacement' gene product from the nucleus (so-called allotopic and xenotopic expression,^{158,159} (ii) the rescue of a primary mitochondrial defect by importing 'wild-type' mtDNA into mitochondria (so-called mtDNA transfection) and (iii) manipulation of the heteroplasmic mtDNA balance (i.e. adjusting the wild-type:mutant type ratio), which can be achieved by improving a patients exercise regime.¹⁶⁰

More recently, and although in very early stages, allogenic haematopoietic stem cell therapy has been successfully used to treat mitochondrial neurogastrointestinal encephalomyopathy, but associated with high mortality.¹⁶¹ Similarly, liver transplants in patients (typically children) suffering from *MPV17*-associated hepatocerebral mitochondrial depletion syndrome have a poor prognosis.¹⁶²

Pre-implantation genetic diagnosis can assist female heteroplasmic mtDNA mutation carriers in determining the risk to their offspring, assisting by preventing transmission of deleterious mtDNA.^{163,164} Briefly, embryos obtained after *in vitro* fertilization are analysed and only those with very low-level mutant levels are transferred to the uterus. However, these techniques are of little help to woman harbouring intermediate-level heteroplasmic mtDNA mutations, where uncertainty regarding the clinical mutation threshold remains.¹⁶³

Advances, harnessing 'pro-nuclear transfer', have made significant steps towards treating primary mitochondrial disease at a mtDNA level.¹⁶⁵ Briefly, the technique involves the transfer of nDNA from a donor zygote (from the mtDNA mutation carrier mother) to an enucleated recipient zygote via fusion. The new 'reconstructed zygote' retains the nDNA from the mother, but the mtDNA from a donor. More recently, a competing group has attempted a similar technique, utilizing 'spindle transfer' of nDNA to an enucleated donor.¹⁶⁶ Unlike pro-nuclear transfer, nDNA isolation occurs pre-fertilization, meaning once the technique is approved it

can be integrated into established *in vitro* fertilization techniques. However, caution is advised, as both pro-nuclear transfer and spindle transfer would only benefit a minority of female mtDNA mutation carriers, whereas prenatal diagnostic testing can be utilized for both all Mendelian mitochondrial disorders and the majority of mtDNA mutations.^{163,167}

Funding

Funding to pay the Open Access publication charges for this article was provided by The Wellcome Trust.

References

- 1 van der Giezen M, Tovar J. Degenerate mitochondria. *EMBO Rep* 2005;**6**:525–30.
- 2 Schon EA, DiMauro S, Hirano M. Human mitochondrial DNA: roles of inherited and somatic mutations. *Nat Rev Genet* 2012;**13**:878–90.
- 3 Schapira AH. Mitochondrial diseases. *Lancet* 2012;**379**:1825–34.
- 4 Koopman WJ, Distelmaier F, Smeitink JA *et al.* OXPHOS mutations and neurodegeneration. *EMBO J* 2013;**32**:9–29.
- 5 Andrews RM, Kubacka I, Chinnery PF *et al.* Reanalysis and revision of the Cambridge reference sequence for human mitochondrial DNA. *Nat Genet* 1999;**23**:147.
- 6 Temperley R, Richter R, Dennerlein S *et al.* Hungry codons promote frameshifting in human mitochondrial ribosomes. *Science* 2010;**327**:301.
- 7 Sutovsky P. Ubiquitin-dependent proteolysis in mammalian spermatogenesis, fertilization, and sperm quality control: killing three birds with one stone. *Microsc Res Tech* 2003;**61**:88–102.
- 8 Vissing JSaM. Paternal Inheritance of mitochondrial DNA. *N Engl J Med* 2002;**208**:1–2.
- 9 Miller FJ, Rosenfeldt FL, Zhang C *et al.* Precise determination of mitochondrial DNA copy number in human skeletal and cardiac muscle by a PCR-based assay: lack of change of copy number with age. *Nucleic Acids Res* 2003;**31**:e61.
- 10 Birky CW Jr. The inheritance of genes in mitochondria and chloroplasts: laws, mechanisms, and models. *Annu Rev Genet* 2001;**35**:125–48.
- 11 Payne BA, Wilson IJ, Yu-Wai-Man P *et al.* Universal heteroplasmy of human mitochondrial DNA. *Hum Mol Genet* 2013;**22**:384–90.
- 12 Calvo S, Jain M, Xie X *et al.* Systematic identification of human mitochondrial disease genes through integrative genomics. *Nat Genet* 2006;**38**:576–82.
- 13 Hirst J. Why does mitochondrial complex I have so many subunits? *Biochem J* 2011;**437**:e1–3.
- 14 Angerer H, Zwicker K, Wumaier Z *et al.* A scaffold of accessory subunits links the peripheral arm and the distal proton-pumping module of mitochondrial complex I. *Biochem J* 2011;**437**:279–88.
