





Canadian Journal of Cardiology 32 (2016) 659-668

Review

Vascular Fibrosis in Aging and Hypertension: Molecular Mechanisms and Clinical Implications

Adam Harvey, PhD, Augusto C. Montezano, PhD, Rheure Alves Lopes, MSc, Francisco Rios, PhD, and Rhian M. Touyz, MBBCh, PhD

Institute of Cardiovascular and Medical Sciences, BHF Glasgow Cardiovascular Research Centre, University of Glasgow, Glasgow, Scotland

ABSTRACT

Aging is the primary risk factor underlying hypertension and incident cardiovascular disease. With aging, the vasculature undergoes structural and functional changes characterized by endothelial dysfunction, wall thickening, reduced distensibility, and arterial stiffening. Vascular stiffness results from fibrosis and extracellular matrix (ECM) remodelling, processes that are associated with aging and are amplified by hypertension. Some recently characterized molecular mechanisms underlying these processes include increased expression and activation of matrix metalloproteinases, activation of transforming growth factor- $\beta 1/SMAD$ signalling, upregulation of galectin-3, and activation of proinflammatory and profibrotic signalling pathways. These events can be induced by vasoactive agents, such as angiotensin II, endothelin-1, and aldosterone, which are increased in the vasculature

RÉSUMÉ

Le vieillissement constitue le principal facteur de risque d'apparition de l'hypertension et de la maladie cardiovasculaire. En vieillissant, le système vasculaire subit des modifications structurelles et fonctionnelles caractérisées par une dysfonction endothéliale ainsi que l'épaississement, la rigidification et la perte d'élasticité des parois vasculaires. La rigidité vasculaire est causée par la fibrose et le remodelage de la matrice extracellulaire, des processus qui sont associés au vieillissement et qui sont amplifiés en présence d'hypertension. Parmi les mécanismes moléculaires sous-jacents du vieillissement récemment identifiés, on retrouve l'augmentation de l'expression et de l'activation des métalloprotéinases matricielles, l'activation des voies de signalisation du facteur de croissance transformant bêta 1 impliquant les protéines SMAD, la régulation positive

Hypertension is the largest contributor to the global burden of cardiovascular disease. The World Health Organization estimates that the number of adults with high blood pressure will increase from 1 billion to 1.5 billion worldwide by 2020. This increase is related in part to the fact that the population is aging. Of all the factors contributing to hypertension, such as genetics, obesity, dyslipidemia, sedentary lifestyle, and diabetes, advancing age is the most important risk factor. Both aging and hypertension are associated with structural, mechanical, and functional changes in the vasculature, characterized by increased arterial stiffness, reduced elasticity, impaired distensibility, endothelial dysfunction, and increased vascular tone. The prevalence of vascular stiffness and high blood pressure increases with age and as such, hypertension has been considered to be a condition of aging. Arterial

Received for publication February 10, 2016. Accepted February 18, 2016.

Corresponding author: Dr Rhian M. Touyz, Institute of Cardiovascular and Medical Sciences, BHF Glasgow Cardiovascular Research Centre, University of Glasgow, 126 University Place, Glasgow G12 8TA, Scotland. Tel.: + 44 (0)141 330 7775/7774; fax: + 44 (0)141 330-3360.

E-mail: rhian.touyz@glasgow.ac.uk See page 666 for disclosure information. stiffening precedes the development of hypertension, and both phenomena occur more frequently in the elderly. The relationship between aging, cardiovascular disease, and vascular stiffening is further exemplified in patients with progeria (premature aging), who exhibit accelerated vascular aging and often die of cardiovascular disease.² Arterial stiffening is caused primarily by excessive fibrosis and reduced elasticity, with associated increased collagen deposition, increased elastin fiber fragmentation/degeneration, laminar medial necrosis, calcification, and cross-linking of collagen molecules by advanced glycation end-products.

Fibrosis as a dynamic process initially is an adaptive repair response that is reversible. However, the fibrogenic process is progressive, leading to further worsening of arterial stiffness and fibrosis that gradually extends into the neighbouring interstitial space. Fibrosis occurs in both large and small arteries. In large vessels, vascular stiffening leads to hemodynamic damage to peripheral tissues.³ Fibrosis and stiffening of the resistance circulation impair endothelial function, increase vasomotor tone, promote vascular rarefaction, and alter tissue perfusion. The combination of "aging" and prohypertensive elements, such as activation of the renin-angiotensinaldosterone system, inflammation, oxidative stress, salt

during aging and hypertension. Complex interplay between the "aging process" and prohypertensive factors results in accelerated vascular remodelling and fibrosis and increased arterial stiffness, which is typically observed in hypertension. Because the vascular phenotype in a young hypertensive individual resembles that of an elderly otherwise healthy individual, the notion of "early" or "premature" vascular aging is now often used to describe hypertension-associated vascular disease. We review the vascular phenotype in aging and hypertension, focusing on arterial stiffness and vascular remodelling. We also highlight the clinical implications of these processes and discuss some novel molecular mechanisms of fibrosis and ECM reorganization.

consumption, and genetic factors, results in excessive arterial fibrosis and extracellular matrix (ECM) deposition with amplification of aging-related vascular injury and stiffness. These processes lead to excessive fibrosis, which often extends from small arteries and replaces parenchymal tissue, thereby leading to tissue fibrosis, scarring, and hypertension-associated target organ damage of the heart, kidney, and brain.

At the molecular and cellular levels, arterial aging and hypertension-associated vascular changes are characterized by reduced nitric oxide production, increased generation of reactive oxygen species (ROS) (oxidative stress), activation of transcription factors, induction of "aging" genes, stimulation of proinflammatory and profibrotic signalling pathways, reduced collagen turnover, calcification, vascular smooth muscle cell proliferation, and ECM remodelling. These processes contribute to increased fibrosis, which is further promoted by prohypertensive vasoactive agents, such as angiotensin II (Ang II), endothelin-1 (ET-1), and aldosterone, which stimulate profibrotic signalling cascades, including p38 mitogen-activated protein kinases (p38 MAPK) and the transforming growth factor-β (TGF-β)/SMAD pathway. Activation of galectin-3 and dysregulation of MMPs and TIMPs are involved in ECM remodelling and further enhance vascular fibrosis. Many of these events are upregulated with advancing age and in human and experimental hypertension. We review the vascular phenotype in physiological aging and in hypertension, focusing particularly on arterial stiffness and fibrosis.

