

# Single-dose Intravenous Toxicology Testing of Daebohwalryeok Pharmacopuncture in Sprague-Dawley Rats

Seung-Ho Sun<sup>1</sup>, Sunju Park<sup>2</sup>, Jong-Jin Jeong<sup>1</sup>, Kwang-Ho Lee<sup>3</sup>, Jun-Sang Yu<sup>4</sup>,  
Hyung-Sik Seo<sup>5</sup>, Ki-Rok Kwon<sup>6\*</sup>

<sup>1</sup> Department of Internal Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>2</sup> Department of Preventive Medicine, College of Korean Medicine, Daejeon University, Daejeon, Korea

<sup>3</sup> Department of Acupuncture & Moxibustion Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>4</sup> Department of Sasang Constitutional Medicine, College of Korean Medicine, Sangji University, Wonju, Korea

<sup>5</sup> Department of Ophthalmology, Otolaryngology, and Dermatology, Korean Medicine Hospital, Pusan National University, Yangsan, Korea

<sup>6</sup> Research Center of the Korean Pharmacopuncture Institute, Seoul, Korea

## Key Words

aqua acupuncture, intravenous injection, pharmacopuncture, single-dose toxicity test

## Abstract

**Objectives:** The aims of the study were to test the single-dose intravenous toxicity of Daebohwalryeok pharmacopuncture (DHRP) in Sprague-Dawley (SD) rats and to estimate the crude lethal dose.

**Methods:** The experiments were conducted at Biototech Co., a Good Laboratory Practice (GLP) laboratory, according to the GLP regulation and were approved by the Institutional Animal Care and Use Committee of Biototech Co. (Approval no: 110156). The rats were divided into three groups: DHRP was injected into the rats in the two test groups at doses of 10 mL/kg and 20 mL/kg, respectively, and normal saline solution was injected into the rats in the control group. Single doses of DHRP were injected intravenously into 6 week old SD rats (5 male and 5 female rats per group). General symptoms were observed and weights were measured during the 14 day observation period after the injection. After the

observation period, necropsies were done. Then, histopathological tests were performed. Weight data were analyzed with a one-way analysis of variance (ANOVA) by using statistical analysis system (SAS, version 9.2).

**Results:** No deaths and no statistical significant weight changes were observed for either male or female SD rats in either the control or the test groups during the observation period. In addition, no treatment related general symptoms or necropsy abnormalities were observed. Histopathological results showed no DHRP related effects in the 20 mL/kg DHRP group for either male or female rats.

**Conclusion:** Under the conditions of this study, the results from single-dose intravenous injections of DHRP showed that estimated lethal doses for both male and female rats were above 20 mL/kg.

## 1. Introduction

Pharmacopuncture is a new type of acupuncture treatment combining acupuncture based on the meridian theory and herbal medicine based on Qi and flavor theory [1]. Pharmacopuncture is categorized as

Received: Nov 27, 2014 Reviewed: Dec 21, 2014 Accepted: Jan 14, 2015

© This is an Open-Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

© This paper meets the requirements of KS X ISO 9706, ISO 9706-1994 and ANSI/NISO Z39.48-1992 (Permanence of Paper).

\*Corresponding Author

Kirok Kwon, Korean Pharmacopuncture Institute, 4F, Association of Korean Oriental Medicine B/D 26-27, Gayang-dong, Gangseo-gu, Seoul 157-200, Korea.  
Tel: +82-33-744-9304 Fax: +82-33-744-9305  
E-mail: drkwon5031@daum.net

© 2015 Korean Pharmacopuncture Institute

<http://www.journal.ac>

meridian field pharmacopuncture (MFP), eight principle pharmacopuncture (EPP), animal based pharmacopuncture (ABP) and mountain ginseng pharmacopuncture (MGP). Pharmacopuncture is prepared by using various methods, such as alcohol immersion, distillation, low temperature extraction, pressing, and dilution [1].

Unlike EPP and MGP, which are made by using the distillation method, pharmacopuncture for Blood and Qi recuperation (BQRP) is made by using the low temperature extraction method. The low temperature extraction method is a novel method for producing pharmacopuncture: It separates the medicinal herb extract after decocting the herb compounds. This is followed by decompression and low temperature distillation. Therefore, BQRP can maximize the inherent effect of the medicinal herbs that were processed [1, 2]. A kind of BQRP is Chukyu (spine-healing) pharmacopuncture [3], Samgihwalryeok pharmacopuncture (SGHRP) [4], Biyeon pharmacopuncture, Cheonghyeol pharmacopuncture, Eunbisan pharmacopuncture, and so on [2].

Daebohwalryeok pharmacopuncture (DHRP) is a kind of BQRP that boosts energy, and it is used for general weakness and fatigue [2]. The indications of DHRP are similar to those of MGP and SGHRP [4-6]. SGHRP has been mainly applied in clinics by using intramuscular or subcutaneous methods. DHRP has a composition similar to that of SGHRP. DHRP, with the same dosage methods and development purposes of MGP, was developed for intravenous administration to increase the effect of SGHRP. Even though toxicity is critical for intravenous injections, studies on the intravenous injection of DHRP have not yet proven its safety whereas studies on the intravenous single-dose toxicity of MGP and on the intramuscular single-dose toxicity of SGHRP have reported those pharmacopunctures to be safe under those injection conditions [4, 6]. Thus, intravenous injection toxicity tests of DHRP are necessary prior to its application. The aims of the study were to test the single-dose intravenous toxicity of DHRP in Sprague-Dawley (SD) rats and to estimate crude lethal dose.

