Research Article

Formulation and Optimization of Complementary Food Based on Its Nutritional and Antinutritional Analysis

Elsabet Getachew Aynalem and Ramesh Duraisamy

Department of Chemistry (Industrial Chemistry Division: Food and Sugar Technology), College of Natural and Computational Sciences, Arba Minch University, Arba Minch, Ethiopia

Correspondence should be addressed to Ramesh Duraisamy; ramesh.duraisamy@amu.edu.et

Received 23 July 2022; Revised 18 September 2022; Accepted 30 September 2022; Published 17 October 2022

Academic Editor: Ravi Pandiselvam

Copyright © 2022 Elsabet Getachew Aynalem and Ramesh Duraisamy. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

This study focuses on formulating and optimizing complementary foods using oat, yellow maize, chickpea, avocado pulp powder, and sugarcane Jaggery to obtain nutritious infant food. Different proportions of the composite food were optimized using Minitab v.19 software upon mixture design by keeping the constant amount of Jaggery (10%). The formulated composite foods had a better proximate composition, minerals, vitamins, and lower amounts of antinutrients. Most attributes have shown significant differences (p < 0.05) among those formulations. The better-optimized formulation is selected from nutritional and antinutritional values through overlaid contour design. The study concludes the optimal complementary food composition ratio: oat (40 g), chickpea (25.81 g), yellow maize (13.78 g), avocado powder (10.4 g), and Jaggery (10 g). Thus, the proposed formulated product becomes nutritious complementary food that may help children's and medium-age hold communities.

1. Introduction

The World Health Organization (WHO) describes complementary feeding as a process starting when breast milk alone is no longer sufficient to meet the nutritional requirements of infants. Therefore, other foods and liquids are needed, along with breast milk [1]. There is a requirement by which complementary foods should include high energy density, balanced protein composition (containing all essential amino acids), and micronutrients like vitamins and minerals (iron, folic acid, and calcium) [2]. Cereals and grains are common foods in most human diets in developed and developing countries, providing the main proportion of dietary energy and nutrients. It contains 75% carbohydrates, primarily starches, and about 6-15% protein, contributing to more than 50% of the energy supply [3, 4].

In Ethiopia, homemade baby foods are prepared primarily from cereal grains, starting around six months. It is ready in the form of solid and liquid (porridge and gruel). These complementary foods typically lack essential amino acids (protein) and micronutrients, which lead to different nutrient deficiency diseases. They are high in antinutrients, which affect the bioavailability of essential minerals. Furthermore, these cereal-based complementary foods have high fiber and less nutrient density, which immediately fill infants' stomachs since high-fiber foods are bulky and low in caloric density [5].

Avocado is a super nutritious fruit that contains essential nutrients for infants but is highly susceptible to microbial spoilage and stored for a short period. Due to this, it needs a proper value addition process like chemical treatment and drying at a lower temperature. Such studies are crucial to coming up with tangible means like blending and compiling with various foodstuffs to produce nutritional rich and good sensory value food products that can promote appetite and habit of consumption. Sugarcane contributes to 90% of the world's sweetener production. Although sugar is added in processing such foods, there is minimal literature on the case of using Jaggery as a sweetener in complementary foods [6]. Worldwide, More than 70% of Jaggery produced from India by processed the sugarcane [4]. Most complementary foods in the market are costly and cannot be economically viable for low-income families [7]. Thus, this research intended to formulate nutritious food products using harmless sweeteners and value-added food products from readily available cereal grains, legumes, fruit, and Jaggery to help overcome nutrient deficiency problems by filling the nutrient gap required by infants. Furthermore, using Jaggery instead of table sugar in the proper amount helps to obtain essential nutrients in addition to its sweetness.

2. Materials and Methods

2.1. Raw Material Selection. Raw materials were selected based on Codex Alimentarius guidelines [8]. Accordingly, the blend was composed of cereal and fruit-based ingredients widely used for preparing complementary food. Ingredients are used in this study based on their nutrients as follows: oat (carbohydrates-rich), chickpea (protein-rich), yellow maize (vitamin-rich), and avocado (fat-rich). Additionally, Jaggery was used as a sweetener in addition to supplementing micronutrients and improving the sensory qualities of composite blends. The components were packed in air-tight polythene bags and stored in a laboratory at room temperature until needed for processing.

