Review Article Surface Modifications and Their Effects on Titanium Dental Implants

A. Jemat,¹ M. J. Ghazali,¹ M. Razali,² and Y. Otsuka³

 ¹Department of Mechanical & Materials Engineering, Faculty of Engineering and Built Environment, UKM, 43600 Bangi, Selangor Darul Ehsan, Malaysia
 ²Department of Peridontology, Faculty of Dentistry, National University of Malaysia, Jalan Raja Muda Abdul Aziz, 50300 Kuala Lumpur, Malaysia
 ³Department of System Safety, Nagaoka University of Technology, 1603-1 Kamitomioka-Cho, Nagaoka-shi, Niigata 940-2188, Japan

Correspondence should be addressed to M. J. Ghazali; mariyam@eng.ukm.my

Received 2 April 2015; Revised 15 June 2015; Accepted 16 June 2015

Academic Editor: Seunghan Oh

Copyright © 2015 A. Jemat et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

This review covers several basic methodologies of surface treatment and their effects on titanium (Ti) implants. The importance of each treatment and its effects will be discussed in detail in order to compare their effectiveness in promoting osseointegration. Published literature for the last 18 years was selected with the use of keywords like titanium dental implant, surface roughness, coating, and osseointegration. Significant surface roughness played an important role in providing effective surface for bone implant contact, cell proliferation, and removal torque, despite having good mechanical properties. Overall, published studies indicated that an acid etched surface-modified and a coating application on commercial pure titanium implant was most preferable in producing the good surface roughness. Thus, a combination of a good surface roughness and mechanical properties of titanium could lead to successful dental implants.

1. Introduction

Surface treatments are normally carried out to modify yet maintain desirable properties of the substrate materials especially in the dental implant industry. The surface area can be increased remarkably by using proper modification techniques, either by addition or subtraction procedures [1, 2]. A surface treatment can also be classified into mechanical, chemical, and physical methods. In dental implant, the surface treatment is used to modify the surface topography and surface energy, resulting in an improved wettability [3-5], increased cell proliferation and growth [3], and accelerated osseointegration process [6]. The quality of dental implant depends on the properties of the surface. In order to have good interaction of the tissue and osseointegration, materials' biocompatibility and roughness of the surface played an important role. Goyal and coworkers [7] observed that the increased roughness can simultaneously increase the surface area of the implant, improve cell migration and attachment

to implant, and enhance osseointegration process. Past literature has revealed most of the surface treatments able to brings a good effect to the dental implants [3-6]. Coating is proved to increase the surface area of the implants substantially [8]. The surface treated with plasma sprayed titanium exhibits the highest value of the surface roughness $(3.43 \pm 0.63 \,\mu\text{m})$ compared to machined surface $(0.15 \pm 0.04 \,\mu\text{m})$ [9]. The healing period was enhanced with hydroxyapatite (HA) coating compared to untreated one [10]. The behavior of modified surface on cells culture studies has revealed that an acid etched zirconia implant surface shows a significant improvement in cell proliferation, except for bone attachment and adhesion on the first day of culture [11-13]. In the study by Parsikia et al. [14], the commercially pure titanium surface was blasted followed by two-step chemical treatment (acid-alkali) resulting in optimized surface topography. The cell bioactivity was improved and expected to have good osseointegration at early stage. Furthermore, a rougher titanium surface promotes shorter healing process [15] than the smoother surfaces. Thus, the surface treatment is used not only to maintain the existing properties of the implants but also to enhance several behaviours as required by dental applications particularly in improving the healing process.

2. Background

2.1. Titanium Implant. Titanium is the material of choice for dental implant as its properties met the most important requirements such as excellent biocompatibility [16], corrosion resistance, high strength, and relatively low modulus of elasticity [17], good formability, and machinability. Additionally, surface modifications are being utilised on implant surfaces, mainly to improve wettability, cell-implant adhesion and attachment, cell proliferation, and osseointegration, and thus faster healing and shorter treatment duration. As a result, many research works have been carried out to improve surface modifications on existing implants to achieve the desired biological responses. The surface topography has also been manipulated such as acid etching and blasting [18] onto the surface to get a better topographies which consequently bring better roughness. In the case of the mechanisms, the roughness of the titanium implants was considered to be one of the significant parameters that affect the rate and the quality of osseointegration [15, 18, 19].

2.2. Biocompatibility of Titanium and Its Alloys. Materials compatibility is the most important issue to be considered for a successful dental implantation. Titanium and its alloys are well known as materials that are well tolerated by living tissues and capable of promoting osseointegration [20]. Ideally, the modification of the implant surface was proposed to enhance osseointegration between materials and bone tissue. The surfaces of materials after treatment should be able to interact with the surrounding tissue to induce direct contact of bone to implant. Kokubo treatment, also known as simulated body fluid (SBF), is a chemical method for inducing or determining a level of biocompatibilities property of dental materials that was established in 1991 [21]. SBF can be described as a solution with ion concentration similar to human blood plasma (see Table 1), kept under mild conditions of pH and identical physiological temperature [21]. The history of SBF usage for apatite formation is shown in Figure 1 [21-25]. In early 1980, Ogino and coworkers [22] have found silicon dioxide (SiO₂) layer and calcium phosphate (CaP) formed on a Bioglass which allows bonding to living bone. In 1990, Kokubo et al. [24] have stated that the formation of apatite is an essential for osseointegration between implant surface and living bone. The full preparation of SBF has been reported in 1995 by Cho et al. [25].

In vivo and in vitro bioactivity of a material can be predicted from the apatite formation on its surface in SBF [26]. Surface conditions, such as surface roughness, surface charge, surface energy, and chemical composition, have important influences on the osseointegration process. Therefore, modifying titanium implant surface seems to be a promising way to achieve stronger and faster osseointegration of the implants and also promoted shorter healing times from implant placement to restoration [27].

 TABLE 1: Ion concentrations (mM) of SBF and human blood plasma
 [21].

| Ion | Simulated body fluid (SBF) | Blood plasma |
|---|----------------------------|--------------|
| Na ⁺ | 142.0 | 142.0 |
| K ⁺ | | |
| K 2 | 5.0 | 5.0 |
| Mg ²⁺ Ca ²⁺ | 1.5 | 1.5 |
| Ca ²⁺ | 2.5 | 2.5 |
| Cl | 148.8 | 103.0 |
| HCO ³⁻ | 4.2 | 27.0 |
| HPO ₄ ^{2–} SO ₄ ^{2–} | 1.0 | 1.0 |
| SO4 ²⁻ | 0.5 | 0.5 |