## 2. Materials and Methods

For the preparation of DHRP, 400 g of DHRP were extracted with 90% ethyl alcohol (EtOH) for 72 hours. The DHRP extract was obtained after the separation of extracting EtOH, which was followed by a 40°C heat treatment to evaporate the volatile EtOH. Medicinal plant powder was mixed with water for injection (WFI) at an appropriate ratio in a low temperature low pressure extractor (Fine FA, Korea). After the extravasation, the pharmacopuncture was extracted at 0.1 Torr. The extract was balanced at a pH of 7.25—7.35 and a salinity of 0.9%. The extract was passed through a N<sub>2</sub> Gas filter (0.1 μm, Sartorius), was transferred to the filling tank/reservoir, and was fired in the vial for high pressure sterilization.

SD rats (Orientbio Inc., Korea) were used in this study as they are widely used for toxicity tests [3-4, 6]. Basic visual examinations were done for incoming animals, which were followed by weight measurements with an electronic scale (CP3202S, Sartorius, Germany). The general condi-

tions were observed daily during the 7 day stabilization/conditioning period. After the general conditions had been observed, the animals were moved to an animal room/chamber for quarantine. The body weights, general basic conditions, and weight changes were measured on the last day of stabilization to confirm/ensure the health statuses of the SD rats. At the time of intravenous injection, the weight range of the 6 week old male rats was 178.4—195.8 g (n = 15) and that of the 6 week old female rats was 148.5—169.5 g (n = 15). Breeding conditions were as follows: a temperature of 19.0—23.2°C, a relative humidity of 33.0%—59.5%, a ventilation rate of 10—15 times/hour, an illumination time of 12 hours/day (7:00 am—7:00 pm), and an intensity of illumination of 150—300 Lux.

Telkad Certified Irradiated Global 18% Protein Rodent Diet 2918C (Harlan Laboratories, Inc., U.S.A.) was used to feed the animal *Ad-libitum*. This study was approved by the Institutional Animal Care and Use Committee of Biototech Co. (Approval no: 110156). The tests were conducted according to the Good Laboratory Practice (GLP) regulation and the toxicity test guidelines of the Korea Ministry of Food and Drug Safety (MFDS). On the last day of stabilization (the grouping day), all animals whose weights were equal to the mean weight were randomly grouped into 3 groups, with 5 male rats and 5 female rats for each group. The groupings were as follows (Table 1).

DHRP were injected intravenously according to the schedule used in clinical settings. Dosages for the control and the high dose groups were 20 mL/kg each, and that for the low dose group was 10 mL/kg. For all groups, half of each dose was injected at a 2 hours interval. The initial dose for each animal was calculated based on the weight at the time of the injection. A 26G needle (3 mL) was used for all animals, and the injection speed was approximately 2 mL/minutes. The expected clinical dosage for DHRP was approximately 20 mL/human/day, which is approximately 0.33 mL/kg for an adult of 60 kg. In a pilot test (Biototech Study no.: B10928P) no deaths were observed for single-dose intravenous injections of 20 mL/kg and 10 mL/kg in male and female rats. As a result, dosages for this study were set at 20 mL/kg, maximum available dose, for the high dose group, which is approximately 60 times the expected clinical dosage, and 10 mL/kg for low dose group. Normal saline (Choongwae Pharma Corp., Korea) was injected into the rats in the control group.

At the day of injection (day 0), general conditions (types of toxicity, onset time, recovery time) and deaths were observed 30 minutes, 1 hour, and 2 hours after the first injection, and at 30 minutes, 1, 2, 4, and 6 hours after the second

**Table 1** Grouping of animals

| Group                      | Dosage (mL/kg) | Number of animals |        |
|----------------------------|----------------|-------------------|--------|
|                            |                | Male              | Female |
| G1: Control group (Saline) | 20             | 5                 | 5      |
| G2: Low-dose group (DHRP)  | 10             | 5                 | 5      |
| G3: High-dose group (DHRP) | 20             | 5                 | 5      |

DHRP, Daebohwalryeok pharmacopuncture.

injection. General conditions were noted daily for 14 day after injection. Body weights were measured on the day of the injection (before injection) and on the third, seventh, and fourteenth (necropsy day) days after injection. After the observation period, necropsy with CO<sub>2</sub> gas anesthesia, followed by abdominal aorta bloodletting, was conducted for all animals. Organs and tissues, including the brain, heart, liver, spleen, kidney, lung, and spinal nerve, were extracted from sacrificed animals and fixed with neutral buffered formalin solution. After the organs and tissues had been fixed, sections were produced with dehydrated paraffin. Histotomies were dyed with hematoxylin & eosin (H&E) solution. Residual organ, tissue and fixed organ, tissue were preserved with 10% neutral buffered formalin solution. Microscopic examinations were performed for all sections made for histotomy in both the control group and the high dose group.