Aging-Associated Vascular Alterations

With aging, the vasculature undergoes functional, structural, and mechanical changes characterized by endothelial dysfunction, thickening (remodelling) of the vascular wall, and increased stiffening, respectively (Fig. 1). These changes result in a reduced capacity of arteries to adapt to tissue demands and accordingly may lead to ischemic injury. Preclinical and clinical studies have clearly demonstrated that with aging, there is impaired endothelium-dependent vasorelaxation with associated increased permeability and vascular inflammation.

de la galectine-3 et l'activation des voies de signalisation proinflammatoires et profibrotiques. Ces mécanismes peuvent être induits par divers agents vasoactifs comme l'angiotensine II, l'endothéline-1 et l'aldostérone dont la présence s'accroît au fil du processus de vieillissement et en présence d'hypertension. Cette interaction complexe entre le « processus de vieillissement » et les facteurs pro-hypertensifs entraîne un remodelage et une fibrose accélérée ainsi que la rigidification des artères qu'on observe habituellement avec l'hypertension. Puisque le phénotype vasculaire de l'hypertendu jeune ressemble à celui de la personne âgée par ailleurs en bonne santé, on fait désormais de plus en plus souvent appel au vocable de vieillissement vasculaire « précoce » ou « prématuré » pour désigner la maladie vasculaire liée à l'hypertension. Nous passons ici en revue le phénotype vasculaire du vieillissement et de l'hypertension en mettant l'accent sur la rigidité artérielle et le remodelage vasculaire. Nous traitons également de l'incidence clinique de ces processus, en plus d'aborder quelques-uns des mécanismes moléculaires de la fibrose et de la réorganisation de la matrice extracellulaire.

Epidemiologic, cross-sectional, clinical, and postmortem studies in healthy individuals of variable ages have clearly demonstrated that intimal wall thickening and dilatation are noticeable structural changes that occur in conduit arteries with advanced age. Findings from noninvasive vascular phenotyping studies in healthy individuals have demonstrated that intima-media thickness increases 2- to 3-fold between 20 and 90 years of age. 4 Studies in aging nonhuman primates also showed a relationship between intimal thickness in the thoracic aorta and aging.5 Exact factors causing progressive intimal thickening with aging in otherwise healthy individuals remain elusive, but a number of distinctive changes at the cellular and morphologic levels have been identified, including fracture of elastin fibres within the tunica media, increased collagen deposition, cellular senescence, and dysregulated cell proliferation. Associated with these events is remodelling of the ECM, which is an essential component of the connective tissue surrounding the vascular wall.

The ECM is composed of basic structural elements (collagen and elastin) and more specialized proteins including fibronectin and proteoglycans. The ECM is a dynamic structure and its components are continuously being turned over through highly regulated systems involving activation of MMPs and TIMPs. Dysregulation of these processes, together with alterations in profibrotic and proinflammatory signalling pathways, likely contribute to aging-associated vascular structural changes.

The Vascular Phenotype in Hypertension Resembles Aging-Associated Vascular Remodelling

The overall vascular phenotype of an individual at any 1 time depends not only on "aging" but also on a combination of multiple interacting factors, such as genetic factors, diet, smoking, diabetes, dyslipidemia, oxidative stress, and obesity. Moreover, in the presence of prohypertensive factors, there is acceleration of aging-associated vascular changes that leads to exaggerated vascular injury and arterial stiffening. In susceptible individuals, the interplay between aging and hypertension leads to "early vascular aging" and arterial

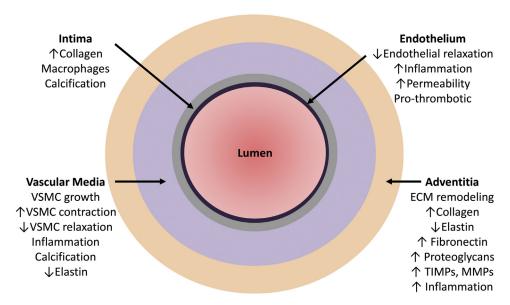


Figure 1. The vascular phenotype in aging and hypertension. With aging and during the development of hypertension, the endothelium, vascular wall, and adventitia undergo functional and structural changes. Endothelial function is impaired and the vascular media is thickened. The adventitial extracellular matrix undergoes remodelling, with increased collagen deposition, reduced elastin content, and increased proinflammatory cells. These processes contribute to vascular fibrosis and stiffening. ECM, extracellular matrix; MMP, matrix metalloproteinases; TIMPs, tissue inhibitory metalloproteinases; VSMC, vascular smooth muscle cell.

stiffness, in which the vascular phenotype in young hypertensive individuals resembles that of elderly otherwise healthy individuals (Fig. 1).

Arterial Stiffness

Normally, conduit arteries distend to accommodate large pressure ejections from the heart during systole to facilitate perfusion to tissues during diastole. This is determined in large part by the elasticity, distensibility, and compliance of the arterial system. Loss of elasticity and increased stiffness demand greater force to accommodate blood flow, leading to increased systolic blood pressure, increased cardiac work load, and consequent cardiac hypertrophy and risk of cardiovascular events. Aortic stiffness also affects the microcirculation and vice versa.^{7,8} Aortic wall stiffening causes increased pulse wave velocity (PWV) and premature reflected waves with elevated central hemodynamic load leading to damage of peripheral small arteries. Remodelling of small arteries in turn leads to increased peripheral vascular and pulse wave reflection, which can further contribute to aortic stiffness. 10 Arterial stiffness can be assessed by measuring PWV, pulse wave analysis, ambulatory arterial stiffness (using 24-hour ambulatory blood pressure monitoring) and evaluating endothelial function (flow-mediated dilation). PWV is the most commonly used approach and measures the speed of the pressure pulse from the heart as it is propagated through the arteries; it is calculated by dividing the distance travelled by the time taken to travel the defined distance. Stiffer arteries result in a more rapid travel time and hence a higher PWV. Various approaches can be used to measure PWV, including applanation tonometry, oscillometry, Doppler echocardiography, and magnetic resonance imaging. Although the measurement of PWV is considered to be the most simple, noninvasive, robust, and reproducible method to determine arterial

stiffness, ¹¹ it is not yet used in routine clinical practice. Carotid-femoral PWV is a direct measure of aortic stiffness and is now considered the gold standard for its evaluation in clinical and epidemiologic studies. ¹²

Arterial stiffness is a natural consequence of advancing age and is accelerated in hypertension. It is also an independent predictive risk factor for cardiovascular events and, as such, aortic PWV is now recognized as an important biomarker in the determination of cardiovascular risk. Arterial stiffness has a bidirectional causal relationship with blood pressure, because high blood pressure causes arterial wall injury, which promotes stiffening, whereas arterial stiffening itself is the major cause of increased systolic blood pressure, especially in the elderly, 8,13 Multiple interacting factors at the systemic (blood pressure, hemodynamics), vascular (vascular contraction/dilatation, ECM remodelling), cellular (cytoskeletal organization, inflammatory responses), and molecular (oxidative stress, intracellular signalling, mechanotransduction) contribute to arterial stiffness in aging and hypertension. Dysregulation of endothelial cells, vascular smooth muscle cells, and adaptive immune responses has also been implicated in arterial aging and vascular damage in hypertension. A detailed discussion of all these mechanisms is beyond the scope of this review and is addressed elsewhere this issue of the Canadian Journal of Cardiology. 14 Here we focus on some molecular and cellular events that contribute to vascular fibrosis and ECM remodelling.