2.2. Raw Material Preparation

2.2.1. Oat Processing. The oats were cleaned using air aspiration. It was dried in an oven at 40°C, then dehulled and heated at 60°C five hours to develop flavor and deactivate microorganisms. Finally, grinding and sieving were performed using 250 μ m to get fine particles [9].

2.2.2. Yellow Maize Processing. Yellow maize was winnowed, sorted, washed with tap water (3:1 water to sample ratio), and soaked for 24 hours at room temperature 25°C. It was removed from the water, washed thrice with water, and dried in an oven maintained at 60°C until <2% free from moisture [10].

2.2.3. Chickpea Processing. It was hand-sorted and soaked (3:1 water to sample ratio) for 24 hours at room temperature (25° C). It was removed from the water, washed thrice with water, and dried in an oven maintained at 60°C until it was free from moisture (<2%). Dehulling and air aspiration was undertaken for removing the chickpea outer-layer shield. followed by grinding the pretreated chickpea using mortar and pestle [11].

2.2.4. Avocado Powder Processing. Pure, fully ripe, and undamaged avocado fruit samples were selected and washed with pure water by applying appropriate soap. It was peeled and sliced using a stainless steel knife. Blanching was performed (under soaking) at 80°C with lemon juice (with 35% citric acid, which was used as an antibrowning agent) for 4 minutes, drained, and placed on aluminum trays. It was dried using an oven dryer at 45°C for 24 hours and packed using moisture-free polyethylene bag-appropriate packing material [12].

2.2.5. Jaggery Processing. Sugarcane Jaggery was prepared using a three-step process: juice extraction, clarification, and concentration. Fresh sugarcane was collected from cultivation land in Mirab-Abaya, which is near the study area (Arba Minch town). The samples were washed with pure water and peeled using a knife, ground using a juice extractor, and extracted the juice. Filtration was followed by waiting (five minutes) to remove heavy impurities and fine bagasse after sedimentation. Clarification was started during the boiling stage at 75°C (to make hot liming) by adding lime (10% calcium oxide) and filtered again after cooling to remove impurities in the form of scum. The clarified juice was heated on a hot plate to raise the temperature with continuous stirring and monitored using a thermometer [13]. Through constant boiling (as with increasing the temperature gradually from 100 to 118°C), the brownish foam was observed at the top and disappeared with uniform boiling at >110°C, which gave a golden color. When the temperature reaches 105°C, the juice starts frothing. Then, the heat is carefully regulated to prevent caramelization by employing a thermostat. The syrup was constantly stirred to avoid charring and to drop over the pan's side. The concentrated juice was removed from the thermostatic hot plate, poured into aluminum foil, and molded with desired shape [14].

2.3. Formulation of Complementary Foods Using Experimental Design. Different mixing ratios prepared the formulation of complementary foods through the Minitab version 19 mixture design model. A mixture design is applicable when the response depends on the component ranges of the mixture; 17 different compositions (shown in Table 1) helped to select the best optimum compositions through other food quality parameters. The preliminary study was conducted, and based on those results, the constraints used are 40-44 g/100 g oat, 15-33 g/100 g chickpea, 12-23 g/100 g yellow maize, and 5-11 g/100 g avocado powder of upper and lower constraints with a constant amount of Jaggery (10 g/100 g) with the total amount of formulation as 100 g.

2.4. Nutrient Analysis of Raw Materials and Complementary Foods. The proximate compositional analysis, including percentages of fat, fiber, protein, carbohydrate, moisture, ash, and energy, was determined by [15] methods. Graphite furnace atomic absorption spectroscopy was used to estimate the mineral concentrations in the food samples. The β -carotene content was determined using a UV/Vis spectrophotometer by measuring the absorbance of the extract and the blank solution at 450 nm. Using the Beers Lambert law from measured absorbance data, its concentration was calculated in mg·l⁻¹ [16].

Total phenol was determined by extracting the phenolic compound from the sample using methanol [17]. Total phenolic contents were expressed as gallic acid equivalents (GAE) in mg/100 g GAE (at 765 nm) using a UV/Vis spectrophotometer.