2.3. Surface Treatment. Recently, many works have been carried out on surface treated commercial titanium implants to enhance the osseointegration function (references). By increasing the surface roughness, an increase in the osseointegration rate and the biomechanical fixation of titanium implants have been observed [27, 28]. The implant modifications can be achieved either by additive or subtractive methods. The additive methods employed the treatment in which other materials are added to the surface, either superficial or integrated, categorized into coating and impregnation, respectively. While impregnation implies that the material/chemical agent is fully integrated into the titanium core, such as calcium phosphate crystals within TiO₂ layer or incorporation of fluoride ions to surface, the coating on the other hand is addition of material/agent of various thicknesses superficially on the surface of core material. The coating techniques can include titanium plasma spraying (TPS), plasma sprayed hydroxyapatite (HA) coating, alumina coating, and biomimetic calcium phosphate (CaP) coating. Meanwhile, the subtractive techniques are the procedure to either remove the layer of core material or plastically deform the superficial surface and thus roughen the surface of core material. The common subtractive techniques are large-grit sands or ceramic particle blasts, acid etch, and anodization [19]. The removal of surface material by mechanical methods involved shaping/removing, grinding, machining, or grit blasting via physical force. A chemical treatment, either by using acids or using alkali solution of titanium alloys in particular, is normally performed not just to alter the surface roughness but also to modify the composition and to induce the wettability or the surface energy of the surface [29]. As for physical treatment such as plasma spray or thermal spray, it is often carried out on the outer coating surface to improve the aesthetic of the material and its performance. Additionally, ion implantation, laser treatment and sputtering [10, 30–33], alkali/acid etching [34–36], and ion deposition [37] are also utilised. Thus, in the light of studying the effects of surface treatments, this review only focuses on various methods that have high potentials in improving the performance of titanium implants. The basic principle of each surface modification and its developments are discussed in the following sections:

- (i) Pretreatment significance.
- (ii) Plasma spray coating.

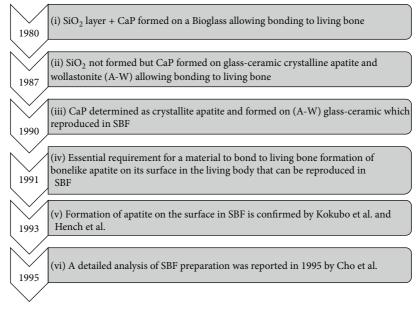


FIGURE 1: History of simulated body fluid (SBF).

- (iii) Grit blasting.
- (iv) Acid etching.
- (v) Dual acid etching (DAE).
- (vi) Sand blast and acid etching (SLA).
- (vii) Other methods.
- (viii) Trends in surface treatment of titanium.
- (ix) Final remarks.

2.3.1. Pretreatment Significance. Prior to the surface modification, pretreatment is required to ensure the substrate surfaces are free from contaminations. Prior to plasma spray procedure, the substrates are normally pretreated by grit blasting [38, 39] to remove the surface impurities and roughened (roughness range $3-5 \mu m$) the surface in order to get better adhesion between substrate and powder [40]. The substrate can also be preheated to reduce residual stress and to avoid crack in the coating [38]. As for an acid etching method, the surface was prepared by polishing with several grits of sand papers [41] to achieve uniform [42] and regular morphology of the surface [43]. Typical surface roughness that is obtained from polishing process is in the range of ~0.1 [44, 45] to $3 \,\mu m$ [43]. Figure 2 shows the typical morphologies of Ti alloy polished using silicon carbide (SiC) grit papers. In short, a pretreatment process is crucial as it provides clean surface, by eliminating undesired defects like scratch and irregularities.

3. Type of Surface Treatment

3.1. Plasma Spray Coating. Plasma spraying technique generally involves thick layer of depositions, such as hydroxyapatite (HA) and titanium (Ti). The coating process includes spraying thermally melted materials on the implant substrates. A combination of HA coating on Ti alloys substrate has received many attentions due to their attractive properties such as good biocompatibility and mechanical properties [32]. The plasma spray substantially increased the surface area of the implants by increasing their surface roughness [46]. The potential of spray plasma spray coatings to enhance the mechanical behaviour has been addressed by many studies [9, 17, 18, 31, 37, 38]. Several techniques were proposed to adhere HA to titanium implants [9, 10, 17], but only the plasma spraying coating technique has been successfully used on commercial implants [19]. A metastable calcium phosphate solution provides excellent bioactivity of the HA/YSZ/Ti-6Al-4V composite coatings, which have the ability to induce bone-like apatite nucleation and growth on implant surface [38]. Fouda et al. [10] reported that HA coated titanium implant could enhance the healing period compared to the uncoated implants. Xie et al. [33] also discovered that HA coatings promote better cell proliferation. However, in some cases, a reverse effect of HA coatings [47, 48] was also noted. According to Liu et al. [49], the bonding strength of HA on titanium alloys decreased long hours of immersion time in the simulated body fluids (SBF). Yang et al. [48] also reported that after an immersion in the SBF, the hydroxyapatite (HA) coatings became weak due to the intermellar or cohesive bonding degradation in the coating. However, Knabe et al. [9] found that a plasma sprayed titanium surface exhibits the highest surface roughness compared to a deep profile surface structure (the surface was acid etched and grit blasted; see Figure 3) and in an in vitro test, the HA coating has less bone contact compared to other surface modifications. Some reports showed that the mechanical properties of HA can be significantly improved by the addition of yttria-stabilized zirconia [40, 50]. Previous study [51] reported that the HA coatings reinforced with zirconia possessed better performance in bond strength and dissolution behaviour of the titanium implants. Over the same period (4 weeks after the

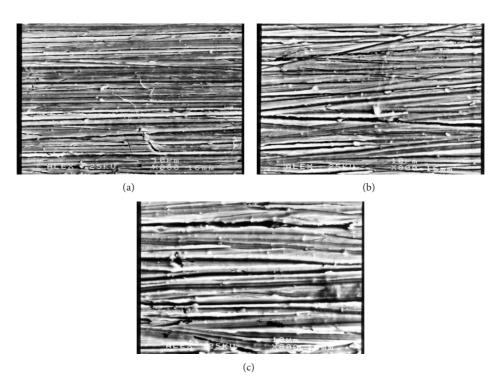


FIGURE 2: Typical morphologies of Ti alloy polished using SiC paper (a) 1200 grit, (b) 600 grit, and (c) 180 grit [42].

SBF immersion), the HA/YSZ/Ti-6Al-4V composite coating showed a reduced tensile strength by ~27.7% compared to the pure HA coatings with ~78.8% [38]. It has been reported that more new bones are formed and grow more rapidly into pores of the surface of alkaline-modified plasma sprayed implants, and this may be beneficial to reduce clinical healing times and thus to improve implant success rates [52].