- 15 Smith PM, Fox JL, Winge DR. Biogenesis of the cytochrome bc(1) complex and role of assembly factors. *Biochim Biophys Acta* 2012;**1817**:276–86.
- 16 Vogel RO, Smeitink JA, Nijtmans LG. Human mitochondrial complex I assembly: a dynamic and versatile process. *Biochim Biophys Acta* 2007;**1767**:1215–27.
- 17 Mick DU, Fox TD, Rehling P. Inventory control: cytochrome c oxidase assembly regulates mitochondrial translation. *Nat Rev Mol Cell Biol* 2011;**12**:14–20.
- 18 Wang ZG, White PS, Ackerman SH. Atp11p and Atp12p are assembly factors for the F(1)-ATPase in human mitochondria. *J Biol Chem* 2001;**276**:30773–8.
- 19 Copeland WC. Defects in mitochondrial DNA replication and human disease. *Crit Rev Biochem Mol Biol* 2012;**47**:64–74.
- 20 Bogenhagen DF. Mitochondrial DNA nucleoid structure. *Biochim Biophys Acta* 2012;**1819**:914–20.

- 21 Kukat C, Wurm CA, Spahr H *et al.* Super-resolution microscopy reveals that mammalian mitochondrial nucleoids have a uniform size and frequently contain a single copy of mtDNA. *Proc Natl Acad Sci USA* 2011;**108**:13534–9.
- 22 McCulloch V, Seidel-Rogol BL, Shadel GS. A human mitochondrial transcription factor is related to RNA adenine methyltransferases and binds S-adenosylmethionine. *Mol Cell Biol* 2002;**22**:1116–25.
- 23 McCulloch V, Shadel GS. Human mitochondrial transcription factor B1 interacts with the C-terminal activation region of h-mtTFA and stimulates transcription independently of its RNA methyltransferase activity. *Mol Cell Biol* 2003;**23**:5816–24.
- 24 Metodiev MD, Lesko N, Park CB *et al.* Methylation of 12S rRNA is necessary for in vivo stability of the small subunit of the mammalian mitochondrial ribosome. *Cell Metab* 2009;**9**:386–97.
- 25 Gaspari M, Larsson NG, Gustafsson CM. The transcription machinery in mammalian mitochondria. *Biochim Biophys Acta* 2004;**1659**:148–52.
- 26 Ma J, Farwell MA, Burkhart WA *et al.* Cloning and sequence analysis of the cDNA for bovine mitochondrial translational initiation factor 2. *Biochim Biophys Acta* 1995;**1261**:321–4.
- 27 Koc EC, Spremulli LL. Identification of mammalian mitochondrial translational initiation factor 3 and examination of its role in initiation complex formation with natural mRNAs. *J Biol Chem* 2002;**277**:35541–9.
- 28 Christian B, Haque E, Spremulli L. Ribosome shifting or splitting: it is all up to the EF-G. *Mol Cell* 2009;**35**:400–2.
- 29 Hammarsund M, Wilson W, Corcoran M *et al.* Identification and characterization of two novel human mitochondrial elongation factor genes, hEFG2 and hEFG1, phylogenetically conserved through evolution. *Hum Genet* 2001;**109**:542–50.
- 30 Ling M, Merante F, Chen HS *et al.* The human mitochondrial elongation factor tu (EF-Tu) gene: cDNA sequence, genomic localization, genomic structure, and identification of a pseudogene. *Gene* 1997;**197**:325–36.
- 31 Smits P, Smeitink J, van den Heuvel L. Mitochondrial translation and beyond: processes implicated in combined oxidative phosphorylation deficiencies. *J Biomed Biotechnol* 2010;**2010**:737385.
- 32 Zhang Y, Spremulli LL. Identification and cloning of human mitochondrial translational release factor 1 and the ribosome recycling factor. *Biochim Biophys Acta* 1998;**1443**:245–50.
- 33 Soleimanpour-Lichaei HR, Kuhl I, Gaisne M *et al.* mtRF1a is a human mitochondrial translation release factor decoding the major termination codons UAA and UAG. *Mol Cell* 2007;**27**:745–57.
- 34 Youle RJ, van der Blik AM. Mitochondrial fission, fusion, and stress. *Science* 2012;**337**:1062–5.
- 35 Schaefer AM, McFarland R, Blakely EL *et al.* Prevalence of mitochondrial DNA disease in adults. *Ann Neurol* 2008;**63**:35–9.
- 36 Skladal D, Halliday J, Thorburn DR. Minimum birth prevalence of mitochondrial respiratory chain disorders in children. *Brain* 2003;**126**(Pt 8):1905–12.