Data were analyzed with statistical analysis system (SAS, version 9.2, SAS Institute Inc., U.S.A). A homoscedasticity check was done with Bartlett test (significance level of 0.05). When homoscedasticity was satisfied, a one-way analysis of variance (ANOVA) was done, followed by a Dunnett's *t*-test for a post-hoc analysis (significance levels of 0.05 and 0.01 were used for the two sided test).

### 3. Results

No deaths were observed for both male and female rats in the control and the test groups during the observation period (Table 2). In addition, no abnormal conditions were observed for both male and female rats in the control and the test groups during the observation period (Table 3). Moreover, no statistically significant weight changes were observed for both male and female SD rats in the test groups, compared with the SD rats in the control group,

during the observation period (Figs. 1, 2).

As a result of necropsy, no macroscopic abnormalities were observed in the control and the test groups during the observation period (Table 4). No DHRP related effects were observed in the 20 mL/kg DHRP group for both male and female rats. Other findings that were observed in the kidneys and the livers in the control and the high dose groups, which were spontaneous and adventitious ones, were lesions commonly observed in age relevant SD rats and had no toxicological significance (Fig. 3, Table 5).

### 4. Discussion

The composition of DHRP is equal to that of SGHRP, which consists of *Panax ginseng radix*, *Cornu cervi parvum* (CC), *Angelicae gigantis radix* (AGR), *Ophiopogonis radix* (OR), *Liriope platyphylla*, and *Schisandrae fructus* (SF) [2, 4], but the dose rate of each herb medicine is different. The major organized herbal medicines of DHRP are as follows:

*Panax ginseng radix* is the dried root of *Panax ginseng* C. A. Mey. (family Aralicaceae). It has been used to tonify Qi, the spleen, and the lung, to tranquilize, to stimulate the secretion of body fluids to quench thirst, and to prevent shock and prostration. It has been demonstrated to excite and inhibit the central nervous system (CNS), to have cardiotoxic, anti-fatigue, anti-shock, anti-aging, and anti-cancer effects, to strengthen anti-stress and immune functions, to stimulate hematogenesis, and to improve learning and memory [7-15].

CC is the hairy, young horn of a male deer or stag, *Cervus nippon* Temminck or *Cervus elaphus* L. (family Cervidae). It has been used to reinforce the vital function of the kidney's yang, to strengthen bones and muscles, and to treat the loss of spontaneous seminal emission and strength,

**Table 2** Summary of mortalities

| Sex    | Group / Dose (mL/kg) | No. of animals | Days after dosing |   |   |   |   |   |   |   |   |   |    |    |    |    | Mortality (dead/total) |    |    |       |
|--------|----------------------|----------------|-------------------|---|---|---|---|---|---|---|---|---|----|----|----|----|------------------------|----|----|-------|
|        |                      |                | 0                 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 |                        | 14 |    |       |
| Male   | G1/ Saline (20)      | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |
|        | G2/ DHRP (10)        | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |
|        | G3/ DHRP (20)        | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |
| Female | G1/ Saline (20)      | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |
|        | G2/ DHRP (10)        | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |
|        | G3/ DHRP (20)        | 5              | 0                 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0  | 0  | 0  | 0  | 0                      | 0  | 0% | (0/5) |

**Table 3** Summary of clinical signs

| Sex     | Group / Dose (mL/kg) | No. of animals | Clinical sign | Hours (Day 0) after dosing |   |   |                        |   |   |   |   |   |
|---------|----------------------|----------------|---------------|----------------------------|---|---|------------------------|---|---|---|---|---|
|         |                      |                |               | 1 <sup>st</sup> dosing     |   |   | 2 <sup>nd</sup> dosing |   |   |   |   |   |
|         |                      |                |               | 0.5                        | 1 | 2 | 0.5                    | 1 | 2 | 4 | 6 |   |
| Male    | G1/ Saline (20)      | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |
|         | G2/ DHRP (10)        | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |
|         | G3/ DHRP (20)        | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |
| Fe-male | G1/ Saline (20)      | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |
|         | G2/ DHRP (10)        | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |
|         | G3/ DHRP (20)        | 5              | NOA           | 5                          | 5 | 5 | 5                      | 5 | 5 | 5 | 5 | 5 |

| Sex     | Group / Dose (mg/kg) | No. of animals | Clinical signs | Days after dosing |   |   |   |   |   |   |   |   |   |    |    |    |    |
|---------|----------------------|----------------|----------------|-------------------|---|---|---|---|---|---|---|---|---|----|----|----|----|
|         |                      |                |                | 0                 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 |
| Male    | G1/ Saline (20)      | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |
|         | G2/ DHRP (10)        | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |
|         | G3/ DHRP (20)        | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |
| Fe-male | G1/ Saline (20)      | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |
|         | G2/ DHRP (10)        | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |
|         | G3/ DHRP (20)        | 5              | NOA            | 5                 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5 | 5  | 5  | 5  | 5  |