Vitamin C was determined by titration by preparing a standard vitamin C solution and using standard iodine

| Run order | Oat (g) | Chickpea (g) | Yellow maize (g) | Avocado powder (g) |
|-----------|---------|--------------|------------------|--------------------|
| CF1 | 40 | 22 | 23 | 5 |
| CF2 | 41.4 | 28.8 | 13.3 | 6.5 |
| CF3 | 45 | 22 | 12 | 11 |
| CF4 | 43.8 | 23.4 | 13.3 | 9.5 |
| CF5 | 40 | 27 | 12 | 11 |
| CF6 | 48 | 25 | 12 | 5 |
| CF7 | 50.1 | 22.1 | 12.6 | 5.2 |
| CF8 | 53.6 | 17.3 | 12.8 | 6.3 |
| CF9 | 55 | 18 | 12 | 5 |
| CF10 | 46.8 | 23.4 | 13.3 | 6.5 |
| CF11 | 40 | 33 | 12 | 5 |
| CF12 | 50 | 15 | 14 | 11 |
| CF13 | 55 | 15 | 15 | 5 |
| CF14 | 48.6 | 20.6 | 12.6 | 8.2 |
| CF15 | 51 | 22 | 12 | 5 |
| CF16 | 52.1 | 16.6 | 13.6 | 7.7 |
| CF17 | 48.8 | 20.5 | 15.1 | 5.6 |
| Levels: | 40-55 g | 15-33 g | 12-23 g | 5-11 g |

TABLE 1: Mixture design for the formulation of complementary foods (CF).

TABLE 2: Full cubic regression model ANOVA *p* values and model coefficients for proximate composition of formulated complementary food samples.

| Source | Moisture | Ash | Protein | Fat | Fiber | СНО | Energy |
|-----------------------------|----------|--------|---------|--------|--------|--------|--------|
| Linear | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.002 | ≤0.001 | ≤0.001 |
| Special cubic | ≤0.001 | 0.001 | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 |
| Oat*chickpea*maize | ≤0.001 | 0.416 | ≤0.001 | ≤0.001 | 0.393 | ≤0.001 | ≤0.001 |
| Oat*chickpea*avocado powder | ≤0.001 | 0.692 | ≤0.001 | ≤0.001 | 0.873 | ≤0.001 | ≤0.001 |
| Oat*maize*avocado powder | 0.046 | 0.047 | 0.022 | 0.048 | 0.987 | 0.004 | 0.264 |
| Chickpea*maize*avocado | ≤0.001 | 0.234 | ≤0.001 | ≤0.001 | 0.829 | ≤0.001 | 0.005 |
| R ² (adjusted) | 0.875 | 0.864 | 0.998 | 0.995 | 0.976 | 0.988 | 0.992 |

TABLE 3: ANOVA p values for full cubic regression model for Ca, Fe, Zn, Mg, Mn, and Cu.

| Source | Ca | Fe | Zn | Mg | Mn | Cu |
|-------------------------------|--------|--------|--------|--------|--------|-------------|
| Linear | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.004 |
| Quadratic | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.206 | ≤ 0.001 |
| Special cubic | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.096 | 0.022 |
| Oat*chickpea*maize | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.328 | 0.125 |
| Oat* chickpea* avocado powder | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.349 | 0.099 |
| Oat*maize*avocado powder | 0.036 | ≤0.001 | ≤0.001 | 0.574 | 0.329 | 0.080 |
| Chickpea*maize*avocado powder | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.292 | 0.086 |
| Full cubic | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.009 |
| R ² (adjusted) | 0.992 | 0.992 | 0.982 | 0.977 | 0.330 | 0.872 |

solution as a titrant by adding starch indicator until the observation of blue color [18]. In all cases, sample preparation was processed based on a method designated by the AOAC method [15].

2.5. Analysis of Antinutrients of Formulated Complementary Food. Phytate content was determined as described by [19]. Follwed by extraction, filtration, titrated the sample solution with standard iron (III) chloride solution using ammonium