- 37 Majamaa K, Moilanen JS, Uimonen S *et al.* Epidemiology of A3243G, the mutation for mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episodes: prevalence of the mutation in an adult population. *Am J Hum Genet* 1998;**63**:447–54.
- 38 Darin N, Oldfors A, Moslemi AR *et al.* The incidence of mitochondrial encephalomyopathies in childhood: clinical features and morphological, biochemical, and DNA abnormalities. *Ann Neurol* 2001;**49**:377–83.
- 39 Magner M, Szentivanyi K, Svandova I *et al.* Elevated CSF-lactate is a reliable marker of mitochondrial disorders in children even after brief seizures. *Eur J Paediatr Neurol* 2011;**15**:101–8.
- 40 Barkovich AJ, Good WV, Koch TK *et al.* Mitochondrial disorders: analysis of their clinical and imaging characteristics. *AJNR Am J Neuroradiol* 1993;**14**:1119–37.
- 41 Lin DD, Crawford TO, Barker PB. Proton MR spectroscopy in the diagnostic evaluation of suspected mitochondrial disease. *AJNR Am J Neuroradiol* 2003;**24**:33–41.
- 42 Leonard JV, Schapira AH. Mitochondrial respiratory chain disorders I: mitochondrial DNA defects. *Lancet* 2000;**355**:299–304.
- 43 Leonard JV, Schapira AH. Mitochondrial respiratory chain disorders II: neurodegenerative disorders and nuclear gene defects. *Lancet* 2000;**355**:389–94.

- 44 Rahman S, Poulton J, Marchington D *et al.* Decrease of 3243 A→G mtDNA mutation from blood in MELAS syndrome: a longitudinal study. *Am J Hum Genet* 2001;68:238–40.
- 45 Harman D. The biologic clock: the mitochondria? *J Am Geriatr Soc* 1972;20:145–7.
- 46 Hegde ML, Mantha AK, Hazra TK *et al.* Oxidative genome damage and its repair: implications in aging and neurodegenerative diseases. *Mech Ageing Dev* 2012;133:157–68.
- 47 Howell N. Origin, cellular expression, and cybrid transmission of mitochondrial CAP-R, PYR-IND, and OLI-R mutant phenotypes. *Somat Cell Genet* 1983;9:1–24.
- 48 Chinnery PF, Howell N, Lightowlers RN *et al.* Molecular pathology of MELAS and MERRF. The relationship between mutation load and clinical phenotypes. *Brain* 1997;120:1713–21.
- 49 White SL, Collins VR, Wolfe R *et al.* Genetic counseling and prenatal diagnosis for the mitochondrial DNA mutations at nucleotide 8993. *Am J Hum Genet* 1999;65:474–82.
- 50 Weber K, Wilson JN, Taylor L *et al.* A new mtDNA mutation showing accumulation with time and restriction to skeletal muscle. *Am J Hum Genet* 1997;60:373–80.
- 51 Poulton J, O’Rahilly S, Morten KJ *et al.* Mitochondrial DNA, diabetes and pancreatic pathology in Kearns-Sayre syndrome. *Diabetologia* 1995;38:868–71.
- 52 Torroni A, Huoponen K, Francalacci P *et al.* Classification of European mtDNAs from an analysis of three European populations. *Genetics* 1996;144:1835–50.
- 53 Pello R, Martin MA, Carelli V *et al.* Mitochondrial DNA background modulates the assembly kinetics of OXPHOS complexes in a cellular model of mitochondrial disease. *Hum Mol Genet* 2008;17:4001–11.
- 54 Pyle A, Foltynie T, Tiangyou W *et al.* Mitochondrial DNA haplogroup cluster UKJT reduces the risk of PD. *Ann Neurol* 2005;57:564–7.
- 55 Santoro A, Balbi V, Balducci E *et al.* Evidence for sub-haplogroup h5 of mitochondrial DNA as a risk factor for late onset Alzheimer’s disease. *PLoS One* 2010;5:e12037.
- 56 Ridge PG, Maxwell TJ, Corcoran CD *et al.* Mitochondrial genomic analysis of late onset Alzheimer’s disease reveals protective haplogroups H6A1A/H6A1B: the Cache County Study on Memory in Aging. *PLoS One* 2012;7:e45134.
- 57 Udar N, Atilano SR, Memarzadeh M *et al.* Mitochondrial DNA haplogroups associated with age-related macular degeneration. *Invest Ophthalmol Vis Sci* 2009;50:2966–74.
- 58 Chinnery PF, Elliott HR, Hudson G *et al.* Epigenetics, epidemiology and mitochondrial DNA diseases. *Int J Epidemiol* 2012;41:177–87.
- 59 Ghezzi D, Zeviani M. Assembly factors of human mitochondrial respiratory chain complexes: physiology and pathophysiology. *Adv Exp Med Biol* 2012;748:65–106.