3.2. Grit Blasting. Another route for roughening the surface is grit blasting, through pressurised particle projection either using ceramic materials or silica onto the implant surface. Materials such as sand, hydroxyapatite, alumina, or TiO₂ particles are usually employed for the purposes [35, 36]. Grit blasting is always followed by an acid etching to remove the residual blasting particles. Hence, the grit blasting is also considered as one of the means to embed surface contaminants on the substrates [51]. Surface microhardness of zirconia particles on titanium surface via blasting was found to be far greater than a controlled polished titanium surface [19]. Al-Radha and coworkers [53] evaluated the effect of bacterial adhesion on several titanium implants with different treatments. The results showed that ZrO₂-blasted titanium exhibited greater bacterial adhesion compared to other surface treatments. In a similar case, Aparicio et al. [34] applied alumina blasting with particle sizes ranging $425-600 \,\mu\text{m}$ to gain high value of surface roughness between $4.15 \pm 0.26 \,\mu\text{m}$. In in vivo studies by Bacchelli et al. [54], they discovered that deposited titanium treated with commercially pure Ti shows the highest surface roughness of 8.55 \pm 0.78 μ m, followed by ZrO₂ sandblasting with improved osteogenesis. This indicates that the blasting method also has an effective role in inducing optimum roughness of dental implants surface [3].

However, this technique is only promising in a good surface but not in terms of osseointegration itself. Besides, bacteria will tend to accumulate more on the rough surface substrate compared to smooth substrate. Thus, further study on how this technique affects the important properties like bone implant contact, removal torque values, tissues response, and bacterial adhesion, and biocompatibility must be carried out.

3.3. Acid Etching. In acid etching, the use of acids on metal surfaces is not only to clean the surface but also to modify the roughness. A strong acid like hydrofluoric (HF), nitric (HNO₃), and sulphuric (H_2SO_4) or a combination of these acids is commonly used in this technique. Acid etched surfaces had increased cell adhesion and bone formation, thus enhancing the osseointegration [3, 49-51, 53, 54, 59-62]. Due to its dissolution ability [63, 64], HF has been used for etching restorative ceramic materials in order to increase the bonding surface for luting agents. The significance of this technique also renders the substrate with homogeneous roughening regardless of the sizes and shapes [63]. The roughness of titanium is one of the factors that helps in determining the stability of bone formation and resorption at the interface of bone implants [65]. Alla et al. [66] reported that a nanotopography that allows bone ingrowth via acid etching on an implant may improve the roughness. Previous study has reported that the rate of etching depends on the type and concentration of the acid used [35]. However, the suitability of these acids in etching was not determined as they required further tests particularly on the bone implant contact and torque removal. Titanium samples etched by H₂SO₄ with different concentrations demonstrated an increase in surface roughness [57]. Concentrated H₂SO₄ has been proven as an

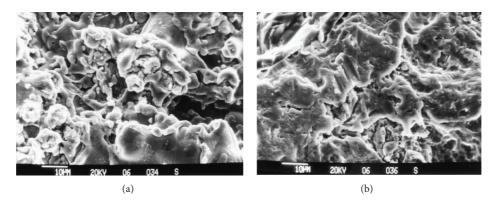


FIGURE 3: Surface morphology by (a) plasma sprayed titanium (b) deep profile structure [9].

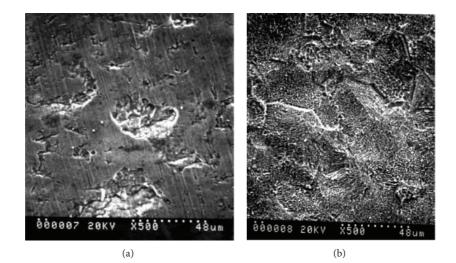


FIGURE 4: Titanium implant with (a) a machined surface and (b) treated dual acid 48% HF + HCl/H₂SO₄ [51].

effective solution to roughen the surfaces particularly for biological applications [66].

3.3.1. Dual Acid Etching (DAE). Similar to acid etching, the DAE is also able to treat the surface via chemical or acid whether in sequence [45] or with the combination of both [67, 68]. Rapid osseointegration can be achieved by dual etching through micro rough surface [55]. A comparative study between a machined surface and those using HF and HCl/H₂SO₄ (DAE) has shown the acid treated surface has greater resistance to reverse torque removal and better osseointegration [55]. In order to examine the surface roughened by the DAE, Yang et al. [48] inserted fifteen implants into rabbit's tibias. It was remarkable to note that roughened surfaces implants showed greater value of a removal torque at 2, 4, and 8 weeks than the machined surface. At the same time, a histomorphometric analysis demonstrated that the bone-to-implant contact significantly increased along with the peri implant bone formation. Thus, the DAE can provide a surface with a certain microroughness, thus contributing to a rapid osseointegration [35]. However, the acid etching treatment is strongly dependent on the acid selection and the

process. Juodzbałys et al. [69] observed that an acid etched titanium implant exhibited similar surface topography as those gained from a sand-blasted large-grit acid etched (SLA) surface treatment. They found that the sample of titanium shows a good surface roughness with 1–10 μ m micropits after etching with H₂SO₄ and then HCl compared to a poor surface microtexture by HCl and then H₂SO₄ [69]. A comparison study had also been carried out between a machined surface and a dual-etched surface as shown in Figure 4.

It was noted that the acid treated surface gave greater resistance in a reverse torque removal and better osseointegration than the machined surface implants [55]. A surface treatment via acid etching on zirconia implant has been reported to have similar effects on density of the bone implant and relative capacity for an osseointegration [61]. However, side effects like porosities, with sizes ranging from 0.5 to 2 μ m, were also formed due to the use of these acids [58, 70]. This process somehow is also believed to benefit tissue ingrowth and cell surface interactions in the dental implant [70]. The success of osseointegration or implant anchorage was measured using a resistance to reverse torque rotation. As torque rotation force value increased, bone-to-implant contact

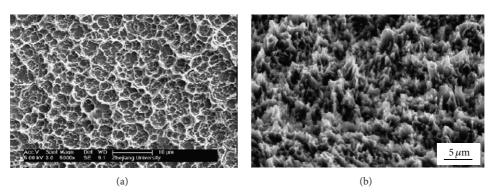


FIGURE 5: The surface morphology of (a) sandblasted and treated Ti6Al4V alloy implants with DAE (HCl and H_2SO_4) [57] and (b) sandblasted and etched Ti implant with warm HCl [58].

(BIC) also increased which lead to greater osseointegration [51].

3.4. Sandblast, Large-Grit, and Acid Etching (SLA). SLA is used to induce surface erosion by applying a strong acid onto the blasted surface [17]. This treatment combines blasting with large-grit sand particles and acid etching sequentially to obtain macro roughness and micro pits [58] to increase the surface roughness as well as osseointegration [71–74]. Cho and Jung [71] discovered that the SLA surface possessed wide cavities (from $5\,\mu m$ to $20\,\mu m$ in diameter) and micro pits (from ~0.5 μ m to 3 μ m in diameter), indicating an increase in the surface roughness and the surface area. Hence, the SLAtreated surface was found to be useful for improving tissue integration and cell proliferation. In vivo studies on six adult dogs carried out by Xue et al. [52] indicated that the surface after sequential grit blasting and alkaline treatment showed high shear strength, improving early bone growth and osseointegration. A recent investigation on a two-step chemical treatment (acid-alkali) noticed that optimised morphology and good bioactivity resulted in good osseointegration during the early stage of the implantation [75]. Similarly, He et al. [76] also discovered that the implants treated with blasting followed by the DAE (HCl and H₂SO₄) promote better osseointegration during the healing phase, indicating a great improvement in the bioactivities. In addition, biological evaluation by Kim et al. [58] discovered that human osteoblasts grow splendidly on the SLA surface which provides greater space for cell attachment and proliferation. Surface morphology for SLA typically became rough and irregular after sandblasting, but then after the acid etching treatment the surface is more uniform and small micro pits (1-2 μ m in diameter) are created as shown in Figures 5(a) and 5(b).