| Source | Phytate (mg/100 g) | Tannin (mg/100g) | Oxalate (mg/100 g) | eta-Carotene (mg) | Vit. C (mg) | Polyphenol (mg/100 g) |
|-------------------------------|--------------------|------------------|--------------------|-------------------|-------------|-----------------------|
| Linear | ≤0.001 | 0.002 | ≤0.001 | ≤0.001 | 0.453 | ≤0.001 |
| Quadratic | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.106 | ≤0.001 |
| Special cubic | ≤0.001 | ≤0.001 | ≤0.001 | ≤0.001 | 0.188 | ≤0.001 |
| Oat*chickpea*maize | ≤0.001 | 0.002 | ≤0.001 | ≤0.001 | 0.424 | 0.728 |
| Oat*chickpea*avocado powder | 0.008 | 0.001 | ≤0.001 | ≤0.001 | 0.312 | 0.841 |
| Oat*maize*avocado powder | ≤0.001 | 0.005 | ≤0.001 | ≤0.001 | 0.058 | 0.772 |
| Chickpea*maize*avocado powder | ≤0.001 | 0.001 | ≤0.001 | ≤0.001 | 0.173 | 0.991 |
| Full cubic | ≤0.001 | 0.001 | ≤0.001 | ≤0.001 | 0.030 | 0.871 |
| R^2 (adjusted) | 0.998 | 0.890 | 0.996 | 0.993 | 0.893 | 0.905 |

| 5 | |
|---|--|
| a | |
| nt | |
| ne | |
| ler | |
| omp | |
| Ä | |
| ö | |
| sd | |
| are | |
| g | |
| Ľ. | |
| μ | |
| 0 | |
| 0 | |
| ler | |
| hd | |
| lyp | |
| g | |
| σ | |
| , anc | |
| | |
| Ë. | |
| itamins | |
| Ita | |
| 5 | |
| ts, | |
| Sn: | |
| Ľ. | |
| Ħ | |
| tin | |
| ц | |
| r a | |
| s for | |
| S | |
| jų, | |
| Ğ | |
| Æ | |
| oe | |
| Ū. | |
| e. | |
| 0 | |
| lod | |
| В | |
| q m | |
| В | |
| q m | |
| lues and m | |
| s and m | |
| p values and m | |
| A p values and m | |
| A p values and m | |
| NOVA p values and m | |
| ANOVA p values and m | |
| I ANOVA p values and m | |
| del ANOVA p values and m | |
| odel ANOVA p values and m | |
| model ANOVA p values and m | |
| odel ANOVA p values and m | |
| model ANOVA p values and m | |
| ession model ANOVA p values and m | |
| egression model ANOVA p values and m | |
| gression model ANOVA p values and m | |
| c regression model ANOVA p values and m | |
| egression model ANOVA p values and m | |
| l cubic regression model ANOVA p values and m | |
| ull cubic regression model ANOVA p values and m | |
| l cubic regression model ANOVA p values and m | |
| 4: Full cubic regression model ANOVA p values and m | |
| ${\rm E}$ 4: Full cubic regression model ANOVA p values and m | |
| ${\rm E}$ 4: Full cubic regression model ANOVA p values and m | |
| ${\rm E}$ 4: Full cubic regression model ANOVA p values and m | |
| ABLE 4: Full cubic regression model ANOVA p values and m | |

thiocynate as an indicator. Then, end point (persistence of golden color for 5 minutes) was noticed. Phytate content was calculated using equation (1):

Phytic acid = Titer value
$$\times$$
 (0.00195) \times 1.19 \times 100, (1)

where 0.00195 is the weight of FeCl₃.

Tannin was determined spectrophotometrically at 500 nm as described by [20]. Using the tannic acid (Merck chemicals, India, with 90% purity) solution standard, the calibration curve was used to determine the concentration [21].

Oxalate was determined by using the titration method using $KMnO_4$ as a titrant after extraction [15]. Titration was performed until observation of blue color and persisted for at least one minute [22] and calculated as

$$\text{\%Oxalate} = \left(\frac{\text{Titer value}}{W}\right) \times 0.06303 \times 100.$$
(2)

The relationship, that is, 1 mL of KMnO_4 solution = 0.006303 g oxalate, was used, where titer is the volume of KMnO_4 consumed and W is the weight of the sample.

2.6. Statistical Analysis. The results of experimental designs of the proportions of models described by each response were validated, and the analysis of variance (ANOVA) helps to evaluate each response, the effect of each factor, and interactions among factors.