- 60 MITOMAP. A human mitochondrial genome database, 2000. <http://www.gen.emory.edu/mitomap.html>.
- 61 Rossignol R, Faustin B, Rocher C *et al.* Mitochondrial threshold effects. *Biochem J* 2003;370 (Pt 3):751–62.
- 62 Schon EA, Rizzuto R, Moraes CT *et al.* A direct repeat is a hotspot for large-scale deletion of human mitochondrial DNA. *Science* 1989;244:346–9.
- 63 Krishnan KJ, Reeve AK, Samuels DC *et al.* What causes mitochondrial DNA deletions in human cells? *Nat Genet* 2008;40:275–9.
- 64 Wallace DC, Singh G, Lott MT *et al.* Mitochondrial DNA mutation associated with Leber’s hereditary optic neuropathy. *Science* 1988;242:1427–30.
- 65 Carelli V, Ghelli A, Ratta M *et al.* Leber’s hereditary optic neuropathy: biochemical effect of 11778/ND4 and 3460/ND1 mutations and correlation with the mitochondrial genotype. *Neurology* 1997;48:1623–32.
- 66 Carelli V, La Morgia C, Valentino ML *et al.* Retinal ganglion cell neurodegeneration in mitochondrial inherited disorders. *Biochim Biophys Acta* 2009;1787:518–28.
- 67 Hudson G, Keers S, Yu Wai Man P *et al.* Identification of an X-chromosomal locus and haplotype modulating the phenotype of a mitochondrial DNA disorder. *Am J Hum Genet* 2005;77:1086–91.
- 68 Achilli A, Iommarini L, Olivieri A *et al.* Rare primary mitochondrial DNA mutations and probable synergistic variants in Leber’s hereditary optic neuropathy. *PLoS One* 2012;7:e42242.
- 69 Kirkman MA, Yu-Wai-Man P, Korsten A *et al.* Gene-environment interactions in Leber hereditary optic neuropathy. *Brain* 2009;132(Pt 9):2317–26.

- 70 Hudson G, Carelli V, Spruijt L *et al.* Clinical expression of Leber hereditary optic neuropathy is affected by the mitochondrial DNA-haplogroup background. *Am J Hum Genet* 2007;**81**:228–33.
- 71 Carvalho MR, Muller B, Rotzer E *et al.* Leber's hereditary optic neuroretinopathy and the X-chromosomal susceptibility factor: no linkage to DXS7. *Hum Hered* 1992;**42**:316–20.
- 72 Chen JD, Cox I, Denton MJ. Preliminary exclusion of an X-linked gene in Leber optic atrophy by linkage analysis. *Hum Genet* 1989;**82**:302–7.
- 73 Abu-Amero K, Jaber M, Hellani A *et al.* Genome-wide expression profile of LHON patients with the 11778 mutation. *Br J Ophthalmol* (2009/09/04 ed), 2009.
- 74 Phasukkijwatana N, Kunhapan B, Stankovich J *et al.* Genome-wide linkage scan and association study of PARL to the expression of LHON families in Thailand. *Hum Genet* (2010/04/22 ed).
- 75 Prezant TR, Agapian JV, Bohlman MC *et al.* Mitochondrial ribosomal RNA mutation associated with both antibiotic-induced and non-syndromic deafness. *Nat Genet* 1993;**4**:289–94.
- 76 Zifa E, Giannouli S, Theotokis P *et al.* Mitochondrial tRNA mutations: clinical and functional perturbations. *RNA Biol* 2007;**4**:38–66.
- 77 McFarland R, Elson JL, Taylor RW *et al.* Assigning pathogenicity to mitochondrial tRNA mutations: when 'definitely maybe' is not good enough. *Trends Genet* 2004;**20**:591–6.
- 78 Pavlakis SG, Phillips PC, DiMauro S *et al.* Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes: a distinctive clinical syndrome. *Ann Neurol* 1984;**16**:481–8.
- 79 Hirano M, Ricci E, Koenigsberger MR *et al.* Melas: an original case and clinical criteria for diagnosis. *Neuromuscul Disord* 1992;**2**:125–35.
- 80 Manwaring N, Jones MM, Wang JJ *et al.* Population prevalence of the MELAS A3243G mutation. *Mitochondrion* 2007;**7**:230–3.
- 81 Goto Y, Nonaka I, Horai S. A mutation in the tRNA(Leu)(UUR) gene associated with the MELAS subgroup of mitochondrial encephalomyopathies. *Nature* 1990;**348**:651–3.
- 82 Santorelli FM, Tanji K, Kulikova R *et al.* Identification of a novel mutation in the mtDNA ND5 gene associated with MELAS. *Biochem Biophys Res Commun* 1997;**238**:326–8.