3.5. Other Methods. Ion implantation, laser treatment, sputtering, and other combinations of several mentioned techniques are also briefly discussed in this review. An ion implantation, for example, involved accelerating ions of materials in an electrical field and impact onto the substrate to a depth of approximately 1 μ m [1]. Braceras and his colleagues [37] used this method to investigate the osseointegration properties of the treated implant surface. They found out

that the ion implantation of cobalt onto titanium alloys significantly improved the osseointegration. Deposition via dip coating of nanocomposite (HA-ZrO₂-Al₂O₃) on titanium substrate showed the highest adhesion strength compared to the HA coatings [37]. Another technique observed by Pető et al. [77] involve Nd glass laser, in which the removal torque of implants was 20% larger for laser-treated surface compared to the machined and blasted implants. These results corresponded well with the data reported by Hallgren et al. [75] who demonstrated that the removal torque value was larger for the laser-modified implant (52 Ncm) than the machined surface implant (35 Ncm) after 12 weeks of healing. This result was also in agreement with other studies [9, 78-81]. Using pulsed magnetron sputtering method [72], ZrO₂-Ag and ZrO₂-Cu deposited titanium surface had improved the antibacterial performance relative to pure Ti implant materials [58]. In another study, combination method of laser-treated and acid etched surface was proven to have better osseointegration than the laser-treated surface with BIC value 49.71% [80].

4. Trends in Surface Treatment of Titanium

The greatest interest has been noted in the use of plasma spraying and acid etching techniques. Clearly, the plasma spray method is the most preferable (see Figure 6) due to its advantages in providing porous implant surfaces for greater bone contact [30]. The qualities of the coating surface are strongly dependent on the types of the coating materials. Other than that, study on plasma spray showed good growth cells on the implant surface [9] and a good bone contact which accelerated the bone formation [30]. Relatively, coated implants like ZrO₂ possessed high surface roughness with an approximation of 5.7 \pm 0.2 μ m. This value could be increased up to $8.68 \pm 0.37 \,\mu\text{m}$ [47] when acid etching is applied prior to coating. Even the dual acid etching played an important role in producing good surfaces, with roughness ranging from 0.44 to $3.51 \,\mu\text{m}$ [34, 35, 82]. In general, the DAE is better than a single acid etching due to its high composition, amount, and concentration. In the case of dental implants, the effect of acid etching is based on the concentration and the type of the acid as well as the temperature and time, in which

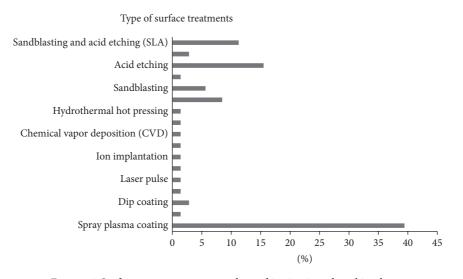


FIGURE 6: Surface treatments commonly used in titanium dental implants.

the surface roughness normally increased with an increasing concentration of acid [67]. Furthermore, the blasting and the SLA techniques were also commonly used to improve the surface roughness and have high potential to improve the implant bone healing. Every single technique has its own advantages and limitations. Thus, to ensure high quality of the coated materials, the importance of a pretreatment on the surface prior to depositing work must also be considered.

In this paper, important methodologies that have been extensively utilised in the surface treatment work of titanium dental implants are summarised in Table 2. In spite of the high number on plasma spray method of studies to date, the results in the literature demonstrate difficulties in deciding the optimum value of surface roughness for better osseointegration yet decrease bacterial adhesion.

5. Final Remarks

All in all, the coating techniques contribute to important positive effects of dental implant application. Most authors [6, 9, 30, 31, 33, 56, 79-81, 83-86] agreed that a good coating technique may give high impact on the mechanical properties of the dental implants. However, this technique has several limitations including poor long-term adherence of the coating to the substrate material [76], nonuniformity in thickness of the deposited layer [84], and variations in crystallinity [87] and composition of the coating. On the practical side, a better understanding of the suitable parameters during plasma spray is important in order to control these limitations. In contrast, most studies could not determine any major advantages or disadvantages with blasted surface implants. Blasting is one popular technique for surface treatments which can easily roughen the implant surface but is inadequate to give credit to the important properties like bone implant contact, removal torque values, tissues response, and biocompatibilities. Ion implantation technique on the other hand is useful to harden the surface of titanium but not applicable for dental implant [64]. It is most useful in orthopaedic devices which

are subject to articulating or in wear situations. Another preferable surface treatment technique is the DAE that has high composition, amount, and concentration [63]. To date, ceramic coatings (calcium phosphate, HA, and TiO₂) still remain the most popular bioceramic materials in the surface treatments area. Nevertheless, HA is recognised as the best candidate in bioceramics compared to TiO₂ [76]. Meanwhile, zirconia also has good potential as dental implants whereby it promotes higher microhardness [33] and better mechanical properties when coated onto Ti alloy. Zirconia stabilized with yttria (YSZ) particles as a secondary phase in coatings is also believed to be dispersion-strengthened due to the homogeneous distribution of YSZ particles in the matrix [84], resulting in good bonding within the composite, and hence improves the mechanical properties.

Currently, surface roughening (e.g., grit blasting, acid etching, and SLA) and coating (e.g., with CaP and HA) are commonly used techniques in practice. Both methods have their advantages and drawbacks as we have discussed in this paper. It has been reported that the improvement of bone implant interface and greater resistance of failure were influenced by acid etched surface [45]. In addition, sandblasted with large-grit (25-50 mm) and acid etched surface were found to have a 50-60% mean value of bone implant contact compared to titanium plasma sprayed surface which had only a 30–40% mean value of bone implant contact after 6 weeks [46]. BIC value is very important in long-term success of dental implants. Numerous studies have demonstrated that rough implants surface show better bone apposition and BIC than implants with smooth surfaces [22, 46, 75]. Surface roughness also stimulated the cell migration and proliferation which in turn leads to better BIC [50]. Different modification methods have been studied, namely, sandblasted, large-grit, acid etched (SLA) and coated surfaces that were chemically different but had the same physical properties that were conducted to assess BIC as a measure of osseointegration.