3. Results and Discussion

3.1. Optimization of Composite Complementary Foods by Its Proximate Composition. The proximate compositions of composite food samples' such as moisture, ash, protein, fat, fiber, carbohydrate contents, and gross energy were analyzed. ANOVA has been used, and characteristics and quantities evaluated the variants. Amongst, p-values and predicted R^2 values are used and analyzed the responses. Generally, if the *p* value is less than alpha (<0.05) which is more significant, then the model of the parameter or interaction can be considered statistically significant. Tables 2 and 3 summarize the ANOVA p values of all the quantities of responses, such as proximate composition, antinutrients, vitamins, and minerals. The full cubic with special cubic models was used for the statistical data analysis of responses. These results were obtained from mixture components as model terms, and the mixture regression design model was selected as a model-fitting method [23].

Table 2 reveals that the changes in proximate compositional of samples to moisture, protein, fat, and carbohydrate (CHO) contents were found to be significantly influenced (p < 0.05) by the blending of the constituents. It shows a statistically significant (p < 0.05) relationship between the oat, chickpea, maize, and avocado pulp powder blending ratios. The other interactions had also shown significant changes. Still, 3-way interaction between oats, maize, and avocado powder has the least nonsignificant, which is proved by their p values (0.987 in fiber and 0.264 in energy) in this case. But,

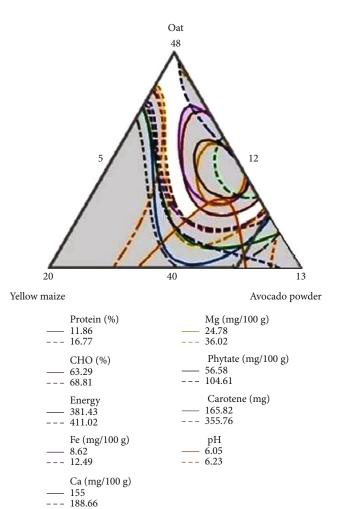


FIGURE 1: Overlaid contour plots that show the sweet spot. Notes: the white area shows the "sweet spot" that optimizes the response variables listed in the respective legends.

the overall selected model was found to a significant (p < 0.05), with the adjusted R^2 values that ranged from 0.875 to 0.998 (shown in Table 2).

The cubic regression ANOVA results (Table 3) for this case also reveal that in copper and manganese, all interactions have not shown statistically significant difference (p > 0.05). It is confirmed with a lower R^2 adjusted value, but iron, calcium, zinc, and magnesium in all cases of interactions show a significant difference (p value < 0.05) with R^2 adjusted value from 0.977 to 0.992.

Table 4 describes that the results of phytate, oxalate, and β -carotene fit very well (R^2 – adjusted > 0.95). It is confirmed that it provides a reasonably good explanation of the relationship between the independent factors (food components) and the responses of the processing of blending themselves. R^2 values of polyphenol, tannin, and vitamin C were found to be 0.905, 0.890, and 0.893, respectively. Similarly, the cubic models for vitamin C, polyphenol, and tannin also fit the data well because the adjusted R^2 is greater than 0.80. On the other hand, for polyphenol and vitamin

| Ontimized feed and in a | | Comp | Overall composite desirability | | | |
|-------------------------|---------|----------|--------------------------------|----------------|---------|--------------------------------|
| Optimized food coding | Oat | Chickpea | Yellow maize | Avocado powder | Jaggery | Overall composite desirability |
| F-1 | 40 | 31.2262 | 12.9589 | 5.81491 | 10.00 | 1.000000 |
| F-2 | 40 | 25.8148 | 13.7778 | 10.4074 | 10.00 | 0.908463 |
| F-3 | 49.3036 | 15 | 14.6964 | 11 | 10.00 | 1.000000 |
| F-4 | 40.9091 | 31.5989 | 12.2789 | 5.21306 | 10.00 | 1.000000 |

TABLE 5: Response optimization using Minitab D-optimizer for formulated food.

C content, except for linear and quadratic models, all three interactions (cubic model) do not fit the data well (p > 0.05).

3.2. Optimal Responses Based on Contour Plate. In the contour plot (Figure 1), the white "sweet spot" that optimizes the responses was determined using the researchers' lower and upper goals for a response [23]. For optimization, resolved responses that show significant differences in the blends were considered. The overlaid contour plot design (Figure 1) for proximate, antinutrient, vitamins, and minerals was obtained by considering significant values (p < 0.05) to find the optimization region using the contour plot design.