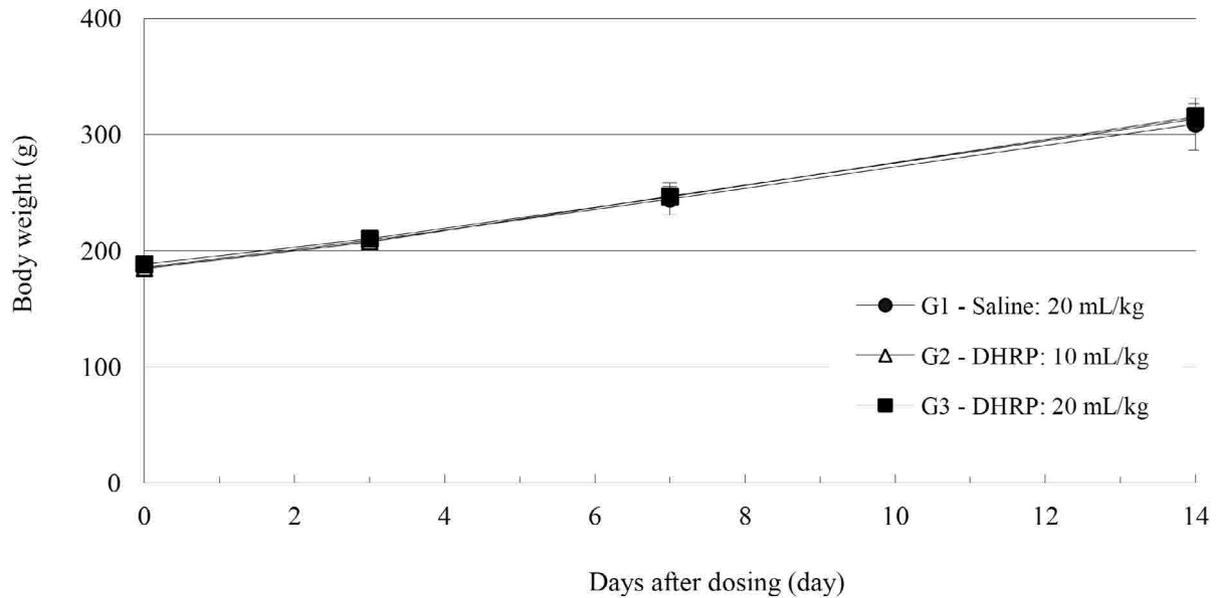
DHRP, Daebohwalryeok pharmacopuncture; NOA, no observable abnormality.

impotence, and leukorrhea due to deficiencies of vital functions caused by chronic diseases [7, 8]. Also, CC has been reported to stimulate the synthesis of proteins and nucleic acids, hematogenesis, and the immune and sexual functions, to strengthen learning and memory ability, and to protect against fatigue and aging [7, 8, 16, 17].

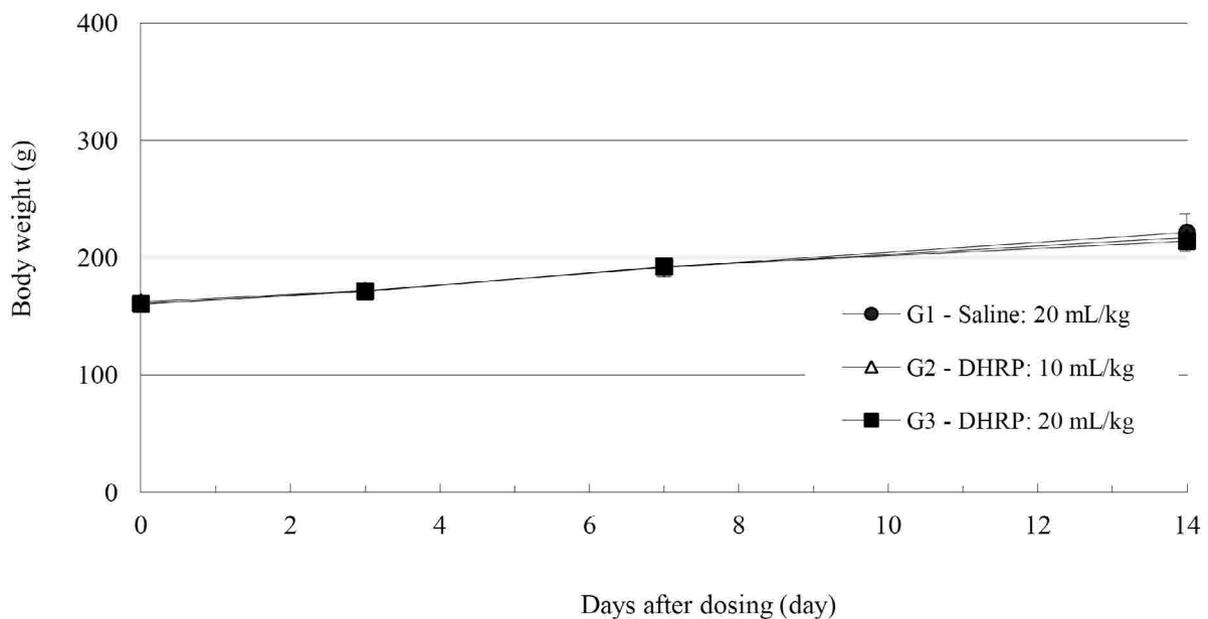
AGR is the root of *Angelica gigas* NAKAI (family Umbelliferae) and has been known as the best medicine in gynecology as it nourishes the blood. It has been used to nourish the blood and to invigorate the blood circulation for the treatment of menstrual disorders and as an emollient and