The ECM and Vascular Fibrosis in Aging and Hypertension

The ECM is an essential component of the connective tissue that surrounds cells. In addition to maintaining cellular and vascular integrity, it plays a fundamental role in cell signalling and regulation of cell-cell interactions. The ECM

comprises multiple structural proteins, including collagens, elastin, fibronectin, and proteoglycans. Composition of the ECM varies from organ to organ, with collagen types I and III representing the predominant isoforms in the vascular ECM. 15 The absolute and relative quantities of collagen and elastin determine biomechanical properties of vessels, in which an elastin deficiency/collagen excess leads to vascular fibrosis and increased stiffness. 4,15 In healthy individuals, collagen deposition and turnover are tightly regulated, and the ratio of collagen to elastin remains relatively constant. However, an imbalance in these processes leads to excessive ECM protein deposition, particularly collagen and fibronectin, contributing to vascular fibrosis and stiffening in aging and during the development of hypertension. 15 Collagens are particularly important in these processes because they are the most abundant and stiffest of the ECM proteins. Increased collagen content and destruction of the elastin fiber network together with a proinflammatory microenvironment contribute to ECM remodelling and increased intima-media thickening and vascular stiffness in small and large arteries in human and experimental hypertension.

Contributing to the profibrotic process is transglutaminase (TG2), which is secreted into the ECM, where it catalyzes formation of ε -(γ -glutamyl)lysine isopeptide, in a Ca²⁺-dependent manner. ¹⁶ TG2 acts as an extracellular scaffold protein as well as a cross-linking enzyme. Numerous ECM proteins are TG2 substrates, such as fibronectin, collagen, and laminin. 16 Under physiological conditions, TG2 regulates fibroblast activity and ECM organization, with little protein cross-linking. However, in pathologic conditions, increased TG2/ECM protein crosslinking and altered TG2 activity cause increased rigidity and stiffening of the vascular wall, processes that may contribute to remodelling in aging and cardiovascular disease. Recent evidence indicates altered TG2 activity and functionality in large arteries of hypertensive rats. TG2 dysregulation has also been implicated in small-vessel changes and inward remodelling in hypertension. 18 Fundamental to many of the processes underlying ECM reorganization and fibrosis in aging and hypertension is activation of MMPs and TIMPs.

MMPs and TIMPs

ECM proteins, including collagen and elastin, are regulated by MMPs, a family of endopeptidases, which are activated by many factors associated with aging and hypertension, such as proinflammatory signalling molecules (cytokines, interleukins), growth factors, vasoactive agents (Ang II, ET-1, aldosterone) and ROS. MMP activity is controlled at 3 levels: gene transcription, proenzyme activation, and activity inhibition. 18 Signalling pathways involved in regulating MMP transcription include p38 MAPK, which can enhance or repress MMP expression in a cell type-dependent manner (Fig. 2). Commonly, MMPs are activated in the pericellular space by other MMPs, including membrane-type MMPs and MMP-3, or by serine proteases like plasmin and chymase. Activated MMPs degrade collagen, elastin, and other ECM proteins, resulting in a modified ECM, often associated with a proinflammatory microenvironment that triggers a shift of endothelial and vascular smooth muscle cells to a more secretory, migratory, proliferative, and senescent phenotype, which contributes to fibrosis, calcification, endothelial dysfunction, and increased intima-media thickness, further impacting on vascular remodelling and arterial stiffness.

The effect that MMPs have on vascular fibrosis in hypertension is not completely elucidated, with both inhibitory and stimulatory modulation observed. 19 This probably relates to activation of different MMP isoforms and downstream signalling pathways. For instance, MMP-1 over-expression attenuates fibrosis, ²⁰ whereas MMP-9 activation potentiates fibrosis and DNA damage.²¹ MMP2 activation leads to stimulation of TGF-β1 signalling; increased vascular smooth muscle cell production of collagens I, II, and III; and increased fibronectin secretion, processes that lead to collagen accumulation in the vascular wall. Although activation of vascular MMP2 and MMP9 in hypertension is associated with collagen accumulation, activation of MMP8 and MMP13 is associated with collagen degradation, processes especially important in arterial wall plaque and plaque rupture.^{22,23} MMP2/MMP9 activation through TGF-β1/ SMAD signalling also induces activation of myofibroblasts and increased infiltration of monocytes/macrophages, leading to oxidative stress, inflammation, and vascular wall injury. Vascular MMP2 and MMP9 are activated by numerous prohypertensive factors, including Ang II, ET-1, and salt, as well as mechanical and physical factors, such as shear stress and pressure. MMP2, MMP7, MMP9, and MMP14 are upregulated by aging. MMP2 activation is increased in aged rat aorta, leading to increased TGF-\$\beta\$1 and SMAD activation.²⁴ Young rats infused with Ang II exhibit increased MMP2 activation with intima-media thickness and vascular fibrosis changes that are typical in old untreated rats.²⁴ The importance of MMPs in vascular fibrosis in aging and hypertension is further evidenced by MMP inhibitors, such as PD166793, which blunted age-associated vascular fibrosis and remodelling in experimental models. 25,26

MMPs are normally inhibited by endogenous inhibitors called TIMPs, of which there are multiple isoforms. Alterations in the balance between ECM MMPs and TIMPs may contribute to the profibrotic phenotype in aging and hypertension. ^{19,24} The 4 TIMP isoforms—TIMP1, TIMP2, TIMP3, and TIMP4—are responsible for the inhibition of > 20 MMPs, and the relationship between MMPs and TIMPs changes with age. For instance, increased MMP2 expression and activity is observed in vessels of old rats and nonhuman primates compared with young counterparts. ^{5,27} Furthermore, TIMPs are downregulated in aged animals with heart failure but not in young animals. ²⁸

Molecular and Cellular Mechanisms of Vascular Fibrosis in Aging and Hypertension

TGF-β/SMAD signalling

The TGF- β superfamily consists of > 40 members that share common sequence elements and structural motifs and includes TGF- β , bone morphogenetic protein, activin, inhibin, and growth differentiation factors. ^{29–32} Disruption of the TGF- β pathway has been implicated in arterial aging and

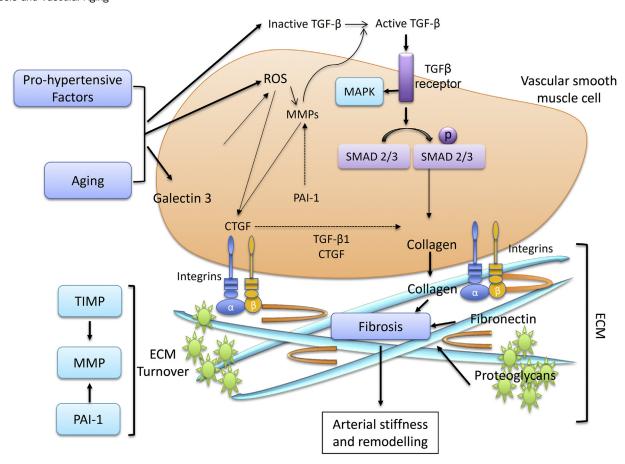


Figure 2. Vascular signalling mediating extracellular matrix (ECM) remodelling, fibrosis, and arterial stiffening in aging and hypertension. Prohypertensive factors and physiological aging promote ECM remodelling through activation of transforming growth factor-β (TGF-β) and subsequently, mitogen-activated protein kinase (MAPK) and SMAD pathways, reactive oxygen species (ROS) production, leading to matrix metalloproteinase (MMP) and connective tissue growth factor (CTGF) activation and upregulation of galectin-3. Subsequently, collagen, fibronectin, and proteoglycan deposition is increased, leading to fibrosis and increased arterial stiffness. PAI, plasminogen activator inhibitor.