- 83 Shoffner JM, Lott MT, Lezza AM *et al.* Myoclonic epilepsy and ragged-red fiber disease (MERRF) is associated with a mitochondrial DNA tRNA(Lys) mutation. *Cell* 1990;**61**:931–7.
- 84 Zeviani M, Moraes CT, DiMauro S *et al.* Deletions of mitochondrial DNA in Kearns-Sayre syndrome. *Neurology* 1988;**38**:1339–46.
- 85 Moraes CT, DiMauro S, Zeviani M *et al.* Mitochondrial DNA deletions in progressive external ophthalmoplegia and Kearns-Sayre syndrome. *N Engl J Med* 1989;**320**:1293–9.
- 86 Rotig A, Cormier V, Blanche S *et al.* Pearson's marrow-pancreas syndrome. A multisystem mitochondrial disorder in infancy. *J Clin Invest* 1990;**86**:1601–8.
- 87 Horvath R, Hudson G, Ferrari G *et al.* Phenotypic spectrum associated with mutations of the mitochondrial polymerase gamma gene. *Brain* 2006;**129**(Pt 7):1674–84.
- 88 Spelbrink JN, Li FY, Tiranti V *et al.* Human mitochondrial DNA deletions associated with mutations in the gene encoding Twinkle, a phage T7 gene4-like protein localized in mitochondria. *Nat Genet* 2001;**28**:223–31.
- 89 Agostino A, Valletta L, Chinnery PF *et al.* Mutations of ANT1, Twinkle, and POLG1 in sporadic progressive external ophthalmoplegia (PEO). *Neurology* 2003;**60**:1354–6.
- 90 Wolf A, Cowen D. The cerebral atrophies and encephalomalacias of infancy and childhood. *Res Publ Assoc Res Nerv Ment Dis* 1955;**34**:199–330.
- 91 Kollberg G, Moslemi AR, Darin N *et al.* POLG1 mutations associated with progressive encephalopathy in childhood. *J Neuropathol Exp Neurol* 2006;**65**:758–68.
- 92 Naviaux RK, Nguyen KV. POLG mutations associated with Alpers' syndrome and mitochondrial DNA depletion. *Ann Neurol* 2004;**55**:706–12.
- 93 Clayton DA. Replication of animal mitochondrial DNA. *Cell* 1982;**28**:693–705.
- 94 Spinazzola A, Zeviani M. Disorders of nuclear-mitochondrial intergenomic communication. *Biosci Rep* 2007;**27**:39–51.
- 95 Haack TB, Haberberger B, Frisch EM *et al.* Molecular diagnosis in mitochondrial complex I deficiency using exome sequencing. *J Med Genet* 2012;**49**:277–83.
- 96 Loeffen JL, Smeitink JA, Trijbels JM *et al.* Isolated complex I deficiency in children: clinical, biochemical and genetic aspects. *Hum Mutat* 2000;**15**:123–34.

- 97 Lebre AS, Rio M, Faivre d’Arcier L *et al.* A common pattern of brain MRI imaging in mitochondrial diseases with complex I deficiency. *J Med Genet* 2011;**48**:16–23.
- 98 Benit P, Chretien D, Kadhom N *et al.* Large-scale deletion and point mutations of the nuclear NDUFV1 and NDUFS1 genes in mitochondrial complex I deficiency. *Am J Hum Genet* 2001;**68**:1344–52.
- 99 Benit P, Slama A, Cartault F *et al.* Mutant NDUFS3 subunit of mitochondrial complex I causes Leigh syndrome. *J Med Genet* 2004;**41**:14–7.
- 100 van den Heuvel L, Ruitenbeek W, Smeets R *et al.* Demonstration of a new pathogenic mutation in human complex I deficiency: a 5-bp duplication in the nuclear gene encoding the 18-kD (AQDQ) subunit. *Am J Hum Genet* 1998;**62**:262–8.
- 101 Smeitink J, van den Heuvel L. Human mitochondrial complex I in health and disease. *Am J Hum Genet* 1999;**64**:1505–10.
- 102 Loeffen J, Smeitink J, Triepels R *et al.* The first nuclear-encoded complex I mutation in a patient with Leigh syndrome. *Am J Hum Genet* 1998;**63**:1598–608.
- 103 Schuelke M, Smeitink J, Mariman E *et al.* Mutant NDUFV1 subunit of mitochondrial complex I causes leukodystrophy and myoclonic epilepsy. *Nat Genet* 1999;**21**:260–1.
- 104 Hoefs SJ, van Spronsen FJ, Lenssen EW *et al.* NDUFA10 mutations cause complex I deficiency in a patient with Leigh disease. *Eur J Hum Genet* 2011;**19**:270–4.