laxative for chronic constipation of the aged and debilitated. It tonifies an aversion to cold, a fever, and deficiencies induced by consumptive disease [7, 8, 18]. It has been known for its anti-thrombotic, anti-anemic, vasodilation, and hypotensive functions. Furthermore, AGR has been shown to have possible effects on hematogenesis stimulation, immune function improvement, and excitement or inhibition of bronchial and uterine smooth muscles [8, 19-24].

OR is the root of *Ophiopogon japonicas* Ker.-Gawl or *Liriope platphylla* WANG et TANG (family Liliaceae). It is used



**Figure 1** Body weights in male Sprague-Dawley rats. DHRP, Daebohwalryeok pharmacopuncture.



**Figure 2** Body weights in female Sprague-Dawley rats. DHRP, Daebohwalryeok pharmacopuncture.

to replenish the vital essence, to promote the secretion of body fluids, to stop coughing by soothing the lung for the treatment of dipsois, dry throat, dry cough and bloody sputum, and to nourish the heart for the treatment of palpitation and fearfulness [7, 8]. OR has been suggested to have a possible effect when used to treat atopic dermatitis, diabetes mellitus, and inflammation in autoimmune diabetes mellitus. It also has an immunomodulatory effect to protect against lung or liver injury and can be used to improve the lung's capacity, to improve the production and

the secretion of respiratory mucus, to reduce apoptosis, and to improve neuro protection of neuron cells at the hippocampus [25-34].

SF is the dried fruit of *Schizandra chinensis* (Turez) Bailon (family Magnoliaceae) and causes contractions, arrests discharges, boosts Qi, engenderd fluid, tonifies the kidneys, and tranquilizes the heart [7, 8]. It has been used as an astringent for the treatment of dry cough, asthma, night sweating, liver hepatitis, seminal emission and chronic diarrhea; it has also been used as a tonic for neu-

rasthenia [7]. SF has been proposed to have hypotensive, anti-oxidative, anti-inflammation, and hypoglycemic effects, to suppress CNS, to protect liver function, and to inhibit the growth of prostate cancer [8, 35-40].

On the basis of the effect of the above herbs, DHRP can greatly tonify both the Qi and the blood system of the body. DHRP has an effect similar to that of SGHRP because DHRP has the same composition as the SGHRP herbs that tonify Qi, blood, fluid and humor. Thereby, we postulate that not only does DHRP produce healthy circulation that results in enhanced yang Qi, but also it can be applied to treat diverse consumptive or intractable diseases due to Qi deficiency, blood deficiency, and Qi and blood deficiency, such as lassitude, gastric atony, chronic fatigue, stroke and cancer.

As MGP was developed for intravenous injection to en-

hance the effect of tonifying Qi and blood [6], DHRP was developed for intravenous administration to improve the effect of SGHRP [4]. Not only has MGP been widely used intravenously in clinics in Korea, but also the safety of MGP has been proven by using intravenous single-dose toxicity test [6].

On the basis of our single-dose toxicity study, we conclude that intravenous injection of DHRP can be safe because all SD rats showed tolerance to doses over 20 mL/kg. We secured the basic safety evidence that DHRP can be applied intravenously to patients with deficiency. However, further safety studies, such as 4-week recovery tests, 13-week, repeated intravenous dose toxicity tests and so on, and further efficacy studies will be needed in order to provide more conclusive results. In addition, we think DHRP can be injected into subcutaneous tissue or muscle to treat