vascular fibrosis.²⁹⁻³² Three isoforms (TGF-β1, TGF-β2, and TGF-\(\beta\)3) exist; TGF-\(\beta\)1 is most frequently upregulated in ECM remodelling and fibrosis and is consequently regarded as an important regulator of the ECM. In the vascular system, TGF-\(\beta\)1 is expressed in endothelial cells, vascular smooth muscle cells, myofibroblasts, and adventitial macrophages. Activation of vascular TGF-β1, and its downstream signalling effector SMAD, increases the synthesis of ECM proteins such as fibronectin, collagen, and plasminogen activator inhibitor-1 (PAI-1). 33,34 TGF-β reduces collagenase production and stimulates expression of TIMPS, resulting in excessive matrix accumulation, in part resulting from inhibition of ECM degradation. 35 TGF- β signalling predominantly occurs TGF-β signalling predominantly occurs through the cytoplasmic proteins, SMADs, which translocate to the nucleus and act as transcription factors. The SMAD family comprises receptor-activated SMADs (SMAD2, SMAD3, SMAD5, and SMAD8), inhibitory SMADs (SMAD6, SMAD 7) and common-partner SMADs (SMAD4). SMAD2 and SMAD3 are specific mediators of TGFB/activin pathways, whereas SMAD7 inhibits both BMP and TGF-β/activin signalling. SMAD activation results in increased transcription of many genes involved in ECM formation, including fibronectin, procollagens, PAI-1, and

connective tissue growth factor (CTGF). In vascular smooth muscle cells, overexpression of SMAD7 inhibits TGF- β —induced fibronectin, collagen, and CTGF production. Important non-SMAD pathways implicated in TGF- β profibrotic signalling include extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), p38 MAPK, and phosphoinositide 3-kinase/Akt. MAD translocation to the nucleus can be modulated by Ras-activated ERK1/2. ERK inhibition reduces TGF- β —stimulated SMAD phosphorylation as well as collagen production, suggesting that ERK activation is necessary for an optimal response to TGF- β 1.

Activation of TGF- $\beta1$ and receptor-mediated signalling are increased in the aortic wall with aging and during development of hypertension. Higher Important in the context of these conditions, Ang II, mechanical stress, 34,40 ET-1, 36 and ROS are all elevated and are known to mediate TGF- β activation, with resultant vascular fibrosis. Additionally, MMPs (particularly MMP2 and MMP9) enhance release of TGF- $\beta1$, whereas TGF- $\beta1$ stimulates TIMP, resulting in inhibition of ECM degradation, which further induces ECM accumulation and vascular remodelling and fibrosis. Ang II can activate the SMAD pathway independent of TGF- $\beta1$, with implications for fibrosis. 36,42

Plasminogen activator inhibitor-1

Plasminogen activator inhibitor-1 (PAI-1) is a member of the serine protease inhibitor (serpin) gene family and functions as an inhibitor of the serine proteases, urokinase-type plasminogen activator (uPA), and tissue-type plasminogen activator (tPA). PAI-1 inhibits fibrinolysis and hence regulates dissolution of fibrin and inhibits degradation of the ECM by reducing plasmin generation. PAI-1 normally maintains tissue homeostasis through regulating the activities of uPA, tPA, plasmin, and MMPs. In pathophysiological conditions, PAI-1 upregulation contributes to accumulation of ECM proteins and tissue fibrosis by preventing tissue proteolytic activity and reducing collagen degradation. Together with increased TGFβ1 activity, PAI-1 activity and expression are increased in experimental models of aging and in aged individuals. 43,44 PAI-1 is upregulated in aging-associated pathologic conditions, including hypertension. 45 Increased PAI-1 is also recognized as a biomarker of cellular senescence in aging and hypertension. 46

Connective tissue growth factor

CTGF is a 38-kDa cysteine-rich secreted potent profibrotic factor implicated in fibroblast proliferation, cellular adhesion, and ECM synthesis. CTGF expression in the vasculature is enhanced by several stimuli, including TGF- β 1, tumor necrosis factor- α , and mechanical stress. Ang II—induced vascular fibrosis is mediated by CTGF, and vascular smooth muscle cells treated with CTGF antisense oligonucleotides are protected against agonist-induced ECM protein expression. CTGF may play an important role in arterial aging and vascular fibrosis; a number of experimental models have demonstrated increased levels of CTGF and associated vascular fibrosis with increasing age.

Galectin-3

Galectin-3 (LGALS3) is a 29- to 35-kDa carbohydratebinding lectin expressed on the cell surface of many cell types, including fibroblasts and endothelial and inflammatory cells. It is secreted mainly by activated macrophages, and it is ligand activated by oligosaccharides. Galectin-3 is also activated by other ligands, including glycosylated matrix proteins such as laminin, collagen, elastin, fibronectin, and integrin. The cellular actions of galectin-3 lead to cell proliferation, adhesion, and fibrosis. Galectin-3 has been shown to play an important role in fibrosis and tissue remodelling. In heart failure, plasma galectin-3 levels are increased.⁵¹ In the recent Prevention of Renal and Vascular End-Stage Disease (PREVEND) study in which plasma galectin-3 levels were measured in 7968 individuals, plasma levels correlated positively with increasing age and cardiovascular risk factors, including hypertension. 52 Because of its role in fibrosis, galectin-3 is now considered by many to be an important biomarker of cardiovascular fibrosis. The precise mechanisms through which galectin-3 influences ECM remodelling and fibrosis are still unclear, although activation of the Janus kinase (JAK)/signal transducer and activator of transcription (STAT) and protein kinase C (PKC) pathways, 53,54 as well as oxidative stress and inflammation, have been suggested. In addition, galectin-3 may directly increase production of ECM proteins. In rat vascular smooth muscle cells, over-expression of galectin-3 enhanced aldosterone-induced collagen 1 synthesis, whereas spironolactone or modified citrus pectin (galectin-3 inhibitor) reversed these effects. ⁵⁵ Galectin-3 inhibition also attenuated cardiovascular fibrosis and left ventricular dysfunction in a mouse model of heart failure. ⁵⁶

The Role of Prohypertensive Vasoactive Factors in Vascular Aging and Fibrosis

Many vasoactive factors activate profibrotic pathways, including Ang II, ET-1, and aldosterone (Figs. 2 and 3). Downstream signalling involves activation of redox-sensitive genes and transcription factors, early growth response factor-1, and activation of TGF- β 1, MMPs, galectin-3, and MAP kinases. ⁵⁷⁻⁶¹ The aging vasculature is characterized by increased levels of Ang II, ⁵ angiotensin-converting enzyme, ^{17,31,61} mineralocorticoid receptors, ⁶² and endothelin-converting enzyme-1. ^{63,64} As such, increased levels of these factors, their receptors, and downstream targets could represent an important event during aging that leads to vascular stiffness.