- 105 Hoefs SJ, Dieteren CE, Distelmaier F *et al.* NDUFA2 complex I mutation leads to Leigh disease. *Am J Hum Genet* 2008;**82**:1306–15.
- 106 Shoubridge EA. Nuclear genetic defects of oxidative phosphorylation. *Hum Mol Genet* 2001;**10**:2277–84.
- 107 Loeffen J, Elpeleg O, Smeitink J *et al.* Mutations in the complex I NDUFS2 gene of patients with cardiomyopathy and encephalomyopathy. *Ann Neurol* 2001;**49**:195–201.
- 108 Kirby DM, Salemi R, Sugiana C *et al.* NDUFS6 mutations are a novel cause of lethal neonatal mitochondrial complex I deficiency. *J Clin Invest* 2004;**114**:837–45.
- 109 Benit P, Beugnot R, Chretien D *et al.* Mutant NDUFV2 subunit of mitochondrial complex I causes early onset hypertrophic cardiomyopathy and encephalopathy. *Hum Mutat* 2003;**21**:582–6.
- 110 Berger I, Hershkovitz E, Shaag A *et al.* Mitochondrial complex I deficiency caused by a deleterious NDUFA11 mutation. *Ann Neurol* 2008;**63**:405–8.
- 111 Haack TB, Danhauser K, Haberberger B *et al.* Exome sequencing identifies ACAD9 mutations as a cause of complex I deficiency. *Nat Genet* 2010;**42**:1131–4.
- 112 Calvo SE, Tucker EJ, Compton AG *et al.* High-throughput, pooled sequencing identifies mutations in NUBPL and FOXRED1 in human complex I deficiency. *Nat Genet* 2010;**42**:851–8.
- 113 Gerards M, Sluiter W, van den Bosch BJ *et al.* Defective complex I assembly due to C20orf7 mutations as a new cause of Leigh syndrome. *J Med Genet* 2010;**47**:507–12.
- 114 Saada A, Edvardson S, Rapoport M *et al.* C6ORF66 is an assembly factor of mitochondrial complex I. *Am J Hum Genet* 2008;**82**:32–8.
- 115 Dunning CJ, McKenzie M, Sugiana C *et al.* Human CIA30 is involved in the early assembly of mitochondrial complex I and mutations in its gene cause disease. *EMBO J* 2007;**26**:3227–37.
- 116 Baysal BE. Hereditary paraganglioma targets diverse paraganglia. *J Med Genet* 2002;**39**:617–22.
- 117 Petruzzella V, Tiranti V, Fernandez P *et al.* Identification and characterization of human cDNAs specific to BCS1, PET112, SCO1, COX15, and COX11, five genes involved in the formation and function of the mitochondrial respiratory chain. *Genomics* 1998;**54**:494–504.
- 118 de Lonlay P, Valnot I, Barrientos A *et al.* A mutant mitochondrial respiratory chain assembly protein causes complex III deficiency in patients with tubulopathy, encephalopathy and liver failure. *Nat Genet* 2001;**29**:57–60.
- 119 Haut S, Brivet M, Touati G *et al.* A deletion in the human QP-C gene causes a complex III deficiency resulting in hypoglycaemia and lactic acidosis. *Hum Genet* 2003;**113**:118–22.
- 120 Barel O, Shorer Z, Flusser H *et al.* Mitochondrial complex III deficiency associated with a homozygous mutation in UQCRCQ. *Am J Hum Genet* 2008;**82**:1211–6.
- 121 Ghezzi D, Arzuffi P, Zordan M *et al.* Mutations in TTC19 cause mitochondrial complex III deficiency and neurological impairment in humans and flies. *Nat Genet* 2011;**43**:259–63.

- 122 Massa V, Fernandez-Vizarra E, Alshahwan S *et al.* Severe infantile encephalomyopathy caused by a mutation in COX6B1, a nucleus-encoded subunit of cytochrome c oxidase. *Am J Hum Genet* 2008;**82**:1281–9.
- 123 Antonicka H, Leary SC, Guercin GH *et al.* Mutations in COX10 result in a defect in mitochondrial heme A biosynthesis and account for multiple, early-onset clinical phenotypes associated with isolated COX deficiency. *Hum Mol Genet* 2003;**12**:2693–702.
- 124 Coenen MJ, van den Heuvel LP, Ugalde C *et al.* Cytochrome c oxidase biogenesis in a patient with a mutation in COX10 gene. *Ann Neurol* 2004;**56**:560–4.
- 125 Indrieri A, van Rahden VA, Tiranti V *et al.* Mutations in COX7B cause microphthalmia with linear skin lesions, an unconventional mitochondrial disease. *Am J Hum Genet* 2012;**91**:942–9.
- 126 Zvulunov A, Kachko L, Manor E *et al.* Reticulolinear aplasia cutis congenita of the face and neck: a distinctive cutaneous manifestation in several syndromes linked to Xp22. *Br J Dermatol* 1998;**138**:1046–52.