The overlaid contour plot was drawn using the protein, CHO, energy, Fe, Ca, Mg, phytate, and pH. All these responses were used and found the optimal region of food composnents of the current study.

3.3. Response Optimization Using D-Optimizer. A constrained D-optimal mixture design was used and predicted the best six optimum blending ratios of the food mixture components (mixture ratio proposed ranged in all optimum mixture: oat: 40-49 g/100 g, chickpea: 15-31.77 g/100 g, maize: 12.17-21.6 g/100 g), and avocado powder: 5-11 g/ 100 g with appropriate responses of each food quality parameters, which is presented in Table 5.

Different parameters were analyzed for the bestoptimized product to compare the predicted and actual values. The obtained experimental result was parallel with the predicted value. From the proximate compositional analysis, protein, CHO, and fat contents were analyzed and received 15.12 g, 63.67 g, and 10.32 g, respectively, which is very close to the predicted value. The result is within the WHO/FAO standard limit. The experimental result shows the mineral composition for Ca, Fe, Zn, and Mg, 169.29 mg, 11.12 mg, 3.84 mg, and 32.48 mg, respectively, which approaches the predicted value. From the mineral content, Zn and Mg are within the WHO recommended limit, but iron and calcium need minor modification to reach the recommended levels. The optimized product's antinutrient composition (phytate, oxalate, and tannin) and β -carotene show a close relationship with the predicted value, which is an acceptable limit. Thus, the optimizer gives the predicted response congruous with the actual value recommended for the formulation of infant foods to reach the nutrient requirement.

The results are feasible to develop a nutritious complementary food by compositing oats, chickpea, yellow maize, avocado powder, and Jaggery. The optimal mixtures of 4049 g/100 g, chickpea: 15-31.77 g/100 g, yellow maize: 12.17-21.6 g/100 g, and avocado powder: 5-11 g/100 g with 10 g/100 g Jaggery enhance significant improvement in the composite food nutritional quality which can contribute to preventing malnutrition in infant food.

4. Conclusions

This study was carried out to formulate complementary food to meet the nutritional needs of infants. It was done by mixing readily available raw materials (oat, chickpea, yellow maize, avocado powder, and Jaggery). Thus, this research found that the optimum composite food mixture as the ratio of 40 g of oat, 25.81 g of chickpea, 13.78 g of yellow maize, and 10.41 g of avocado pulp powder with 10 g sugarcane Jaggery is considered a better optimal food composition among the studied components with the response optimizer composite desirability of 0.9085. Also, studied results proved that the appropriate raw materials were processed and formulated well-being complementary food with improved nutritional value suitable for infant feeding and all age hold communities.

Data Availability

All the necessary data have been included in the manuscript.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- D. B. Seboka, "Review on quality characteristics of complementary food and look for policy gap in case of Ethiopia," *American Journal of Health Research*, vol. 7, no. 4, pp. 51–58, 2019.
- [2] M. A. Abeshu, A. Lelisa, and B. Geleta, "Complementary feeding: review of recommendations, feeding practices, and adequacy of homemade complementary food preparations in developing countries–lessons from Ethiopia," *Frontiers in Nutrition*, vol. 3, no. 41, 2016.
- [3] W. Laskowski, H. Górska-Warsewicz, K. Rejman, M. Czeczotko, and J. Zwolińska, "How important are cereals and cereal products in the average polish diet?," *Nutrients*, vol. 11, no. 3, p. 679, 2019.
- [4] D. A. Pawar, M. S. Jadhav, and C. A. Nimbalkar, "Techniques and advances in jaggery processing: a review," *Research Journal of Chemical and Environmental Sciences*, vol. 5, no. 2, pp. 14–20, 2017.