Ang II signalling and vascular fibrosis

The renin-angiotensin-aldosterone system plays a central role in structural and mechanical changes in the vasculature. Ang II acts through activation of 2 receptors—AT1 and AT2_in which AT1 plays a major role in the production of ECM proteins. 65-68 This is highlighted by studies demonstrating that antagonism of Ang II receptors results in decreased fibrosis. 69,70 The precise signalling events involved in Ang II-induced vascular fibrosis are incompletely determined; however, in mesangial cells, TGF-β1 activity is increased by Ang II, an effect not observed when activator protein 1 binding sites or PKC- and p38 MAPK-dependent pathways are inhibited.⁶⁵ In addition, galectin-3 seems to be associated with Ang II-induced fibrosis, and its expression is related to the severity of renal dysfunction in aging; mice subjected to Ang II infusion develop cardiac fibrosis, 1 an effect not observed in galectin-3 knockout animals. Furthermore, cultured fibroblasts exposed to galectin-3 have reduced collagen production and deposition. 60 Ang II—induced activation of p38 MAPK is also associated with the development and progression of fibrosis, commonly observed in aging and hypertension.⁷²⁻⁷⁴ It has been suggested that Ang II induces activity of MMPs and TIMPs and upregulation of CTGF during aging.75

Aldosterone and vascular fibrosis

Accumulating evidence implicates aldosterone as an important pathophysiological mediator in cardiovascular remodelling by promoting vascular hypertrophy, fibrosis, inflammation, and oxidative stress. Evidence from animal models and clinical trials of heart failure and hypertension demonstrate that chronic blockade of mineralocorticoid receptors, through which aldosterone signals, reduces cardiovascular fibrosis. In rats, aldosterone infusion increases aortic media cross-sectional area associated with elevated collagen levels, particularly increased collagen I synthesis. 84,85

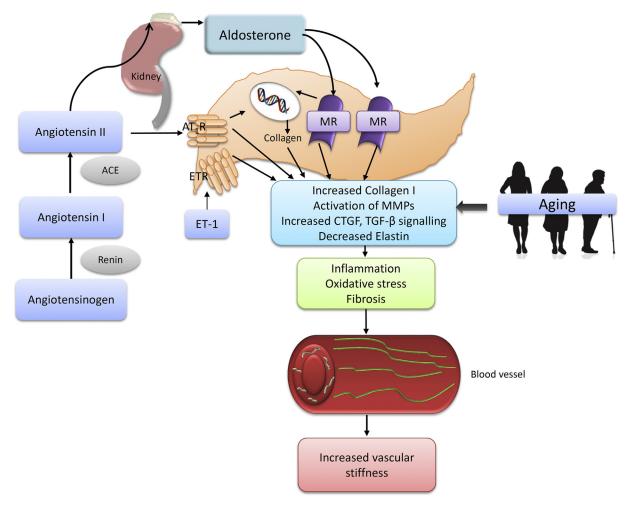


Figure 3. Influence of prohypertensive factors and aging in the development of vascular fibrosis and arterial stiffening. The renin-angiotensinaldosterone system, acting through angiotensin receptor type 1 (AT1R) and mineralocorticoid receptor (MR), and endothelin-1 (ET-1) acting through endothelin receptor (ETR) activate matrix metalloproteinase (MMPs), connective tissue growth factor (CTGF), and transforming growth factor- β (TGF- β) signalling, resulting in inflammation, oxidative stress, and fibrosis, leading to increased arterial stiffness. This process is also induced by ET-1 signalling through ETR, aldosterone signalling through MR, and aging. ACE, angiotensin converting enzyme.

In the context of aging, aldosterone levels have been shown to decline in older age. ^{86,87} This is associated with increased expression of mineralocorticoid receptors in intact vessels, as well as in cultured vascular smooth muscle cells, and has been shown to correlate with markers of vascular fibrosis. ⁶² Whether increased signalling through mineralocorticoid receptors plays a role in vascular fibrosis associated with aging has yet to be confirmed.

ET-1 and vascular fibrosis

ET-1 is a secreted peptide produced primarily in endothelial cells after conversion of preproendothelin to proendothelin and subsequently to mature endothelin, which has potent vasoconstrictor activity. The vascular actions of ET-1 are mediated by 2 distinct endothelin receptor subtypes: the ETA and ETB receptors located on both vascular smooth muscle and endothelial cells. In addition to well-established hypertrophic and mitogenic properties, ET-1 can modulate ECM remodelling by stimulating fibroblast-induced collagen

synthesis. ET-1 stimulates synthesis of collagen through both ETA and ETB receptor subtypes. Reduced cardiac and renal MMP activity and expression has been reported after administration of ETA receptor antagonists. Similarly, treatment with an endothelin antagonist normalizes expression of the collagen I gene and leads to the regression of renal vascular fibrosis and improved survival.

Numerous findings have reported elevated ET-1 levels in healthy older adult humans. ^{94,95} In cultured aortic endothelial cells, ET-1 synthesis is greater in cells obtained from older donors vs young adult donors. ⁹⁶ In Wistar-Kyoto (WKY) rats, aging is associated with a 3.6-fold elevation in kidney ET-1 protein expression in the kidney. In rodent models, dual ETA/ETB receptor antagonism had no effect on the age-associated increase in aortic MMP-2 activity in WKY rats but markedly reduced pro and active MMP-2 activity in aged hypertensive rats, demonstrating that ET-1 may represent an important mediator of vascular stiffness in aging in the presence of other vascular diseases. ⁶³

Conclusions

With aging, the vasculature undergoes structural and functional changes characterized by arterial remodelling, vascular fibrosis, and stiffening, which are processes that are evident in aging and hypertension. Arterial stiffening is common, occurring in > 60% of individuals older than 70 years and is a major independent predictor for serious cardiovascular events. Accordingly, there is a need to understand the fundamental processes that cause vascular stiffness so that mechanism-based therapeutic strategies can be developed to ameliorate or prevent processes of "vascular aging" in hypertension and associated cardiovascular diseases. Arterial stiffening is caused primarily by excessive fibrosis from excessive accumulation of vascular collagen and degradation of elastin. It is a dynamic phenomenon, which initially is an adaptive repair response that is reversible. However, the fibrogenic process is progressive, leading to further worsening of arterial stiffness and fibrosis that gradually extends into the neighbouring interstitial space, causing tissue and organ damage. A number of noninvasive methods are currently available to evaluate large-artery stiffness in the clinical setting, including carotid-femoral PWV. Increased PWV in aging and hypertension reflects increased arterial stiffness and is emerging as a biomarker for cardiovascular risk stratification. Perhaps over the next decade, PWV assessment may become a routine investigation in the clinical tool kit to better predict hypertension and cardiovascular disease.

Funding Sources

This work was supported by grants from the British Heart Foundation (BHF) (RG/13/7/30099). R.M.T. is supported through a BHF Chair (CH/12/4/29762) and R.A.L. is supported by a PhD scholarship from FAPESP-Brazil (2012/12178-6).