- 127 Shteyer E, Saada A, Shaag A *et al.* Exocrine pancreatic insufficiency, dyserythropoietic anemia, and calvarial hyperostosis are caused by a mutation in the COX4I2 gene. *Am J Hum Genet* 2009;**84**:412–7.
- 128 Zhu Z, Yao J, Johns T *et al.* SURF1, encoding a factor involved in the biogenesis of cytochrome c oxidase, is mutated in Leigh syndrome. *Nat Genet* 1998;**20**:337–43.
- 129 Tiranti V, Jaksch M, Hofmann S *et al.* Loss-of-function mutations of SURF-1 are specifically associated with Leigh syndrome with cytochrome c oxidase deficiency. *Ann Neurol* 1999;**46**:161–6.
- 130 Weraarpachai W, Sasarman F, Nishimura T *et al.* Mutations in C12orf62, a factor that couples COX I synthesis with cytochrome c oxidase assembly, cause fatal neonatal lactic acidosis. *Am J Hum Genet* 2012;**90**:142–51.
- 131 Huigsloot M, Nijtmans LG, Szklarczyk R *et al.* A mutation in C2orf64 causes impaired cytochrome c oxidase assembly and mitochondrial cardiomyopathy. *Am J Hum Genet* 2011;**88**:488–93.
- 132 Ghezzi D, Saada A, D'Adamo P *et al.* FASTKD2 nonsense mutation in an infantile mitochondrial encephalomyopathy associated with cytochrome c oxidase deficiency. *Am J Hum Genet* 2008;**83**:415–23.
- 133 Mayr JA, Havlickova V, Zimmermann F *et al.* Mitochondrial ATP synthase deficiency due to a mutation in the ATP5E gene for the F1 epsilon subunit. *Hum Mol Genet* 2010;**19**:3430–9.
- 134 De Meirleir L, Seneca S, Lissens W *et al.* Respiratory chain complex V deficiency due to a mutation in the assembly gene ATP12. *J Med Genet* 2004;**41**:120–4.
- 135 Saada A, Shaag A, Arnon S *et al.* Antenatal mitochondrial disease caused by mitochondrial ribosomal protein (MRPS22) mutation. *J Med Genet* 2007;**44**:784–6.
- 136 Zeharia A, Fischel-Ghodsian N, Casas K *et al.* Mitochondrial myopathy, sideroblastic anemia, and lactic acidosis: an autosomal recessive syndrome in Persian Jews caused by a mutation in the PUS1 gene. *J Child Neurol* 2005;**20**:449–52.
- 137 Smirnova E, Shurland DL, Ryazantsev SN *et al.* A human dynamin-related protein controls the distribution of mitochondria. *J Cell Biol* 1998;**143**:351–8.
- 138 Waterham HR, Koster J, van Roermund CW *et al.* A lethal defect of mitochondrial and peroxisomal fission. *N Engl J Med* 2007;**356**:1736–41.
- 139 Cree LM, Samuels DC, de Sousa Lopes SC *et al.* A reduction of mitochondrial DNA molecules during embryogenesis explains the rapid segregation of genotypes. *Nat Genet* 2008;**40**:249–54.
- 140 Carling PJ, Cree LM, Chinnery PF. The implications of mitochondrial DNA copy number regulation during embryogenesis. *Mitochondrion* 2011;**11**:686–92.
- 141 DiMauro S, Schon EA. Mitochondrial DNA mutations in human disease. *Am J Med Genet* 2001;**106**:18–26.
- 142 Clark MJ, Chen R, Lam HY *et al.* Performance comparison of exome DNA sequencing technologies. *Nat Biotechnol* 2011;**29**:908–14.
- 143 McCormick E, Place E, Falk MJ. Molecular Genetic Testing for Mitochondrial Disease: From One Generation to the Next. *Neurotherapeutics* 2013;**10**:251–61.
- 144 Pagliarini DJ, Calvo SE, Chang B *et al.* A mitochondrial protein compendium elucidates complex I disease biology. *Cell* 2008;**134**:112–23.

- 145 Prokisch H, Andreoli C, Ahting U *et al*. MitoP2: the mitochondrial proteome database—now including mouse data. *Nucleic Acids Res* 2006;**34**(database issue):D705–11.
- 146 Pierson TM, Adams D, Bonn F *et al*. Whole-exome sequencing identifies homozygous AFG3L2 mutations in a spastic ataxia-neuropathy syndrome linked to mitochondrial m-AAA proteases. *PLoS Genet* 2011;**7**:e1002325.