**Table 4** Summary of necropsy findings

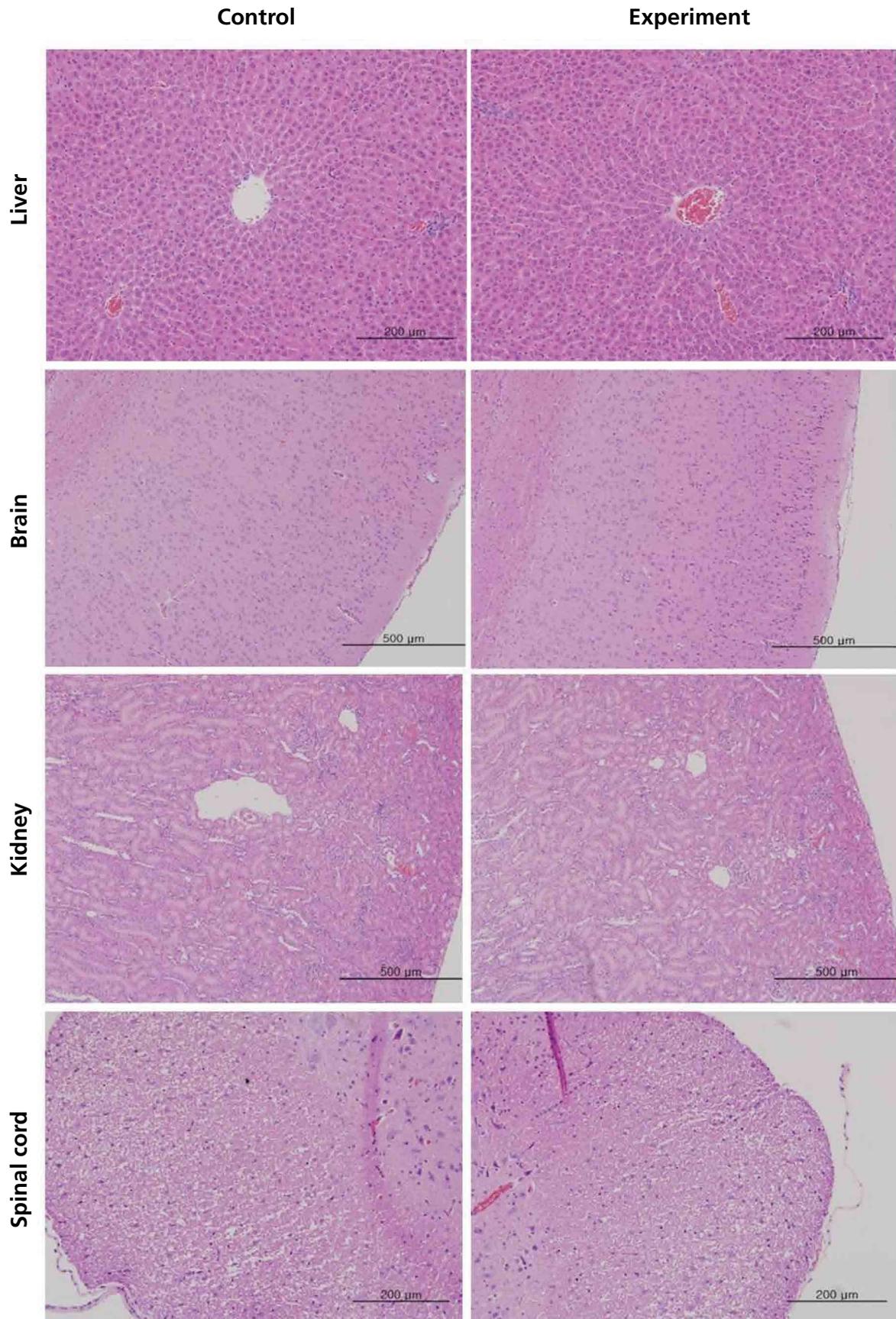
| Sex                   | Male   |      |    | Female |      |    |
|-----------------------|--------|------|----|--------|------|----|
| Group                 | G1     | G2   | G3 | G1     | G2   | G3 |
| Dose (mL/kg)          | Saline | DHRP |    | Saline | DHRP |    |
|                       |        | 20   | 10 | 20     | 20   | 10 |
| No. of animals        | 5      | 5    | 5  | 5      | 5    | 5  |
| Unremarkable findings | 5      | 5    | 5  | 5      | 5    | 5  |
| No. of examined       | 5      | 5    | 5  | 5      | 5    | 5  |

External surfaces and all organs in the body cavities were unremarkable.  
DHRP, Daebohwalryeok pharmacopuncture.

**Table 5** Summary of histopathological findings

| Sex              | Group                               | Male   |      | Female |      |
|------------------|-------------------------------------|--------|------|--------|------|
|                  |                                     | G1     | G3   | G1     | G3   |
| Organ / Findings | Dose (mL/kg)                        | Saline | DHRP | Saline | DHRP |
|                  |                                     |        | 20   | 20     | 20   |
|                  | No. of animals                      | 5      | 5    | 5      | 5    |
| Kidney           | Basophilic tubules                  | ±      | 0    | 0      | 1    |
|                  | Mineralization, outer medulla       | ±      | 0    | 0      | 1    |
|                  | No. of examined                     |        | 5    | 5      | 5    |
| Liver            | Microgranuloma                      | ±      | 0    | 1      | 0    |
|                  | Vacuolation, hepatocyte, periportal | ±      | 0    | 0      | 1    |
|                  | No. of examined                     |        | 5    | 5      | 5    |

There were unremarkable changes in the brain, heart, lung, spleen and spinal nerves of Groups 1 and 3.  
DHRP, Daebohwalryeok pharmacopuncture; Grade- ±, minimal.



**Figure 3** Histopathological observations (hematoxylin & eosin staining  $\times 200$ ).

diseases, just as SGHRP can [4], because DHRP consists of SGHRP and has the same applications in clinics. Thus, safety and efficacy studies for different dose methods will be needed.