It is clearly noted that by altering or modifying the surface texture, namely, the roughness of titanium implants,

| Source(s) | Ti type | Surface treatment | Findings | Average roughness Ra (µm) |
|-------------------------------|---|---|---|------------------------------|
| Knabe et al. | CP-Ti ASTM-F67 | Plasma spray Ti coating, acid etching, and sandblasting | All implants except HA coating surface | Ti coating 3.43 ± 0.63 |
| [9] | | HA coating | showed good growth cells. | HA coating 2.07 ± 0.36 |
| Depprich et al. [11] | ZrO_2 | Acid etching | Acid etched surface shows similar properties | 0.598 |
| | Ti | Acid etching | of osseointegration with titanium implant. | 1.77 |
| Hung et al. [17] | CP-Ti (Ti-6Al-4V ELI, ASTM-F136) | Plasma sprayed hydroxyapatite (HA) | Treated implants indicate high biocompatibility for bone regeneration of titanium implants. | Sa 9.36 |
| | Ti | (1) Blasting HA | | 1.2–1.8 |
| Eom et al. [18] | | (2) Blasting and dual acid etching (SLA) | Hybrid type coating shows higher bone implant contact and removal torque value $(259.9 \pm 6.2 \text{ Ncm})$ than other surfaces. | 2.5-3.0 |
| | | (3) hybrid-type coating with HA and blasting | | 3.0-3.5 |
| Darimont et al. [30] | CP-Ti | HA coating Titanium plasma sprayed | HA coating exhibited higher value of bone contact and accelerated the formation of bone. | NR |
| Simmons et al. [32] | CP-Ti | Sintered porous surface Ti spray plasma | The adhesion properties of the porous surface implants are more stiffer and stronger than plasma sprayed implants | NR |
| Xie et al. [33] | CP-Ti | Plasma sprayed dicalcium silicate/ $\rm ZrO_2$ | Higher ZrO_2 content coating layer exhibits smaller dissolution and lesser degree of degradation. | NR |
| Aparicio et al. [34] | CP-Ti ASTM B348 | (1) Acid etching | Blasted and alkaline etched plus thermal | 1.69 ± 0.1 |
| | | (2) Grit blasting | formed rough and bioactive surface lead to accelerate bone tissue regeneration and | 4.74 ± 0.2 |
| | | (3) Grit blasted and alkaline etched+ thermos chemical treatment | increased mechanical retention in the bone. | 4.23 ± 0.2 |
| Ban et al. [35] | CP-Ti | Acid etching with variable parameter (temperature and time) | Surface roughness increased as temperature and time increased. Weight loss increased linearly with time and temperature. | 0.44-3.51 |
| Velasco-Ortega et al. [36] | CP-Ti | Sandblasting with alumina and nitric acid etching (SLA) | After surface treatment, cpTi implant achieved high biocompatibility with no cytotoxic. | NR |
| Yang et al. [48] | Ti | YSZ plasma spray Acid etching | After acid etching, the Ti surface is roughened and may enhance the osseointegration. | 8.68 ± 0.37 |
| Al-Radha et al. [53] | Ti | (1) Blasting with ZrO_2 | Blasted $\rm ZrO_2$ surface showed a very good | 0.158 ± 0.003 |
| | | (2) Blasting with ZrO_2 and acid etching (SLA) | effect on adhesion reducing almost similar to pure ZrO_2 properties. | 0.150 ± 0.005 |
| Chou and Chang [55] | Ti | Grit blasting with alumina and then ${\rm ZrO}_2$ sprayed plasma | ZrO ₂ bond coat promotes adhesion mechanism for Ti substrate. | NR |
| Simon et al. [56] | срТі | Ti plasma spray | Surface roughness by Ti coating may optimize the osseointegration and enhance the clinical function. | 4.4 ± 0.37 |

| TABLE 2: Studies of | n the surface treatment | on Ti dental implants. |
|---------------------|-------------------------|------------------------|
| | | |

NR = No Result.

in particular, desired effects can be obtained like bone implant contact, removal torque values, tissues response, and biocompatibility. Thus, most works still favour surface treatment of dental implants via coating and acid etching over other methods in producing good substrate surfaces for osseoin-tegration, with surface roughness ranging from 0.44 to 8.68 μ m. In short, a good surface with the right roughness and mechanical properties could lead to better osseointegration for successful dental implants.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- W. R. Lacefield, "Materials characteristics of uncoated/ceramiccoated implant materials," *Advances in Dental Research*, vol. 13, pp. 21–26, 1999.
- [2] M. Özcan and C. Hämmerle, "Titanium as a reconstruction and implant material in dentistry: advantages and pitfalls," *Materials*, vol. 5, no. 9, pp. 1528–1545, 2012.
- [3] J. I. Rosales-Leal, M. A. Rodríguez-Valverde, G. Mazzaglia et al., "Effect of roughness, wettability and morphology of engineered titanium surfaces on osteoblast-like cell adhesion," *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, vol. 365, no. 1–3, pp. 222–229, 2010.
- [4] H. Nakae, M. Yoshida, and M. Yokota, "Effects of roughness pitch of surfaces on their wettability," *Journal of Materials Science*, vol. 40, no. 9-10, pp. 2287–2293, 2005.
- [5] L. Ponsonnet, K. Reybier, N. Jaffrezic et al., "Relationship between surface properties (roughness, wettability) of titanium and titanium alloys and cell behaviour," *Materials Science and Engineering C*, vol. 23, no. 4, pp. 551–560, 2003.
- [6] V. Sollazzo, F. Pezzetti, A. Scarano et al., "Zirconium oxide coating improves implant osseointegration in vivo," *Dental Materials*, vol. 24, no. 3, pp. 357–361, 2008.
- [7] N. Goyal and R. K. Priyanka, "Effect of various implant surface treatments on osseointegration—a literature review," *Indian Journal of Dental Sciences*, vol. 4, pp. 154–157, 2012.
- [8] A. B. Novaes Jr., S. L. S. de Souza, R. R. M. de Barros, K. K. Y. Pereira, G. Iezzi, and A. Piattelli, "Influence of implant surfaces on osseointegration," *Brazilian Dental Journal*, vol. 21, no. 6, pp. 471–481, 2010.
- [9] C. Knabe, F. Klar, R. Fitzner, R. J. Radlanski, and U. Gross, "In vitro investigation of titanium and hydroxyapatite dental implant surfaces using a rat bone marrow stromal cell culture system," *Biomaterials*, vol. 23, no. 15, pp. 3235–3245, 2002.
- [10] M. F. A. Fouda, A. Nemat, A. Gawish, and A. R. Baiuomy, "Does the coating of titanium implants by hydroxyapatite affect the elaboration of free radicals. An experimental study," *Australian Journal of Basic and Applied Sciences*, vol. 3, pp. 1122–1129, 2009.
- [11] R. Depprich, M. Ommerborn, H. Zipprich et al., "Behavior of osteoblastic cells cultured on titanium and structured zirconia surfaces," *Head & Face Medicine*, vol. 4, no. 1, article 29, 2008.
- [12] R. M. London, F. A. Roberts, D. A. Baker, M. D. Rohrer, and R. B. O'Neal, "Histologic comparison of a thermal dual-etched implant surface to machined, TPS, and HA surfaces: bone contact in vivo in rabbits," *The International Journal of Oral & Maxillofacial Implants*, vol. 17, no. 3, pp. 369–376, 2002.