- [5] B. B. Desalegn, K. Abegaz, and E. Kinfe, "Effect of blending ratio and processing technique on physicochemical composition, functional properties and sensory acceptability of quality protein maize (QPM) based complementary food," *International Journal of Food Science and Nutrition Engineering*, vol. 5, no. 3, pp. 121–129, 2015.
- [6] A. G. Lamdande, S. T. Khabeer, R. Kulathooran, and I. Dasappa, "Effect of replacement of sugar with jaggery on pasting properties of wheat flour, physico-sensory and storage characteristics of muffins," *Journal of Food Science and Technology*, vol. 55, no. 8, pp. 3144–3153, 2018.
- [7] A. H. Mahmoud and A. M. El Anany, "Nutritional and sensory evaluation of a complementary food formulated from rice, faba beans, sweet potato flour, and peanut oil," *Food and Nutrition Bulletin*, vol. 35, no. 4, pp. 403–413, 2014.
- [8] Codex Alimentarius Commission, Guidelines on formulated complementary foods for older infants and young children CAC/GL 8-1991, World Health Organization & Food and Agriculture Organization of the United Nations, Rome, Italy, 2013.
- [9] E. A. Decker, D. J. Rose, and D. Stewart, "Processing of oats and the impact of processing operations on nutrition and health benefits," *British Journal of Nutrition*, vol. 112, Supplement 2, pp. S58–S64, 2014.
- [10] T. I. Ademulegun and A. T. Koleosho, "Effects of processing method on the nutrients composition of maize/soya complementary food," *IOSR Journal of Pharmacy and Biological Sciences*, vol. 4, no. 1, pp. 39–43, 2012.
- [11] L. N. Malunga, S. D. Bar-El, E. Zinal, Z. Berkovich, S. Abbo, and R. Reifen, "The potential use of chickpeas in development of infant follow-on formula," *Nutrition Journal*, vol. 13, no. 1, pp. 1–6, 2014.
- [12] N. P. Minh, N. T. Le Trinh, N. T. M. Linh, and C. N. C. Tu, "Several parameters influencing to the production of avocado (Persea americand) powder," *Journal of Pharmaceutical Sciences and Research*, vol. 11, no. 2, pp. 289–294, 2019.
- [13] K. Salelign and R. Duraisamy, "Sugar and ethanol production potential of sweet potato (*Ipomoea batatas*) as an alternative energy feedstock: processing and physicochemical characterizations," *Heliyon*, vol. 7, no. 11, article e08402, 2021.
- [14] K. Akshay, R. Akshay, and D. Rahul, "Developing a sugarcane feeding system for jaggery making plants for rural," *International Journal of Science Technology & Management*, vol. 4, no. 1, pp. 225–228, 2015.
- [15] AOAC (Association of Official Analytical Chemists), Official methods of analysis of the association of analytical chemists international, AOAC, Maryland, 18th edition, 2005.
- [16] T. Z. Mamo, A. G. Mezgebe, and A. Haile, "Developing orange-fleshed sweet potato (*Ipomoea batatas*) juice: analysis of physico-chemical, nutritional," *International Journal of Food Science and Nutrition Engineering*, vol. 4, no. 5, pp. 128–137, 2014.
- [17] J. Orwa, Q. Ansam, Z. Oday, S. Hazem, and Z. Motasem, "Determination of total polyphenolic antioxidants contents invest-bank olive oil," *Journal of Natural Science Research*, vol. 4, pp. 15–23, 2014.
- [18] L. Satpathy, N. Pradhan, D. Dash, P. P. Baral, and S. P. Parida, "Quantitative determination of vitamin C concentration of common edible food sources by redox titration using iodine solution," *Letters in Applied Bioscience NanoBioScience*, vol. 10, pp. 2361–2369, 2021.

- [20] W. Eyasu, F. Tileye, and T. Kassahun, "Proximate, mineral and antinutrient contents of cocoyam (*Xanthosoma sagittifolium* (L.) Schott) from Ethiopia," *International Journal of Food Science*, vol. 2019, Article ID 8965476, 7 pages, 2019.
- [21] A. Tesfay, G. Feyera, W. Aselefech, and E. Tarekegn, "Chemical formulation and characterization of complementary foods from blend of orange-fleshed sweet potato, brown teff, and dark red kidney beans," *International Journal of Food Science*, vol. 2020, Article ID 4803839, 13 pages, 2020.
- [22] O. Oluwaniyi and J. Oladipo, "Comparative studies on the phytochemicals, nutrients and antinutrients content of cassava varieties," *Journal of the Turkish Chemical Society, Section A: Chemistry*, vol. 4, no. 3, pp. 661–674, 2017.
- [23] S. F. Forsido, H. T. Duguma, T. B. Lema, B. Sturm, and O. Hensel, "Nutritional and sensory quality of composite extruded complementary food," *Food Science & Nutrition*, vol. 7, no. 2, pp. 882–889, 2019.