## 5. Conclusions

Under the conditions of this study, single-dose intravenous injection of DHRP is safe because lethal doses were estimated to be over 20 mL/kg for both male and female rats.

## Acknowledgment

The authors have no financial interests related to the material of this manuscript.

## Conflict of interest

The authors declare that there are no conflict of interest.

## References

1. Korean Pharmacopuncture Insitute. [Pharmacopunctureology: principles and clinical application]. Seoul: Elsevier Korea LLC; 2012. Chapter 1, Definition and history; p. 3-5. Chapter 3, Types of pharmacopuncture; p. 9-12. Korean.
2. Lee JS. editor. [Blood and Qi recuperated pharmacopuncture]. Proceedings of 2013 Korean Pharmacopuncture Institute Academic Lecture; 2013 May 25; Wonju, Korea: c2013. 46-9 p. Korean.
3. Jeong HH, Cho SH, Lee EY, Lee SD, Ahn SH, Kim SC. Single dose toxicity of chukyu (spine-healing) pharmacopuncture injection in the muscle of rats. *J Pharmacopuncture*. 2014;17(1):35-43.
4. Kim SC, Ahn SH. Single intramuscular-dose toxicity of samgihwalryeok-pharmacopuncture in sprague-dawley rats. *J Pharmacopuncture*. 2014;17(2):46-56.
5. Lee YH, Kim CW, Lee KH. A case report of monitoring PSA level changes in two prostate cancer patients treated with mountain ginseng pharmacopuncture and sweet bee venom along with western anticancer therapy. *J Pharmacopuncture*. 2011;14(4):81-8. Korean.
6. Lee KH, Sun SH, Yu JS, Lim CS, Kwon KR. Intravenous single-dose toxicity of mountain ginseng pharmacopuncture in sprague-dawley rats. *J Pharmacopuncture*. 2014;17(3):50-6.
7. Xie Z, Huang XK. Dictionary of traditional Chinese medicine. Hong Kong: The Commercial Press, Ltd; 1984. p. 205-18.
8. Han JH, Kim KY. [Korean traditional pharmacology]. Seoul: Euseongdang publishing INC; 2004. p. 479-83, 529-32. Korean.
9. Joo IK, Kim HY, Kim JH, Shehzad O, Kim YS, Han YM. [Effects of ginsenosides Rd and Rg1 on proliferation of B cells and antibody]. *Yakhak Hoeji*. 2013;57(1):1-7. Korean.
10. Kim SM. [Effects of ginseng radix on the rat hypothyroidism induced by PTU (6-n-propyl-2-thiouracil)]. *Kor J Herbology*. 2010;25(3):11-8. Korean.
11. Ko SK, Leem KH. [Discussion of ginseng properties through a historical research of Korean ginseng]. *Kor J Herbology*. 2009;24(3):169-72. Korean.
12. Liu Z, Li W, Li X, Zhang M, Chen L, Zheng YN, *et al*. Antidiabetic effects of malonyl ginsenosides from panax ginseng on type 2 diabetic rats induced by high-fat diet and streptozotocin. *J Ethnopharmacol*. 2013;145(1):233-40.
13. Shin YM, Jung HJ, Choi WY, Lim CJ. Antioxidative, anti-inflammatory, and matrix metalloproteinase inhibitory activities of 20(S)-ginsenoside Rg3 in cultured mammalian cell lines. *Mol Biol Rep*. 2013;40(1):269-79.
14. Wang J, Li S, Fan Y, Chen Y, Liu D, Cheng H, *et al*. Anti-fatigue activity of the water-soluble polysaccharides isolated from panax ginseng C. A. Meyer. *J Ethnopharmacol*. 2010;130(2):421-3.
15. Ye R, Li N, Han J, Kong X, Cao R, Rao Z, *et al*. Neuroprotective effects of ginsenoside Rd against oxygen-glucose deprivation in cultured hippocampal neurons. *Neurosci Res*. 2009;64(3):306-10.
16. Kim YK, Choi YH, Song JH, Jang SJ, Kim HJ, Lee CH, *et al*. [Inhibitory effect of deer antler on osteoclastic bone resorption]. *Korean J Orient Physiol Pathol*. 2009;23(6):1299-304. Korean.
17. Lee KB, Park SK. [Effects of cornu cervi parvum pharmacopuncture on the blood picture and antioxidative activity in rats]. *Korean J Acupunct*. 2010;27(2):25-34. Korean.
18. Heo J. [Donguibogam: principles and practice of eastern medicine. part III-2 miscellaneous disorders]. Seoul: Ministry of Health & Welfare. Korea Institute of Oriental Medicine; 2012. p. 150-6. Korean.
19. Kim KS, Song YJ. [Effects of aqua-acupuncture with radix angelicae gigantis and radix astragali on the blood in the liver damage rats with CCL4]. *The acupuncture*. 1996;13(1):1-10. Korean.
20. Hwang HS, Ahn BC, Park DS. [The effects of angelicae gigantis radix aqua - acupuncture by density on immune response induced by radiation in mice]. *The acupuncture*. 1994;11(1):113-29. Korean.
21. Kim SJ, Song BK, Lee EJ, Kim HK, Kim JK. [Effects of radix angelicae gigantis and resina ferulae on the relaxation of smooth muscle and expression of iNOS]. *J Korean Oriental Med*. 2000;20(2):60-7. Korean.
22. Kim YJ, Hwang CW. [An experimental study on brain damage and cardiovascular system effect of angelicae gigantis radix extract]. *J Korean Oriental Med*. 2000;21(4):37-46. Korean.
23. Park YC, Lee JS, Kim MH, Kim DY, Lee SD. [Pharmacological action and toxicity of angelica sinensis]. *Herbal Formula Science*. 2011;19(2):93-108. Korean.
24. Sun Y, Tang J, Gu X, Li D. Water-soluble polysaccharides from angelica sinensis (oliv.) diels: preparation, characterization and bioactivity. *Int J Biol Macromol*. 2005;36(5):283-9.
25. Jang SE, Kim YB. [The effects of radix ophiopogon