- [13] E. A. Bonfante, C. Marin, R. Granato et al., "Histologic and biomechanical evaluation of alumina-blasted/acid-etched and resorbable blasting media surfaces," *Journal of Oral Implantol*ogy, vol. 38, no. 5, pp. 549–556, 2012.
- [14] F. Parsikia, P. Amini, and S. Asgari, "Influence of mechanical and chemical surface treatments on the formation of bonelike structure in cpTi for endosseous dental implants," *Applied Surface Science*, vol. 259, pp. 283–287, 2012.
- [15] J. He, W. Zhou, X. Zhou et al., "The anatase phase of nanotopography titania plays an important role on osteoblast cell morphology and proliferation," *Journal of Materials Science: Materials in Medicine*, vol. 19, no. 11, pp. 3465–3472, 2008.
- [16] S. Vishnu and D. Kusum, "Advances in surface modification of dental implants from micron to nanotopography," *International Journal of Research in Dentistry*, vol. 1, pp. 1–10, 2011.
- [17] K.-Y. Hung, S.-C. Lo, C.-S. Shih, Y.-C. Yang, H.-P. Feng, and Y.-C. Lin, "Titanium surface modified by hydroxyapatite coating for dental implants," *Surface and Coatings Technology*, vol. 231, pp. 337–345, 2013.
- [18] T.-G. Eom, G.-R. Jeon, C.-M. Jeong et al., "Experimental study of bone response to hydroxyapatite coating implants: boneimplant contact and removal torque test," *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, vol. 114, no. 4, pp. 411–418, 2012.
- [19] L. Le Guéhennec, A. Soueidan, P. Layrolle, and Y. Amouriq, "Surface treatments of titanium dental implants for rapid osseointegration," *Dental Materials*, vol. 23, no. 7, pp. 844–854, 2007.
- [20] C. Y. Guo, A. T. H. Tang, and J. P. Matinlinna, "Insights into surface treatment methods of titanium dental implants," *Journal* of Adhesion Science and Technology, vol. 26, no. 1–3, pp. 189–205, 2012.
- [21] T. Kokubo, "Bioactive glass ceramics: properties and applications," *Biomaterials*, vol. 12, no. 2, pp. 155–163, 1991.
- [22] M. Ogino, F. Ohuchi, and L. L. Hench, "Compositional dependence of the formation of calcium phosphate films on bioglass," *Journal of Biomedical Materials Research*, vol. 14, no. 1, pp. 55– 64, 1980.
- [23] T. Kitsugi, T. Nakamura, T. Yamamura, T. Kokubu, T. Shibuya, and M. Takagi, "SEM-EPMA observation of three types of apatite-containing glass-ceramics implanted in bone: the variance of a Ca-P-rich layer," *Journal of Biomedical Materials Research*, vol. 21, no. 10, pp. 1255–1271, 1987.
- [24] T. Kokubo, S. Ito, Z. T. Huang et al., "Ca, P-rich layer formed on high-strength bioactive glass-ceramic A-W," *Journal of Biomedical Materials Research*, vol. 24, no. 3, pp. 331–343, 1990.
- [25] S.-B. Cho, K. Nakanishi, T. Kokubo et al., "Dependence of apatite formation on silica gel on its structure: effect of heat treatment," *Journal of the American Ceramic Society*, vol. 78, no. 7, pp. 1769–1774, 1995.
- [26] T. Kokubo and H. Takadama, "How useful is SBF in predicting in vivo bone bioactivity?" *Biomaterials*, vol. 27, no. 15, pp. 2907– 2915, 2006.
- [27] D. L. Cochran, R. K. Schenk, A. Lussi, F. L. Higginbottom, and D. Buser, "Bone response to unloaded and loaded titanium implants with a sandblasted and acid-etched surface: a histometric study in the canine mandible," *Journal of Biomedical Materials Research*, vol. 40, no. 1, pp. 1–11, 1998.
- [28] A. Wennerberg, C. Hallgren, C. Johansson, and S. Danelli, "A histomorphometric evaluation of screw-shaped implants each prepared with two surface roughnesses," *Clinical Oral Implants Research*, vol. 9, no. 1, pp. 11–19, 1998.

- [29] X. Liu, P. K. Chu, and C. Ding, "Surface modification of titanium, titanium alloys, and related materials for biomedical applications," *Materials Science and Engineering R: Reports*, vol. 47, no. 3-4, pp. 49–121, 2004.
- [30] G. L. Darimont, R. Cloots, E. Heinen, L. Seidel, and R. Legrand, "In vivo behaviour of hydroxyapatite coatings on titanium implants: a quantitative study in the rabbit," *Biomaterials*, vol. 23, no. 12, pp. 2569–2575, 2002.
- [31] A. Ochsenbein, F. Chai, S. Winter, M. Traisnel, J. Breme, and H. F. Hildebrand, "Osteoblast responses to different oxide coatings produced by the sol-gel process on titanium substrates," *Acta Biomaterialia*, vol. 4, no. 5, pp. 1506–1517, 2008.
- [32] C. A. Simmons, N. Valiquette, and R. M. Pilliar, "Osseointegration of sintered porous-surfaced and plasma spray-coated implants: an animal model study of early postimplantation healing response and mechanical stability," *Journal of Biomedical Materials Research*, vol. 47, no. 2, pp. 127–138, 1999.
- [33] Y. Xie, X. Liu, X. Zheng, C. Ding, and P. K. Chu, "Improved stability of plasma-sprayed dicalcium silicate/zirconia composite coating," *Thin Solid Films*, vol. 515, no. 3, pp. 1214–1218, 2006.
- [34] C. Aparicio, A. Padrós, and F.-J. Gil, "In vivo evaluation of micro-rough and bioactive titanium dental implants using histometry and pull-out tests," *Journal of the Mechanical Behavior* of Biomedical Materials, vol. 4, no. 8, pp. 1672–1682, 2011.
- [35] S. Ban, Y. Iwaya, H. Kono, and H. Sato, "Surface modification of titanium by etching in concentrated sulfuric acid," *Dental Materials*, vol. 22, no. 12, pp. 1115–1120, 2006.
- [36] E. Velasco-Ortega, A. Jos, A. M. Cameán, J. Pato-Mourelo, and J. J. Segura-Egea, "In vitro evaluation of cytotoxicity and genotoxicity of a commercial titanium alloy for dental implantology," *Mutation Research—Genetic Toxicology and Environmental Mutagenesis*, vol. 702, no. 1, pp. 17–23, 2010.
- [37] I. Braceras, J. I. Alava, J. I. Oñate et al., "Improved osseointegration in ion implantation-treated dental implants," *Surface and Coatings Technology*, vol. 158-159, pp. 28–32, 2002.
- [38] Y. W. Gu, K. A. Khor, D. Pan, and P. Cheang, "Activity of plasma sprayed yttria stabilized zirconia reinforced hydroxyapatite/Ti-6Al-4V composite coatings in simulated body fluid," *Biomaterials*, vol. 25, no. 16, pp. 3177–3185, 2004.
- [39] H. Zhou, F. Li, B. He, J. Wang, and B.-D. Sun, "Air plasma sprayed thermal barrier coatings on titanium alloy substrates," *Surface and Coatings Technology*, vol. 201, no. 16-17, pp. 7360– 7367, 2007.
- [40] J.-G. Qian, H.-T. Li, P.-R. Li, and Y.-C. Chen, "Preparation of hydroxyapatite coatings by acid etching-electro deposition on pure titanium," in *Proceedings of the International Conference on Biomedical Engineering and Biotechnology (iCBEB '12)*, pp. 433– 436, May 2012.
- [41] P. Y. Lim, P. L. She, and H. C. Shih, "Microstructure effect on microtopography of chemically etched $\alpha + \beta$ Ti alloys," *Applied Surface Science*, vol. 253, no. 2, pp. 449–458, 2006.
- [42] D. D. Deligianni, N. Katsala, S. Ladas, D. Sotiropoulou, J. Amedee, and Y. F. Missirlis, "Effect of surface roughness of the titanium alloy Ti-6Al-4V on human bone marrow cell response and on protein adsorption," *Biomaterials*, vol. 22, no. 11, pp. 1241–1251, 2001.
- [43] S. Ferraris, S. Spriano, G. Pan et al., "Surface modification of Ti-6Al-4V alloy for biomineralization and specific biological response: part I, inorganic modification," *Journal of Materials Science: Materials in Medicine*, vol. 22, no. 3, pp. 533–545, 2011.
- [44] R. Family, M. Solati-Hashjin, S. N. Nik, and A. Nemati, "Surface modification for titanium implants by hydroxyapatite