Disclosures

The authors have no conflicts of interest to disclose.

References

- World Health Organization, International Society of Hypertension Writing Group. 2003 World Health Organization (WHO)/International Society of Hypertension (ISH) statement on management of hypertension. J Hypertens 2003;21:1983-92.
- Baker PB, Baba N, Boesel CP. Cardiovascular abnormalities in progeria. Case report and review of the literature. Arch Pathol Lab Med 1981;105:384-6.
- Huveneers S, Daemen MJ, Hordijk PL. Between Rho(k) and a hard place: the relation between vessel wall stiffness, endothelial contractility, and cardiovascular disease. Circ Res 2015;116:895-908.
- Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: part I: aging arteries: a "set up" for vascular disease. Circulation 2003;107:139-46.
- Stout LC, Whorton EB Jr, Vaghela M. Pathogenesis of diffuse intimal thickening (DIT) in non-human primate thoracic aortas. Atherosclerosis 1983;47:1-6.
- Lopes RA, Neves KB, Tostes RC, Montezano AC, Touyz RM. Downregulation of nuclear factor erythroid 2-related factor and associated antioxidant genes contributes to redox-sensitive vascular dysfunction in hypertension. Hypertension 2015;66:1240-50.

- AlGhatrif M, Strait JB, Morrell CH, et al. Longitudinal trajectories of arterial stiffness and the role of blood pressure: the Baltimore Longitudinal Study of Aging. Hypertension 2013;62:934-41.
- AlGhatrif M, Lakatta EG. The conundrum of arterial stiffness, elevated blood pressure, and aging. Curr Hypertens Rep 2015;17:1-9.
- Nilsson PM, Boutouyrie P, Cunha P, et al. Early vascular ageing in translation: from laboratory investigations to clinical applications in cardiovascular prevention. J Hypertens 2013;31:1517-26.
- Laurent S, Boutouyrie P. The structural factor of hypertension: large and small artery alterations. Circ Res 2015;116:1007-21.
- Laurent S, Cockcroft J, Van Bortel L, et al. European Network for Noninvasive Investigation of Large Arteries. Expert consensus document on arterial stiffness: methodological issues and clinical applications. Eur Heart J 2006;27:2588-605.
- 12. Van Bortel LM, Laurent S, Boutouyrie P, et al. Artery Society. European Society of Hypertension Working Group on Vascular Structure and Function. European Network for Noninvasive Investigation of Large Arteries. Expert consensus document on the measurement of aortic stiffness in daily practice using carotid-femoral pulse wave velocity. J Hypertens 2012;30:445-8.
- Kotsis V, Stabouli S, Karafillis I, Nilsson P. Early vascular aging and the role of central blood pressure. J Hypertens 2011;29:1847-53.
- Kida Y, Goligorsky MS. Sirtuins, cell senescence, and vascular aging. Can J Cardiol 2016;32:634-41.
- Lakatta EG. The reality of aging viewed from the arterial wall. Artery Res 2013;7:73-80.
- Wang Z, Griffin M. TG2, a novel extracellular protein with multiple functions. Amino Acids 2012;42:939-49.
- Petersen-Jones HG, Johnson KB, Hitomi K, et al. Transglutaminase activity is decreased in large arteries from hypertensive rats compared with normotensive controls. Am J Physiol Heart Circ Physiol 2015;308:H592-602.
- Chakraborti S, Mandal M, Das S, Mandal A, Chakraborti T. Regulation of matrix metalloproteinases: an overview. Mol Cell Biochem 2003;253:269-85.
- Giannandrea M, Parks WC. Diverse functions of matrix metalloproteinases during fibrosis. Dis Model Mech 2014;7:193-203.
- Iimuro Y, Nishio T, Morimoto T, et al. Delivery of matrix metalloproteinase-1 attenuates established liver fibrosis in the rat. Gastroenterology 2003;124:445-58.
- Prakobwong S, Yongvanit P, Hiraku Y, et al. Involvement of MMP-9 in peribiliary fibrosis and cholangiocarcinogenesis via Rac1-dependent DNA damage in a hamster model. Int J Cancer 2010;127:2576-87.
- Newby AC. Dual role of matrix metalloproteinases (matrixins) in intimal thickening and atherosclerotic plaque rupture. Physiol Rev 2005;85:1-31.
- 23. Wang M, Kim SH, Monticone RE, Lakatta EG. Matrix metalloproteinases promote arterial remodeling in aging, hypertension, and atherosclerosis. Hypertension 2015;65:698-703.
- 24. Wang M, Zhao D, Spinetti G, et al. Matrix metalloproteinase 2 activation of transforming growth factor-beta1 (TGF-beta1) and TGF-beta1-type II receptor signalling within the aged arterial wall. Arterioscler Thromb Vasc Biol 2006;26:1503-9.
- Wang M, Zhang J, Telljohann R, et al. Chronic matrix metalloproteinase inhibition retards age-associated arterial proinflammation and increase in blood pressure. Hypertension 2012;60:459-66.
- Zavaczki E, Jeney V, Agarwal A, et al. Hydrogen sulfide inhibits the calcification and osteoblastic differentiation of vascular smooth muscle cells. Kidney Int 2011;80:731-9.