- 147 Vedrenne V, Gowher A, De Lonlay P *et al*. Mutation in PNPT1, which encodes a polyribonucleotide nucleotidyltransferase, impairs RNA import into mitochondria and causes respiratory-chain deficiency. *Am J Hum Genet* 2012;**91**:912–8.
- 148 Janer A, Antonicka H, Lalonde E *et al*. An RMND1 Mutation causes encephalopathy associated with multiple oxidative phosphorylation complex deficiencies and a mitochondrial translation defect. *Am J Hum Genet* 2012;**91**:737–43.
- 149 Casey JP, McGettigan P, Lynam-Lennon N *et al*. Identification of a mutation in LARS as a novel cause of infantile hepatopathy. *Mol Genet Metab* 2012;**106**:351–8.
- 150 Galmiche L, Serre V, Beinat M *et al*. Exome sequencing identifies MRPL3 mutation in mitochondrial cardiomyopathy. *Hum Mutat* 2011;**32**:1225–31.
- 151 Gotz A, Tynynmaa H, Euro L *et al*. Exome sequencing identifies mitochondrial alanyl-tRNA synthetase mutations in infantile mitochondrial cardiomyopathy. *Am J Hum Genet* 2011;**88**:635–42.
- 152 Pfeiffer G, Elliott HR, Griffin H *et al*. Titin mutation segregates with hereditary myopathy with early respiratory failure. *Brain* 2012;**135**(Pt 6):1695–713.
- 153 Chinnery PF, Thorburn DR, Samuels DC *et al*. The inheritance of mitochondrial DNA heteroplasmy: random drift, selection or both? *Trends Genet* 2000;**16**:500–5.
- 154 Monnot S, Gigarel N, Samuels DC *et al*. Segregation of mtDNA throughout human embryofetal development: m.3243A>G as a model system. *Hum Mutat* 2011;**32**:116–25.
- 155 Pfeiffer G, Majamaa K, Turnbull DM *et al*. Treatment for mitochondrial disorders. *Cochrane Database Syst Rev* 2012;**4**:CD004426.
- 156 Klopstock T, Yu-Wai-Man P, Dimitriadis K *et al*. A randomized placebo-controlled trial of idebenone in Leber's hereditary optic neuropathy. *Brain* 2011;**134**:2677–86.
- 157 Kyriakouli DS, Boesch P, Taylor RW *et al*. Progress and prospects: gene therapy for mitochondrial DNA disease. *Gene Ther* 2008;**15**:1017–23.
- 158 Nagley P, Farrell LB, Gearing DP *et al*. Assembly of functional proton-translocating ATPase complex in yeast mitochondria with cytoplasmically synthesized subunit 8, a polypeptide normally encoded within the organelle. *Proc Natl Acad Sci USA* 1988;**85**:2091–5.
- 159 Bonnet C, Kaltimbacher V, Ellouze S *et al*. Allotopic mRNA localization to the mitochondrial surface rescues respiratory chain defects in fibroblasts harboring mitochondrial DNA mutations affecting complex I or v subunits. *Rejuvenation Res* 2007;**10**:127–44.
- 160 Taivassalo T, Gardner JL, Taylor RW *et al*. Endurance training and detraining in mitochondrial myopathies due to single large-scale mtDNA deletions. *Brain* 2006;**129**(Pt 12):3391–401.
- 161 Halter J, Schupbach WM, Casali C *et al*. Allogeneic hematopoietic SCT as treatment option for patients with mitochondrial neurogastrointestinal encephalomyopathy (MNGIE): a consensus conference proposal for a standardized approach. *Bone Marrow Transplant* 2011;**46**:330–7.
- 162 Wong LJ, Brunetti-Pierri N, Zhang Q *et al*. Mutations in the MPV17 gene are responsible for rapidly progressive liver failure in infancy. *Hepatology* 2007;**46**:1218–27.
- 163 Hellebrekers DM, Wolfe R, Hendrickx AT *et al*. PGD and heteroplasmic mitochondrial DNA point mutations: a systematic review estimating the chance of healthy offspring. *Hum Reprod Update* 2012;**18**:341–9.
- 164 Thorburn D, Wilton L, Stock-Myer S. Healthy baby girl born following pre-implantation Genetic diagnosis for mitochondrial DNA m.8993t>g Mutation. *Mol Genet Metab* 2009;**98**:5–6.
- 165 Craven L, Tuppen HA, Greggains GD *et al*. Pronuclear transfer in human embryos to prevent transmission of mitochondrial DNA disease. *Nature* 2010;**465**:82–5.
- 166 Tachibana M, Amato P, Sparman M *et al*. Towards germline gene therapy of inherited mitochondrial diseases. *Nature* 2013;**493**:627–31.
- 167 Poulton J, Oakeshott P. Nuclear transfer to prevent maternal transmission of mitochondrial DNA disease. *BMJ* 2012;**345**:e6651.