nanocomposite," *Caspian Journal of Internal Medicine*, vol. 3, no. 3, pp. 460–465, 2012.

- [45] P. Vanzillotta, G. A. Soares, I. N. Bastos, R. A. Simão, and N. K. Kuromoto, "Potentialities of some surface characterization techniques for the development of titanium biomedical alloys," *Materials Research*, vol. 7, no. 3, pp. 437–444, 2004.
- [46] J. L. Ong and D. C. N. Chan, "Hydroxyapatite and their use as coatings in dental implants: a review," *Critical Reviews in Biomedical Engineering*, vol. 28, no. 5-6, pp. 667–707, 2000.
- [47] L. Fu, K. Aik Khor, and J. Peng Lim, "The evaluation of powder processing on microstructure and mechanical properties of hydroxyapatite (HA)/yttria stabilized zirconia (YSZ) composite coatings," *Surface and Coatings Technology*, vol. 140, no. 3, pp. 263–268, 2001.
- [48] G.-L. Yang, F.-M. He, X.-F. Yang, X.-X. Wang, and S.-F. Zhao, "Bone responses to titanium implants surface-roughened by sandblasted and double etched treatments in a rabbit model," *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, vol. 106, no. 4, pp. 516–524, 2008.
- [49] X. Liu, R. W. Y. Poon, S. C. H. Kwok, P. K. Chu, and C. Ding, "Plasma surface modification of titanium for hard tissue replacements," *Surface and Coatings Technology*, vol. 186, no. 1-2, pp. 227–233, 2004.
- [50] E. Chang, W. J. Chang, B. C. Wang, and C. Y. Yang, "Plasma spraying of zirconia-reinforced hydroxyapatite composite coatings on titanium: Part I Phase, microstructure and bonding strength," *Journal of Materials Science: Materials in Medicine*, vol. 8, no. 4, pp. 193–200, 1997.
- [51] S.-A. Cho and K.-T. Park, "The removal torque of titanium screw inserted in rabbit tibia treated by dual acid etching," *Biomaterials*, vol. 24, no. 20, pp. 3611–3617, 2003.
- [52] W. Xue, X. Liu, X. Zheng, and C. Ding, "In vivo evaluation of plasma-sprayed titanium coating after alkali modification," *Biomaterials*, vol. 26, no. 16, pp. 3029–3037, 2005.
- [53] A. S. D. Al-Radha, D. Dymock, C. Younes, and D. O'Sullivan, "Surface properties of titanium and zirconia dental implant materials and their effect on bacterial adhesion," *Journal of Dentistry*, vol. 40, no. 2, pp. 146–153, 2012.
- [54] B. Bacchelli, G. Giavaresi, M. Franchi et al., "Influence of a zirconia sandblasting treated surface on peri-implant bone healing: an experimental study in sheep," *Acta Biomaterialia*, vol. 5, no. 6, pp. 2246–2257, 2009.
- [55] B.-Y. Chou and E. Chang, "Interface investigation of plasmasprayed hydroxyapatite coating on titanium alloy with ZrO₂ intermediate layer as bond coat," *Scripta Materialia*, vol. 45, no. 4, pp. 487–493, 2001.
- [56] M. Simon, C. Lagneau, J. Moreno, M. Lissac, F. Dalard, and B. Grosgogeat, "Corrosion resistance and biocompatibility of a new porous surface for titanium implants," *European Journal of Oral Sciences*, vol. 113, no. 6, pp. 537–545, 2005.
- [57] Y. Iwaya, M. Machigashira, K. Kanbara et al., "Surface properties and biocompatibility of acid-etched titanium," *Dental Materials Journal*, vol. 27, no. 3, pp. 415–421, 2008.
- [58] H. Kim, S.-H. Choi, J.-J. Ryu, S.-Y. Koh, J.-H. Park, and I.-S. Lee, "The biocompatibility of SLA-treated titanium implants," *Biomedical Materials*, vol. 3, no. 2, p. 25011, 2008.
- [59] A. B. Novaes Jr., V. Papalexiou, M. F. M. Grisi, S. S. L. S. Souza, M. Taba Jr., and J. K. Kajiwara, "Influence of implant microstructure on the osseointegration of immediate implants placed in periodontally infected sites," *Clinical Oral Implants Research*, vol. 15, no. 1, pp. 34–43, 2004.