Harvey et al. Fibrosis and Vascular Aging

- Li Z, Froehlich J, Galis ZS, Lakatta EG. Increased expression of matrix metalloproteinase-2 in the thickened intima of aged rats. Hypertension 1999;33:116-23.
- 28. Horn MA, Graham HK, Richards MA, et al. Age-related divergent remodeling of the cardiac extracellular matrix in heart failure: collagen accumulation in the young and loss in the aged. J Mol Cell Cardiol 2012;53:82-90.
- Bonnema DD, Webb CS, Pennington WR, et al. Effects of age on plasma matrix metalloproteinases (MMPs) and tissue inhibitor of metalloproteinases (TIMPs). J Card Fail 2007;13:530-40.
- Wang M, Takagi G, Asai K, et al. Aging increases aortic MMP-2 activity and angiotensin II in nonhuman primates. Hypertension 2003;41: 1308-16
- Wang M, Zhang J, Jiang LQ, et al. Proinflammatory profile within the grossly normal aged human aortic wall. Hypertension 2007;50:219-27.
- Ruiz-Ortega M, Rodriguez-Vita J, Sanchez-Lopez E, Carvajal G, Egido J. TGF-beta signalling in vascular fibrosis. Cardiovasc Res 2007;74: 196-206.
- Douillet CD, Velarde V, Christopher JT, et al. Mechanisms by which bradykinin promotes fibrosis in vascular smooth muscle cells: role of TGF-beta and MAPK. Am J Physiol Heart Circ Physiol 2000;279: H2829-37.
- O'Callaghan CJ, Williams B. Mechanical strain-induced extracellular matrix production by human vascular smooth muscle cells: role of TGFbeta(1). Hypertension 2000;36:319-24.
- Duncan MR, Frazier KS, Abramson S, et al. Connective tissue growth factor mediates transforming growth factor beta-induced collagen synthesis: down-regulation by cAMP. FASEB J 1999;13:1774-86.
- Rodriguez-Vita J, Sanchez-Lopez E, Esteban V, et al. Angiotensin II activates the Smad pathway in vascular smooth muscle cells by a transforming growth factor-beta-independent mechanism. Circulation 2005;111:2509-17.
- Li JH, Huang XR, Zhu HJ, et al. Advanced glycation end products activate Smad signalling via TGF-beta-dependent and independent mechanisms: implications for diabetic renal and vascular disease. FASEB J 2004;18:176-8.
- Gibbons GH, Pratt RE, Dzau VJ. Vascular smooth muscle cell hypertrophy vs. hyperplasia. Autocrine transforming growth factor-beta 1 expression determines growth response to angiotensin II. J Clin Invest 1992;90:456-61.
- Itoh H, Mukoyama M, Pratt RE, Gibbons GH, Dzau VJ. Multiple autocrine growth factors modulate vascular smooth muscle cell growth response to angiotensin II. J Clin Invest 1993;91:2268-74.
- Sucosky P, Balachandran K, Elhammali A, Jo H, Yoganathan AP. Altered shear stress stimulates upregulation of endothelial VCAM-1 and ICAM-1 in a BMP-4- and TGF-beta1-dependent pathway. Arterioscler Thromb Vasc Biol 2009;29:254-60.
- Rhyu DY, Yang Y, Ha H, et al. Role of reactive oxygen species in TGFbeta1-induced mitogen-activated protein kinase activation and epithelialmesenchymal transition in renal tubular epithelial cells. J Am Soc Nephrol 2005;16:667-75.
- Russo I, Frangogiannis NG. Diabetes-associated cardiac fibrosis: cellular effectors, molecular mechanisms and therapeutic opportunities. J Mol Cell Cardiol 2016;90:84-93.
- 43. Takeshita K, Yamamoto K, Ito M, et al. Increased expression of plasminogen activator inhibitor-1 with fibrin deposition in a murine model of aging, "Klotho" mouse. Semin Thromb Hemost 2002;28:545-54.

- Hashimoto Y, Kobayashi A, Yamazaki N, et al. Relationship between age and plasma t-PA, PA-inhibitor, and PA activity. Thromb Res 1987;46:625-33.
- Yamamoto K, Takeshita K, Saito H. Plasminogen activator inhibitor-1 in aging. Semin Thromb Hemost 2014;40:652-9.
- 46. Vlachopoulos C, Xaplanteris P, Aboyans V, et al. The role of vascular biomarkers for primary and secondary prevention. A position paper from the European Society of Cardiology Working Group on peripheral circulation: endorsed by the Association for Research into Arterial Structure and Physiology (ARTERY) Society. Atherosclerosis 2015;241:507-32.
- Oemar BS, Luscher TF. Connective tissue growth factor. Friend or foe? Arterioscler Thromb Vasc Biol 1997;17:1483-9.
- Ruperez M, Lorenzo O, Blanco-Colio LM, et al. Connective tissue growth factor is a mediator of angiotensin II-induced fibrosis. Circulation 2003;108:1499-505.
- van Almen GC, Verhesen W, van Leeuwen RE, et al. MicroRNA-18 and microRNA-19 regulate CTGF and TSP-1 expression in age-related heart failure. Aging Cell 2011;10:769-79.
- Bigot A, Jacquemin V, Debacq-Chainiaux F, et al. Replicative aging down-regulates the myogenic regulatory factors in human myoblasts. Biol Cell 2008;100:189-99.
- Van Kimmenade RR, Januzzi JL, Ellinor PT, et al. Utility of aminoterminal pro-brain natriuretic peptide, galectin-3, and apelin for the evaluation of patients with acute heart failure. J Am Coll Cardiol 2006;48:1217-24.
- De Boer R, van Veldhuisen D, Gansevoort R, et al. The fibrosis marker galectin-3 and outcome in the general population. J Intern Med 2012;272:55-64.
- Koopmans SM, Bot FJ, Schouten HC, Janssen J, van Marion A. The involvement of Galectins in the modulation of the JAK/STAT pathway in myeloproliferative neoplasia. Am J Blood Res 2012;2:119-27.
- Song X, Qian X, Shen M, et al. Protein kinase C promotes cardiac fibrosis and heart failure by modulating galectin-3 expression. Biochim Biophys Acta 2015;1853:513-21.
- Calvier L, Miana M, Reboul P, et al. Galectin-3 mediates aldosteroneinduced vascular fibrosis. Arterioscler Thromb Vasc Biol 2013;33:67-75.
- Vergaro G, Prud'homme M, Fazal L, et al. Inhibition of galectin-3 pathway prevents isoproterenol-induced left ventricular dysfunction and fibrosis in mice. Hypertension 2016;67:606-12.
- Mendoza-Torres E, Oyarzun A, Mondaca-Ruff D, et al. ACE2 and vasoactive peptides: novel players in cardiovascular/renal remodeling and hypertension. Ther Adv Cardiovasc Dis 2015;9:217-37.
- Martinez-Martinez E, Calvier L, Fernandez-Celis A, et al. Galectin-3 blockade inhibits cardiac inflammation and fibrosis in experimental hyperaldosteronism and hypertension. Hypertension 2015;66:767-75.
- Messaoudi S, He Y, Gutsol A, et al. Endothelial Gata5 transcription factor regulates blood pressure. Nat Commun 2015;6:8835.
- Yu L, Ruifrok WP, Meissner M, et al. Genetic and pharmacological inhibition of galectin-3 prevents cardiac remodeling by interfering with myocardial fibrogenesis. Circ Heart Fail 2013;6:107-17.
- Neves K, Nguyen Dinh Cat A, Lopes RA, et al. Chemerin regulates crosstalk between adipocytes and vascular cells through Nox. Hypertension 2015;66:657-66.
- 62. Krug AW, Allenhofer L, Monticone R, et al. Elevated mineralocorticoid receptor activity in aged rat vascular smooth muscle cells promotes a proinflammatory phenotype via extracellular signal-regulated kinase 1/2