- japonicus on the NC/nga atopy model]. J Korean Orient Med Ophthalmol Otolaryngol Dermatol. 2008;21(3):10-9. Korean.
26. Lee ES, Yang SY, Kim MH, Nam GU, Park YC. [Effects of root of *liriope spicata* on LPS-induced lung injury]. Korean J Orient Physiol Pathol. 2011;25(4):641-9. Korean.
27. Nam SH, Choi SL, Goo JS, Kim JE, Lee YK, Hwang IS, *et al.* [LP-M, a novel butanol-extracts isolated from *liriope platyphylla*, could induce the neuronal cell survival and neuritic outgrowth in hippocampus of mice through akt/ERK activation on NGF signal pathway]. Korean J Life Sci. 2011;21(9):1234-43. Korean.
28. Park SD, Lee GH, Lee YS, Kwon YK, Park JH, Choi SM, *et al.* [Comparison of immunomodulatory effects of water-extracted *adenophorae radix*, *liriopis tuber*, *dendrobii herba*, *polygonati odorati rhizoma* and *polygonati rhizome*]. Korean J Orient Physiol Pathol. 2007;21(2):414-24. Korean.
29. Park SH, Kim YS. [Effects of *liriopis tuber* on 4-HNE-induced apoptosis in PC-12 cells]. Kor J Herbology. 2013;28(2):33-8. Korean.
30. Roh SS, Choi HJ, Kim DH, Seo YB. [Studies of anti-inflammation of *liriopis tuber* to autoimmune diabetes in NOD mice]. Korean J Orient Physiol Pathol. 2008;22(4):766-70. Korean.
31. Kim JH, Kim JE, Lee YK, Nam SH, Her YK, Jee SW, *et al.* [The extracts from *liriope platyphylla* significantly stimulated insulin secretion in the HIT-T15 pancreatic  $\beta$ -Cell Line]. J Life Sci. 2010;20(7):1027-33. Korean.
32. Park DI. [Effects of *maekmundong-tang* on the improvement of lung capacity]. Herbal Formula Science. 2013;21(2):165-72. Korean.
33. Sung HK, Min SY, Kim JH. [Effect of *macmundongtang* on production and secretion of respiratory mucus]. J Pediatrics of Korean Medicine. 2013;27(1):69-81. Korean.
34. Rhee IJ, An JY. [Hepatoprotective effects of water extract of *liriopis tuber* on carbon tetrachloride-induced hepatotoxicity in rats]. Korean J Pharmacogn. 2003;34(2):166-71. Korean.
35. Choo BK, Chung KH, Seo YB, Roh SS. [Antioxidant, antiinflammation and hepatoprotective activity of *schizandrae fructus* processed with differentiated steaming number]. Kor J Herbology. 2013;28(2):83-92. Korean.
36. Kwon DY, Kim DS, Yang HJ, Park S. The lignan-rich fractions of *fructus schisandrae* improve insulin sensitivity via the PPAR- $\gamma$  pathways in *in vitro* and *in vivo* studies. J Ethnopharmacol. 2011;135(2):455-62.
37. Lee YM, Moon BC, Ji YU, Seo HS, Kim HK. [Development of RAPD-derived SCAR markers and multiplex-PCR for authentication of the *schisandrae fructus*]. Korean J Med Crop Sci. 2013;21(3):165-73. Korean.
38. Moon JM, Seok GH, Cho SI. [Antiproliferative effect of *schisandrae fructus* extract on PC-3 human prostate cancer cells]. Kor J Herbology. 2012;27(4):17-23. Korean.
39. Park SM, Kim JJ, Jeong KY, Han SK, Jeong TH, Yun MY. [Antioxidant activity and inhibition of MMP-1 expression of *schizandrae fructus* (*schizandra chinensis*) extract]. Korean J Pharmacogn. 2013;44(1):47-52. Korean.
40. Ko BS, Park SK, Choi SB, Jun DW, Choi MK, Park SM. [A study on hypoglycemic effects of crude extracts of *schizandrae fructus*]. J Korean Soc Appl Biol Chem. 2004;47(2):258-64. Korean.