Supplementary Figures

CIDR: Ultrafast and accurate clustering through imputation for single-cell RNA-seq data

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Figure S1: *CIDR* **flowchart.** This flowchart illustrates the implicit imputation process through which *CIDR* calculates a dissimilarity matrix. The *CIDR* dissimilarity matrix is then used to perform three tasks: dimensionality reduction, data visualization and clustering.



Figure S2: **Supplementary plots for the simulation study.** (a) Distribution of tags; (b) Tornado plot; (c) Imputation weighting function; (d) Proportion of variation explained by each of the principal coordinates; (e) Calinski-Harabasz Index versus Number of Clusters; (f) *CIDR* two-dimensional visualization.



Figure S3: Toy example: dropout candidates imputed to the row mean (IRM) of the expressed entries. (a) Both the between- and within-cluster distances shrink very significantly. (b) IRM shrinks the between-cluster distances a lot more than the within-cluster distances, and therefore dilutes the clustering signal in the original non-dropout data set. (c) The hierarchical clustering result using IRM.



Figure S4: *CIDR* performance on varying simulation parameters. (a) Varying dropout level. A higher dropout level parameter means a higher level of dropouts. There are 3 cell types with 50 cells per cell type; (b) Varying number of cells in each cell type. There are 3 cell types, and the dropout level parameter is 9.6; (c) Varying number of cell types. There are 50 cells per cell type, and the dropout level parameter is 9.2; (d) Varying number of cells per cell type. There are 7 cell types, and the dropout level parameter is 9.2.



Figure S5: Repeated *t*-SNE runs on the human brain scRNA-seq data set with the same parameters. The different colors denote the cell types annotated by the study¹. *t*-SNE is nondeterministic and it outputs dramatically different plots after repeated runs with the same input and the same parameters. The *t*-SNE parameters used here are k = 4 and perplexity = 10.



Figure S6: Altering the parameters of *CIDR*. The data set here is the human brain scRNA-seq data set¹. (a) nPC = 2, nCluster by default; (b) nPC = 6, nCluster by default; (c) nPC = 4 (default), nCluster = 5; (d) nPC = 4 (default), nCluster = 9.



Figure S7: **Performance evaluation on the mouse brain scRNA-seq data set**². In this data set there are 1,800 cells in 7 cell types. The different colors denote the cell types annotated by the study; while the different plotting symbols denote the clusters output by each algorithm. (a) - (e) Clustering output by each of the five compared algorithms; (f) Adjusted Rand Index is used to measure the accuracy of the clustering output by each of the compared algorithms. Note that it is expected that *t-SNE* achieves a higher Adjusted Rand Index than *CIDR*, as the cell type annotation in the study was obtained through applying *t-SNE*.



Figure S8: **Robustness of** *CIDR* with respect to bandwidth adjustment. *CIDR* uses the default *density* bandwidth selection method 'nrd0' with 'adjust = 1' in the kernel density estimation. Here we vary the 'adjust' parameter and re-calculate the Adjusted Rand Indices for both the human brain¹ and human pancreatic islet³ scRNA-seq data sets.



Figure S9: Computational study on the rate of shrinkage. For a variety of P and \hat{P} , and for any fixed x_1 , the expected rate of shrinkage becomes smaller when x_2 becomes larger. This suggests that, on average, *CIDR* shrinks the distance between closer points more than it shrinks the distance between points that are further apart. This differential shrinkage property helps to reveal the underlying clustering structure in the original data.



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