- mitogen-activated protein kinase and epidermal growth factor receptor-dependent pathways. Hypertension 2010;55:1476-83.
- 63. Spiers JP, Kelso EJ, Siah WF, et al. Alterations in vascular matrix metalloproteinase due to ageing and chronic hypertension: effects of endothelin receptor blockade. J Hypertens 2005;23:1717-24.
- Park JB, Schiffrin EL. Cardiac and vascular fibrosis and hypertrophy in aldosterone-infused rats: role of endothelin-1. Am J Hypertens 2002;15:164-9.
- Weigert C, Brodbeck K, Klopfer K, Häring H, Schleicher E. Angiotensin II induces human TGF-β1 promoter activation: similarity to hyperglycaemia. Diabetologia 2002;45:890-8.
- 66. Montezano AC, Paravicini TM, Chignalia AZ, et al. Nicotinamide adenine dinucleotide phosphate reduced oxidase 5 (nox5) regulation by angiotensin ii and endothelin-1 is mediated via calcium/calmodulin-dependent pathways in human endothelial cells. Circ Res 2010;106:1363-73.
- Qi G, Jia L, Li Y, et al. Angiotensin II infusion—induced inflammation, monocytic fibroblast precursor infiltration, and cardiac fibrosis are pressure dependent. Cardiovasc Toxicol 2011;11:157-67.
- Carver KA, Smith TL, Gallagher PE, Tallant E. Angiotensin-(1-7) prevents angiotensin II-induced fibrosis in cremaster microvessels. Microcirculation 2015;22:19-27.
- Ishidoya S, Morrissey J, McCracken R, Reyes A, Klahr S. Angiotensin II receptor antagonist ameliorates renal tubulointerstitial fibrosis caused by unilateral ureteral obstruction. Kidney Int 1995;47:1285-94.
- Ruiz-Ortega M, Gonzalez S, Seron D, et al. ACE inhibition reduces proteinuria, glomerular lesions and extracellular matrix production in a normotensive rat model of immune complex nephritis. Kidney Int 1995;48:1778-91.
- AbouEzzeddine OF, Haines P, Stevens S, et al. Galectin-3 in heart failure with preserved ejection fraction: a RELAX trial substudy (Phosphodiesterase-5 Inhibition to Improve Clinical Status and Exercise Capacity in Diastolic Heart Failure). JACC Heart Fail 2015;3:245-52.
- Li Z, Li J, Bu X, et al. Age-induced augmentation of p38 MAPK phosphorylation in mouse lung. Exp Gerontol 2011;46:694-702.
- Wu Z, Yu Y, Liu C, et al. Role of p38 mitogen-activated protein kinase in vascular endothelial aging: interaction with arginase-II and S6K1 signalling pathway. Aging (Albany NY) 2015;7:70-81.
- 74. Hsieh C, Papaconstantinou J. The effect of aging on p38 signalling pathway activity in the mouse liver and in response to ROS generated by 3-nitropropionic acid. Mech Ageing Dev 2002;123:1423-35.
- 75. Pons M, Cousins SW, Alcazar O, Striker GE, Marin-Castaño ME. Angiotensin II—induced MMP-2 activity and MMP-14 and basigin protein expression are mediated via the angiotensin II receptor type 1—mitogenactivated protein kinase 1 pathway in retinal pigment epithelium: implications for age-related macular degeneration. Am J Pathol 2011;178:2665-81.
- Nakai K, Kawato T, Morita T, et al. Angiotensin II induces the production of MMP-3 and MMP-13 through the MAPK signalling pathways via the AT 1 receptor in osteoblasts. Biochimie 2013;95:922-33.
- Yaghooti H, Firoozrai M, Fallah S, Khorramizadeh M. Angiotensin II induces NF-κB, JNK and p38 MAPK activation in monocytic cells and increases matrix metalloproteinase-9 expression in a PKC-and Rho kinase-dependent manner. Braz J Med Biol Res 2011;44:193-9.
- Oelusarz A, Nichols LA, Grunz-Borgmann EA, et al. Overexpression of MMP-7 increases collagen 1A2 in the aging kidney. Physiol Rep 2013;1: e00090.
- Sangaralingham SJ, Wang BH, Huang L, et al. Cardiorenal fibrosis and dysfunction in aging; imbalance in mediators and regulators of collagen. Peptides 2016;76:108-14.

- Odenbach J, Wang X, Cooper S, et al. MMP-2 mediates angiotensin IIinduced hypertension under the transcriptional control of MMP-7 and TACE. Hypertension 2011;57:123-30.
- Sakurabayashi-Kitade S, Aoka Y, Nagashima H, et al. Aldosterone blockade by spironolactone improves the hypertensive vascular hypertrophy and remodeling in angiotensin II overproducing transgenic mice. Atherosclerosis 2009;206:54-60.
- Sontia B, Montezano ACI, Touyz RM. Downregulation of renal TRPM7 and increased cardiovascular and renal inflammation and fibrosis in aldosterone-infused mice—effects of magnesium supplementation. Hypertension 2008;51:915-21.
- 83. Callera GE, Yogi A, Briones AM, et al. Vascular proinflammatory responses by aldosterone are mediated via c-Src trafficking to cholesterol-rich microdomains: role of PDGFR. Cardiovasc Res 2011;91:720-31.
- 84. Savoia C, Touyz RM, Schiffrin EL. Selective mineralocorticoid receptor blocker eplerenone reduces resistance artery stiffness in hypertensive patients. Hypertension 2008;51:432-9.
- 85. Briones AM, Nguyen Dinh Cat A, Callera GE, et al. Adipocytes produce aldosterone through calcineurin-dependent signalling pathways: implications in diabetes mellitus-associated obesity and vascular dysfunction. Hypertension 2012;59:1069-78.
- 86. Weidmann P, Beretta-Piccoli C, Ziegler WH, et al. Age versus urinary sodium for judging renin, aldosterone, and catecholamine levels: studies in normal subjects and patients with essential hypertension. Kidney Int 1978;14:619-28.
- 87. Hegstad R, Brown RD, Jiang N, et al. Aging and aldosterone. Am J Med 1983;74:442-8.
- Horstmeyer A, Licht C, Scherr G, Eckes B, Krieg T. Signalling and regulation of collagen I synthesis by ET-1 and TGF-β1. FEBS J 2005;272:6297-309.
- Hafizi S, Wharton J, Chester AH, Yacoub MH. Profibrotic effects of endothelin-1 via the ETA receptor in cultured human cardiac fibroblasts. Cell Physiol Biochem 2004;14:285-92.
- Park JB, Schiffrin EL. ET(A) receptor antagonist prevents blood pressure elevation and vascular remodeling in aldosterone-infused rats. Hypertension 2001;37:1444-9.
- Ammarguellat FZ, Gannon PO, Amiri F, Schiffrin EL. Fibrosis, matrix metalloproteinases, and inflammation in the heart of DOCA-salt hypertensive rats: role of ET(A) receptors. Hypertension 2002;39:679-84.
- 92. Ebihara I, Nakamura T, Tomino Y, Koide H. Effect of a specific endothelin receptor A antagonist and an angiotensin-converting enzyme inhibitor on glomerular mRNA levels for extracellular matrix components, metalloproteinases (MMP) and a tissue inhibitor of MMP in aminonucleoside nephrosis. Nephrol Dial Transplant 1997;12:1001-6.
- Boffa JJ, Tharaux PL, Dussaule JC, Chatziantoniou C. Regression of renal vascular fibrosis by endothelin receptor antagonism. Hypertension 2001;37:490-6.
- 94. Komatsumoto S, Nara M. Changes in the level of endothelin-1 with aging. Nihon Ronen Igakkai Zasshi 1995;32:664-9.
- Donato AJ, Gano LB, Eskurza I, et al. Vascular endothelial dysfunction with aging: endothelin-1 and endothelial nitric oxide synthase. Am J Physiol Heart Circ Physiol 2009;297:H425-32.
- Tokunaga O, Fan J, Watanabe T, et al. Endothelin. Immunohistologic localization in aorta and biosynthesis by cultured human aortic endothelial cells. Lab Invest 1992;67:210-7.