- [60] V. Papalexiou, A. B. Novaes, M. F. M. Grisi, S. S. L. S. Souza, M. Taba Jr., and J. K. Kajiwara, "Influence of implant microstructure on the dynamics of bone healing around immediate implants placed into periodontally infected sites. A confocal laser scanning microscopic study," *Clinical Oral Implants Research*, vol. 15, no. 1, pp. 44–53, 2004.
- [61] J. Y. Park and J. E. Davies, "Red blood cell and platelet interactions with titanium implant surfaces," *Clinical Oral Implants Research*, vol. 11, no. 6, pp. 530–539, 2000.
- [62] M. Wong, J. Eulenberger, R. Schenk, and E. Hunziker, "Effect of surface topology on the osseointegration of implant materials in trabecular bone," *Journal of Biomedical Materials Research*, vol. 29, no. 12, pp. 1567–1575, 1995.
- [63] C. Y. Guo, J. P. Matinlinna, and A. T. H. Tang, "Effects of surface charges on dental implants: past, present, and future," *International Journal of Biomaterials*, vol. 2012, Article ID 381535, 5 pages, 2012.
- [64] T. R. Rautray, R. Narayanan, and K.-H. Kim, "Ion implantation of titanium based biomaterials," *Progress in Materials Science*, vol. 56, no. 8, pp. 1137–1177, 2011.
- [65] K. Suzuki, K. Aoki, and K. Ohya, "Effects of surface roughness of titanium implants on bone remodeling activity of femur in rabbits," *Bone*, vol. 21, no. 6, pp. 507–514, 1997.
- [66] R. K. Alla, K. Ginjupalli, N. Upadhya, M. Shammas, R. Krishna Ravi, and R. Sekhar, "Surface roughness of implants: a review," *Trends in Biomaterials and Artificial Organs*, vol. 25, no. 3, pp. 112–118, 2011.
- [67] A. S. Santiago, E. A. dos Santos, M. S. Sader, M. F. Santiago, and G. de Almeida Soares, "Response of osteoblastic cells to titanium submitted to three different surface treatments," *Brazilian Oral Research*, vol. 19, no. 3, pp. 203–208, 2005.
- [68] M. Takeuchi, Y. Abe, Y. Yoshida, Y. Nakayama, M. Okazaki, and Y. Akagawa, "Acid pretreatment of titanium implants," *Biomaterials*, vol. 24, no. 10, pp. 1821–1827, 2003.
- [69] G. Juodzbalys, M. Sapragoniene, and A. Wennerberg, "New acid etched titanium dental implant surface," *Stomatologija—Baltic Dental and Maxillofacial Journal*, vol. 5, pp. 101–105, 2003.
- [70] C. Massaro, P. Rotolo, F. De Riccardis et al., "Comparative investigation of the surface properties of commercial titanium dental implants. Part I: chemical composition," *Journal of Materials Science: Materials in Medicine*, vol. 13, no. 6, pp. 535– 548, 2002.
- [71] S.-A. Cho and S.-K. Jung, "A removal torque of the laser-treated titanium implants in rabbit tibia," *Biomaterials*, vol. 24, no. 26, pp. 4859–4863, 2003.
- [72] E. Conforto, B.-O. Aronsson, A. Salito, C. Crestou, and D. Caillard, "Rough surfaces of titanium and titanium alloys for implants and prostheses," *Materials Science and Engineering: C*, vol. 24, no. 5, pp. 611–618, 2004.
- [73] T. Monetta and F. Bellucci, "The effect of sand-blasting and hydrofluoric acid etching on Ti CP 2 and Ti CP 4 surface topography," *Open Journal of Regenerative Medicine*, vol. 1, no. 3, pp. 41–50, 2012.
- [74] O. Zinger, K. Anselme, A. Denzer et al., "Time-dependent morphology and adhesion of osteoblastic cells on titanium model surfaces featuring scale-resolved topography," *Biomaterials*, vol. 25, no. 14, pp. 2695–2711, 2004.
- [75] C. Hallgren, H. Reimers, D. Chakarov, J. Gold, and A. Wennerberg, "An in vivo study of bone response to implants topographically modified by laser micromachining," *Biomaterials*, vol. 24, no. 5, pp. 701–710, 2003.

- [76] F. M. He, G. L. Yang, Y. N. Li, X. X. Wang, and S. F. Zhao, "Early bone response to sandblasted, dual acid-etched and H₂O₂/HCl treated titanium implants: an experimental study in the rabbit," *International Journal of Oral & Maxillofacial Surgery*, vol. 38, no. 6, pp. 677–681, 2009.
- [77] G. Pető, A. Karacs, Z. Pászti, L. Guczi, T. Divinyi, and A. Joób, "Surface treatment of screw shaped titanium dental implants by high intensity laser pulses," *Applied Surface Science*, vol. 186, no. 1–4, pp. 7–13, 2002.
- [78] H.-L. Huang, Y.-Y. Chang, J.-C. Weng, Y.-C. Chen, C.-H. Lai, and T.-M. Shieh, "Anti-bacterial performance of Zirconia coatings on Titanium implants," *Thin Solid Films*, vol. 528, pp. 151– 156, 2013.
- [79] A. Joób-Fancsaly, T. Divinyi, A. Fazekas, G. Petó, and A. Karacs, "Surface treatment of dental implants with high-energy laser beam," *Fogorvosi Szemle*, vol. 93, no. 6, pp. 169–180, 2000.
- [80] M. Rong, L. Zhou, Z. Gou, A. Zhu, and D. Zhou, "The early osseointegration of the laser-treated and acid-etched dental implants surface: an experimental study in rabbits," *Journal of Materials Science: Materials in Medicine*, vol. 20, no. 8, pp. 1721– 1728, 2009.
- [81] Y. T. Zhao, Z. Zhang, Q. X. Dai, D. Y. Lin, and S. M. Li, "Microstructure and bond strength of HA(+ZrO₂ + Y₂O₃)/ Ti₆Al₄V composite coatings fabricated by RF magnetron sputtering," *Surface and Coatings Technology*, vol. 200, no. 18-19, pp. 5354–5363, 2006.
- [82] M. Gahlert, S. Röhling, M. Wieland, C. M. Sprecher, H. Kniha, and S. Milz, "Osseointegration of zirconia and titanium dental implants: a histological and histomorphometrical study in the maxilla of pigs," *Clinical Oral Implants Research*, vol. 20, no. 11, pp. 1247–1253, 2009.
- [83] C. Aparicio, D. Rodriguez, and F. J. Gil, "Variation of roughness and adhesion strength of deposited apatite layers on titanium dental implants," *Materials Science and Engineering C*, vol. 31, no. 2, pp. 320–324, 2011.
- [84] I. Dion, L. Bordenave, F. Lefebvre et al., "Physico-chemistry and cytotoxicity of ceramics," *Journal of Materials Science: Materials in Medicine*, vol. 5, no. 1, pp. 18–24, 1994.
- [85] H. Li, Z.-X. Li, H. Li, Y.-Z. Wu, and Q. Wei, "Characterization of plasma sprayed hydroxyapatite/ZrO₂ graded coating," *Materials and Design*, vol. 30, no. 9, pp. 3920–3924, 2009.
- [86] E. S. Thian, J. Huang, Z. H. Barber, S. M. Best, and W. Bonfield, "Surface modification of magnetron-sputtered hydroxyapatite thin films via silicon substitution for orthopaedic and dental applications," *Surface and Coatings Technology*, vol. 205, no. 11, pp. 3472–3477, 2011.
- [87] C.-Y. Yang, T.-M. Lee, Y.-Z. Lu et al., "The influence of plasmaspraying parameters on the characteristics of fluorapatite coatings," *Journal of Medical and Biological Engineering*, vol. 30, no. 2, pp. 91–98, 2010.